RESEARCH NOTE

Open Access



Associations of dietary inflammatory index, serum levels of MCP-1 and body composition in Iranian overweight and obese women: a cross-sectional study

Nasim Ghodoosi¹, Atieh Mirzababaei¹, Elahe Rashidbeygi¹, Negin Badrooj¹, Seyedeh Forough Sajjadi¹, Leila Setayesh¹, Mir Saeed Yekaninejad², Seyed Ali Keshavarz³, Farideh Shiraseb¹ and Khadijeh Mirzaei^{1*}

Abstract

Objective: Although, several studies have illustrated that there is a relation between dietary inflammatory index (DII) with obesity-related parameters, and inflammation, their results were controversial. This study aimed to investigate this relationship among Iranian women.

Results: Multivariable linear regression showed that fat mass was 0.14 kg lower in the anti-inflammatory diet group, with respect to the pro-inflammatory group, after adjusting covariates such as age, physical activity, economic and job status ($\beta = -0.142, 95\%$ Cl -4.44, -1.71, P = 0.03). Fat-free mass (FFM) was 1.5 kg more in the anti-inflammatory diet group, compared to the pro-inflammatory diet group, after adjusting for potentials cofounders ($\beta = 1.50, 95\%$ Cl 0, 3.01, p = 0.05). Furthermore, after adjusting for potentials cofounders, it was revealed that the subjects with lower DII had lower monocyte chemoattractant protein-1 (MCP-1) levels in serum ($\beta = -18.81, 95\%$ Cl -35.84, -1.79, p = 0.03). These findings suggest an inverse and significant relationship between DII and FFM and also DII is directly related to Fat mass and the level of MCP-1. This finding can be used for developing interventions that aim to promote healthy eating to prevent inflammation and non-communicable disease development among obese females.

Keywords: Dietary inflammatory index, Overweight, Obesity, Body composition, Body fat mass, Fat-free mass, MCP-1

Introduction

Over the past 3 decades, the prevalence of obesity has nearly doubled worldwide [1]. The mean Body mass index(BMI) for women rose by 0.5 kg/m² per decade from 1980 to 2008 globally [2]. Notably, it has been suggested that more than 21% of women will be obese by 2030 internationally [3]. Obesity results in inflammation

Full list of author information is available at the end of the article



cascade [4] and increased the risk of cardiovascular disease (CVD), hypertension (HTN), and stroke [5-8].

Obesity is contributed to low-grade systemic inflammation however, the exact underlying molecular pathway did not understand completely. It has been proposed, however, that this inflammation is created by pro-inflammatory cytokines produced by infiltrated macrophages in adipose tissue [9–11]. The underlying mechanism can be explained by penetration macrophage into adipose tissue, some chemotactic molecules such as monocyte chemoattractant protein-1 (MCP-1), are secreted by white adipose tissue (WAT) [12]. MCP-1 is known as the main ligand of chemokine receptor-2 (CCR2) [13] That plays an important role in obesity-induced inflammatory

© The Author(s) 2020. This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/

^{*}Correspondence: mina_mirzaei101@yahoo.com

¹ Department of Community Nutrition, School of Nutritional Sciences and Dietetics, Tehran University of Medical Sciences (TUMS), P.O. Box: 14155-6117, Tehran, Iran

responses [14]. MCP-1 levels have been related to many diseases, such as CVD, diabetes [15–17]. Recent studies showed that MCP-1 levels can be decreased by an anti-inflammatory diet, such as a Mediterranean diet[18, 19].

Based on the consumption of 45 specific foods and nutrients, dietary inflammatory index (DII) demonstrates the serum levels of six inflammatory biomarkers [Interleukin1beta (IL-1 β), Interleukin 4 (IL-4), Interleukin 6 (IL-6), Interleukin 10 (IL-10), Tumor necrosis factoralpha (TNF α), and C-reactive protein (CRP)] [20]. Moreover, many studies revealed that healthy and unhealthy dietary index such as a vegetarian diet is associated with anthropometric measures or body composition. [21–24]. To our knowledge, a limited number of studies have investigated the relationship between DII, body composition, and serum MCP-1 nevertheless, this association had not been evaluated in Iran. Consequently, the present study decided to assess the relationships between DII with MCP1 and body composition.

Main text

Methods

Study design and population

A cohort of 280 non-postmenopausal and healthy women aged $18 \le$ years, who referred to health centers in Tehran, Iran, in 2018 was recruited in this cross-sectional study. Multistage cluster random sampling method was used to select certain regions from among all the regions of the city; finally, 20 clusters were chosen to select the obese and overweight participants (BMI: 25–40 kg/m²). The exclusion criteria were as follows: a history of any acute or chronic diseases such as hypertension, CVD, diabetes mellitus, hepatic or renal disease or alcohol consumption, regular usage of medicine other than birth control pills, pregnancy, or lactation. They were excluded if adhered to special dietary patterns or had any significant body weight fluctuations over the past year.

Dietary measurements & DII calculation

Dietary assessment was carried out by a validated and reliable 147-item semi-quantitative food frequency question (FFQ) designed according to the Willett study that administered by a trained nutritionist to assess the average daily intake for last year [25]. The FFQ consisted of a list of foods with standard serving sizes. Participants were asked to report their frequency and the amount of each food item consumed during the previous year on a daily (e.g., bread), weekly (e.g., rice, meat), or monthly (e.g., fish) basis. Household measures were used to convert portion sizes of the consumed foods into grams [26], and an estimated average daily intake of food parameters was calculated from the FFQ using NUTRITIONIST IV software (version 7.0; N-Squared Computing, Salem, OR). FFQ-derived dietary data were used to calculate DII scores for all participants. The dietary data were linked to the regionally representative world database that included food consumption from eleven populations around the world and provided a robust estimate of a mean and standard deviation for each parameter [27]. In order to get z-scores, the "standard global mean" was subtracted from the actual dietary intake amount, and this value was divided by the standard deviation. Subsequently, to minimize the effect of 'right skewing', these z-scores were converted into a percentile - each percentile score was doubled and then subtracted by 1. The centered percentile score for each food parameter for each individual was then multiplied by the respective food parameter effect score, to obtain a food parameterspecific DII score for an individual [27]. Subsequently, all of the food parameter-specific DII scores were summed together to calculate the overall DII score. Higher DII scores indicated a more pro-inflammatory diet; whereas lower values represented more anti-inflammatory diets [27]. A total of 29 food parameters were available from the FFQ, were used to calculate DII (namely: energy, carbohydrate, protein, total fat, monounsaturated fat, polyunsaturated fat, saturated fat, omega-3, omega-6 fatty acids, cholesterol, fiber, thiamin, riboflavin, niacin, vitamin B6, folic acid, vitamin B12, iron, magnesium, selenium, zinc, β carotene, vitamin A, C, D, E and tea, onion, caffeine).

Biochemical assessment

All Biochemical analyses were carried out on venous blood samples that were collected after 12 h fasting, and the serum was centrifuged, liquated, then stored at -80 °C. Serum MCP-1 levels were measured by the enzyme-linked immunosorbent assay (ELISA) method with an appropriate kit (Zell Bio GmbH, ULM, Germany, assay range: 5 ng/L-1500 ng/L, sensitivity: 2.4 ng/L, inter-assay variability: CV<12%, intra-assay variability: CV<10%).

Anthropometric assessment

Bioelectrical impedance analysis (BIA) (InBody 720, Korea) was utilized to calculate body composition measures, including body fat mass and fat-free mass. Anthropometric measures such as body weight, BMI, waist circumferences (WC), and waist-hip ratio (WHR) were measured for all participants. Height was characterized while the subjects were in a standing position without shoes, in contact with the wall with their head, shoulders, heels, and hips, and their height was recorded to the nearest 0.1 cm with Seca 206. All of the measurements were done by expert trained technicians and based on specific guidelines, in order to reduce interpersonal variation.

Physical activity assessment

The required information on physical activities in three levels of mild, moderate, and vigorous for research purpose was achieved by using The International Physical Activity Questionnaire (IPAQ) which can be applied by young and middle-aged adults (18–65years). All these sub-components were summed across MET scores and MET-minutes per week (MET-min/wk) were computed and then the total physical activity from all activity categories was reported [28].

Statistical analyses

DII (dichotomous) was examined across the quantitative characteristics. These were analyzed through an independent sample T-test. Linear regression analyses were conducted to determine the relationship between DII score with fat-free mass and serum MCP-1 levels, adjusted for potential confounding factors. The results are reported as a percentage change (β), with 95% confidence intervals (95% CI). P values of <0.05 were considered to be statistically significant. Statistical analysis was performed using SPSS version 21 (SPSS Inc., Chicago, USA).

Results

Study population characteristics

The DII score in this study ranged from -4.14 (most anti-inflammatory score) to 3.89 (most pro-inflammatory score). DII was categorized based on the median value of

the DII (0.05). A total of 280 females were categorized, based on the DII level, into the anti-inflammatory diet (DII \leq 0.05) and the pro-inflammatory diet (DII > 0.06) groups. Participant characteristics by DII categories are provided in Table 1. There were no significant differences observed in participant characteristics across the two DII groups. Notably, there were significant differences in fat-free mass (*P*=0.021) among DII group members and MCP-1 serum levels and physical activity among the DII categories (*P* < 0.0001).

DII food parameters intakes across the anti-inflammatory and pro-inflammatory diet are shown in Table 2. No significant differences were noted regarding energy intakes between DII groups (Table 2). However, among DII groups, some nutrient intakes, such as total fat (P=0.013), saturated fatty acid (SFA), polyunsaturated fatty acid (PUFA), monounsaturated fatty acid (MUFA), omega-6 fatty acids, and Vitamin E (P<0.0001) was higher with those in the pro-inflammatory group, whereas riboflavin (P=0.021), Vitamin C (P=0.031) and Beta-carotene (P<0.0001) were lower intakes mean in the pro-inflammatory diet group compared to the antiinflammatory diet group (Table 2).

Results obtained from modeling DII as a dichotomous variable in relation to the fat-free mass showed a negative association and positive association between DII adherence and fat-free mass and fat mass respectively, after adjustment for age, physical activity, economic and job status (P = 0.05, P = 0.03) (Table 3). As shown in Table 3, the fat-free mass was 1.5 kg more in the anti-inflammatory diet group, compared to the proinflammatory diet group, with adjusted confounders

Table 1 Pa	rticipant chara	acteristics and b	ody com	position b	y DII Level
------------	-----------------	-------------------	---------	------------	-------------

Continuous variables (mean ± SD)	uous variables (mean±SD) Anti-inflammatory diet Pro-inflammato		ory diet P value	
Age (year)	37.60±7.58	35.03±8.48	0.105	
Weight (kg)	81.09 ± 12.56	80.61 ± 12.09	0.317	
Height (cm)	161.25 ± 6.24	161.33±5.75	0.217	
BMI (kg/m ²)	30.94 ± 3.99	30.52 ± 3.56	0.130	
Waist circumference (cm)	99.25 ± 10.24	98.97±9.89	0.391	
Body fat mass (kg)	33.98 ± 8.55	34.27±8.89	0.858	
Visceral fat area(cm ²)	162.39 ± 95.08	175.145±10.75	0.307	
Fat-free mass (kg)	47.6±20.03	46.5±49.20	0.021	
Physical activity (MET h/day)	1162/45±1322/88	773/24±692/04	< 0.0001	
MCP-1	30.76±42.01	51.72 ± 67.13	< 0.0001	

n = 280

Data are presented as mean \pm standard deviation

Nutrients intake adjusted for energy intake before calculating DII

DII values were categorized according to the median (anti-inflammatory diet: DII \leq 0.05, pro-inflammatory diet: DII > 0.06)

Independent sample t-test was used for comparison of continuous variables between DII categories

Table 2	DII	food p	parameters	intak	e accord	ing to	DII group
---------	-----	--------	------------	-------	----------	--------	-----------

	Anti- inflammatory diet Mean SD		Pro- inflammatory diet Mean SD		P value
Energy intake (kcal)	2588.82	707.15	2658.21	790.42	0.140
Carbohydrate (g)	386.41	39.61	357.23	49.32	0.108
Protein (g)	93.67	14.83	83.12	17.38	0.584
Total fat (g)	87.28	15.56	101.38	22.02	0.015
MUFA (g)	29.01	6.52	33.55	10.42	< 0.0001
PUFA (g)	18.85	5.70	21.23	9.40	< 0.0001
SFA (g)	25.98	6.05	30.13	8.55	< 0.0001
Cholesterol (mg)	256.77	79.80	248.50	94.54	0.113
Fiber (g)	50.09	12.70	40.34	14.21	0.439
Thiamine (mg)	2.13	0.31	2.01	0.39	0.156
Riboflavin (mg)	2.31	0.43	2.06	0.64	0.023
Niacin (mg)	26.49	4.88	23.92	6.85	0.165
Vitamin B6 (mg)	2.38	0.37	1.92	0.38	0.354
Folic acid (µg)	648.31	101.31	562.32	92.69	0.222
Vitamin B12 (µg)	4.88	1.88	4.55	2.33	0.402
Vitamin A (RE)	931.58	369.19	616.03	287.89	0.044
Vitamin C (mg)	257.56	98.48	172.09	106.78	0.024
Vitamin E (mg)	16.17	5.92	18.52	10.85	< 0.0001
Vitamin D (µg)	2.84	1.45	2.45	1.63	0.943
Iron (mg)	19.79	2.44	17.43	2.83	0.431
Beta-carotene (µg)	6910.62	3652.64	3581.06	1912.90	< 0.0001
Selenium (µg)	123.90	28.14	115.70	30.25	0.542
Zinc (mg)	13.71	1.97	12.03	2.26	0.214
Magnesium (mg)	510.30	68.62	404.72	66.85	0.811
Caffeine (g)	205.50	189.42	167.97	96.51	0.058
Omega 3 (g)	1.39	0.56	1.35	0.65	0.513
Omega 6 (g)	15.98	5.46	18.67	9.24	< 0.0001
Onion(g)	33.82	23.44	30.97	21.04	0.498
Tea(g)	963.42	946.41	795.08	489.88	0.082

n = 280

Data are presented as mean $\pm\, {\rm standard}$ deviation

Comparisons of nutrient intake across the groups of the DII were analyzed using independent sample t-test

DII values were categorized according to the median (anti-inflammatory diet: ${\sf DII}\,{\le}\,0.05,$ pro-inflammatory

diet: DII > 0.06)

Nutrients intake adjusted for energy intake

 $(\beta = 1.50, 95\% \text{ CI } 0, 3.01, P = 0.05)$, fat mass was 0.14 kg lower in the anti-inflammatory diet group, with respect to the pro-inflammatory group, with adjusted covariates ($\beta = -0.142, 95\% \text{ CI} - 4.44, -1.71, P = 0.03$). Linear regression tests demonstrated a positive relationship between DII and serum MCP-1 level after adjusting for confounders (P = 0.03). MCP-1 serum levels was 18.81 lower in the anti-inflammatory diet group, with respect to the pro-inflammatory group,

with adjusted covariates ($\beta = -18.81$, 95% CI -35.84, -1.79, P = 0.03) (Table 3).

Discussion

The current study set out to examine the associations between dietary inflammatory potential, body composition, and inflammation in Iranian obese women. Previous studies have shown that different dietary components have varying effects on body composition, and inflammation. In this cross-sectional study, it was found that subjects with a higher DII had lower fat-free mass and higher fat mass than others, independent of potential confounders. Furthermore, independent of confounders, the DII, and MCP-1 are related.

A dissident study in Spain demonstrated that a higher DII score was inversely associated with obesity-related parameters such as FFM and weight [29]. The previous papers have shown DII was related to higher average BMI, WC, and WHR [23, 30, 31]. The associations between fat mass, BMI, and waist circumference with inflammatory molecules such as IL-1, IL-6 have been suggested [32, 33]. In this context, increased circulating cytokine levels, have been described in relation to a decrease in FFM [34]. A possible mechanism was theorized that a diet with an inflammation-induced potential is correlated with higher inflammatory response, and may result in lower fat-free mass. Furthermore, individuals with higher adherence to DII had a lower intake of some nutrients like vitamins C and E, magnesium, potassium, and a range of carotenoids such as Beta carotene that the amount of intake of these nutrients was positively associated with muscle mass in women. Moreover, based on previous evidence, higher adherence to the healthy dietary pattern as the Mediterranean Diet, Healthy Diet Indicator, Diet Quality Index, Alternate Healthy Eating Index, and DASH-style was significantly associated with measurements of muscle mass. Consumption of a variety of plant-based nutrients and overall higher quality in some dietary patterns resulted in the conservation of muscle mass [35]. M. Ruiz-Canela et al. shows participants with higher pro-inflammatory diet also had higher BMI and WHR [36]. Some potential mechanisms are hypothesized, such as activation of pathogen-associated molecular patterns, such as nod and toll-like receptors (TLRs), that induce the activation of inflammatory markers particularly in adipose tissues [37]. Also, intestinal microbiota may be affected by highfat or low-fiber dietary patterns, which also appear to be associated with low-grade inflammation and obesity [38-40]. Evidence suggests that weight gain can be predicted by the proteins in plasma that are sensitive to inflammation [41]. Serine phosphorylation is another conceivable mechanism that assumes a function in inflammation. Inflammation causes phosphorylation of both insulin

Variables	В	95% (CI)	P value
Fat free mass			
Anti-inflammatory diet	1.50	(0.01,3.01)	0.050*
Pro-inflammatory diet	Ref.	Ref.	
Fat mass			
Anti-inflammatory diet	- 0.142	(- 4.44,- 0.171)	0.034*
Pro-inflammatory diet	Ref.	Ref.	
MCP-1			
Anti-inflammatory diet	— 18.81	(- 35.84,- 1.79)	0.030**
Pro-inflammatory diet	Ref.	Ref.	

Table 3 The Association between DII, Fat-Free Mass, Fat Mass, and MCP-1

n = 280

DII values were categorized according to the median

(Anti-inflammatory diet: DII \leq 0.05, Pro-inflammatory diet: DII > 0.06)

P value*: adjusted for age, physical activity, economic and job status

P value **: adjusted for age, weight, smoking, physical activity, economic and job status

receptor and its proteins substrate, including the enzyme phosphatidylinositol 3-OH kinase (PI3 K) bind together, then the inflammatory pathway, mainly I kappa B kinase (IKKb) and Nuclear factor-kappa B (NF-kB) pathway will be activated, finally inhibits insulin-mediated PI3K signaling [42]. IKKb/NF-kB activation also blocked cytokine signaling-3, a protein that blocks cell signals from insulin [43, 44]. Ultimately, these processes trigger suppression of appetite [45].

This study was indicated that participants with higher DII had a greater level of MCP-1. In a study conducted in Belgium was seen that DII and inflammatory markers IL-6 have been associated. Although there were no significant associations between CRP and fibrinogen [46]. Moreover, Shivappa et al. showed that pro-inflammatory diets were related to increased levels of various inflammatory markers such as TNF-a, IL-1, 2, IFN-y, and vascular cell adhesion molecules [47]. The research was conducted in Spain demonstrated that an anti-inflammatory diet, such as a Mediterranean diet, was associated with a significant reduction of IL-6, IL-8, and MCP-1 [48]. Therefore, one possible mechanism seems to suggest that anti-inflammatory diets can modify inflammatory responses, is due to their antioxidants, fiber content, and other anti-inflammatory substances [19, 48].

To the researchers' knowledge, this is the first study conducted which examines the association between dietary inflammatory potential, body composition, and MCP-1 levels.

Conclusion

The present results provide evidence supporting the proposition that a higher DII score (pro-inflammatory diet) is directly associated with lower fat-free mass, higher fat mass, and higher levels of inflammatory markers like MCP-1. These results suggest the importance of promoting dietary patterns with low inflammatory potential to reduce low-grade inflammation and improve obesity- parameters among the overweight and obese population.

Limitations

This study has several limitations. First, include the reliability and validity of the estimation of average food intakes, which were based on the relatively limited number of food items (147 items, which is moderately long for an FFQ). Second, the DII was calculated using the data on just 29 food parameters derived from the FFQ. Third, causality cannot be inferred, because of the crosssectional design of the study. Fourth, it is not possible to generalize about dietary patterns throughout the country, because dietary intakes and other lifestyle measurements in Tehran are somewhat different from those in other parts of the country. Moreover, these dietary patterns were confined to women.

Abbreviations

AHEI: Alternate Healthy Eating Index; BMI: Body mass index; CVD: Cardiovascular disease; CCR2: Chemokine receptor-2; CRP: C-reactive protein; DII: Dietary inflammatory index; DQI: Diet Quality Index; ELISA: Enzyme-linked immunosorbent assay; FFQ: Food frequency questionnaire; FM: Fat mass; FFM: Fat-free mass; HTN: Hypertension; Hs-CRP: High-sensitivity C-reactive protein; HDI: Healthy Diet Indicator; IPAQ: International Physical Activity Questionnaires; IFN-y: Interferon-gamma; IL-2: Interleukin-2; IL-4: Interleukin-4; IL-6: Interleukin-6; IL-8: Interleukin-8; IL-10: Interleukin-10; IL-1β: Interleukin-1β; IKKb: I kappa B kinase; MCP-1: Monocyte Chemoattractant Protein-1; MUFA: Monounsaturated fatty acids; METs: Metabolic Equivalent Tasks; NF-kB: Nuclear factor kappa B; PUFA: Polyunsaturated fatty acids; PI3 K: Phosphatidylinositol 3-OH kinase; SFA: Saturated fatty acids; TNFα: Tumor necrosis factora; TC: Total cholesterol; TG: Triglycerides; WHO: World Health Organization; WHR: Waist to hip ratio; WAT: White adipose tissue.

Acknowledgements

We are grateful to all of the participants for their contribution to this research. This study was supported by grants from the Tehran University of Medical Sciences, Tehran, Iran.

Authors' contributions

NG and ER designed the study and collected the samples, SFS and LS entering FFQ data into NUTRITIONIST IV software, AM and NB wrote the paper, MSY performed the statistical analyses, FSH revised the article, SAK contributed to interpretation of the data and revised the article, KhM had full access to all of the data in the study and took responsibility for the integrity and accuracy of the data. All authors read and approved the final manuscript.

Funding

This study is funded by grants from the Tehran University of Medical Sciences (TUMS), (Grant ID: 95-03-161-33142).

Availability of data and materials

Participants of this study did not agree for their data to be shared publicly, so supporting data is not available.

Ethics approval and consent to participate

Ethics approval for the study protocol was confirmed by The Human Ethics Committee of Tehran University of Medical Sciences (Ethics Number IR.TUMS. VCR.REC.1395.1597). All participants signed a written informed consent that was approved by the Ethics committee.

Consent for publication

Not applicable.

Competing interests

All authors declared that they have no competing interests.

Author details

¹ Department of Community Nutrition, School of Nutritional Sciences and Dietetics, Tehran University of Medical Sciences (TUMS), P.O. Box: 14155-6117, Tehran, Iran. ² Department of Epidemiology and Biostatistics, School of Public Health, Tehran University of Medical Sciences (TUMS), Tehran, Iran. ³ Department of Clinical Nutrition, School of Nutritional Sciences and Dietetics, Tehran University of Medical Sciences, Tehran, Iran.

Received: 12 August 2020 Accepted: 13 November 2020 Published online: 23 November 2020

References

- 1. Gallus S, Lugo A, Murisic B, Bosetti C, Boffetta P, La Vecchia C. Overweight and obesity in 16 European countries. Eur J Nutr. 2015;54(5):679–89.
- Finucane MM, Stevens GA, Cowan MJ, Danaei G, Lin JK, Paciorek CJ, et al. National, regional, and global trends in body-mass index since 1980: systematic analysis of health examination surveys and epidemiological studies with 960 country-years and 9-1 million participants. Lancet. 2011;377(9765):557–67.
- Poston L, Caleyachetty R, Cnattingius S, Corvalán C, Uauy R, Herring S, et al. Preconceptional and maternal obesity: epidemiology and health consequences. Lancet Diabetes Endocrinol. 2016;4(12):1025–36.
- Esser N, Legrand-Poels S, Piette J, Scheen AJ, Paquot N. Inflammation as a link between obesity, metabolic syndrome and type 2 diabetes. Diabetes Res Clin Pract. 2014;105(2):141–50.
- Ahluwalia N, Dalmasso P, Rasmussen M, Lipsky L, Currie C, Haug E, et al. Trends in overweight prevalence among 11-, 13-and 15-year-olds in 25 countries in Europe, Canada and USA from 2002 to 2010. Eur J Public Health. 2015;25(suppl 2):28–32.
- Busetto L, Maggi S. Incidence and prevalence of obesity. Minimally Invasive Bariatric and Metabolic Surgery: Springer; 2015. p. 11–5.
- Fierabracci P, Tamberi A, Santini F. Obesity-related comorbidities. Minimally Invasive Bariatric and Metabolic Surgery: Springer, 2015. p. 25–34.
- Williams EP, Mesidor M, Winters K, Dubbert PM, Wyatt SB. Overweight and obesity: prevalence, consequences, and causes of a growing public health problem. Current obesity reports. 2015;4(3):363–70.

Page 6 of 7

- Calder PC, Ahluwalia N, Brouns F, Buetler T, Clement K, Cunningham K, et al. Dietary factors and low-grade inflammation in relation to overweight and obesity. Br J Nutr. 2011;106(S3):S1–78.
- 10. Sun S, Ji Y, Kersten S, Qi L. Mechanisms of inflammatory responses in obese adipose tissue. Annu Rev Nutr. 2012;32:261.
- Rodríguez-Hernández H, Simental-Mendía LE, Rodríguez-Ramírez G, Reyes-Romero MA. Obesity and inflammation: epidemiology, risk factors, and markers of inflammation. Int J Endocrinol. 2013. https://doi. org/10.1155/2013/678159.
- Fock KM, Khoo J. Diet and exercise in management of obesity and overweight. J Gastroenterol Hepatol. 2013;28(S4):59–63.
- Chen A, Mumick S, Zhang C, Lamb J, Dai H, Weingarth D, et al. Diet induction of monocyte chemoattractant protein-1 and its impact on obesity. Obesity. 2005;13(8):1311–20.
- Bastarrachea RA, Lopez-Alvarenga JC, Bolado-Garcia VE, Tellez-Mendoza J, Laviada-Molina H, Comuzzie AG. Macrophages, inflammation, adipose tissue, obesity and insulin resistance. Gac Med Mex. 2007;143(6):505–12.
- Dawson J, Miltz W, Mir AK, Wiessner C. Targeting monocyte chemoattractant protein-1 signalling in disease. Expert opinion on therapeutic targets. 2003;7(1):35–48.
- Niu J, Kolattukudy PE. Role of MCP-1 in cardiovascular disease: molecular mechanisms and clinical implications. Clin Sci. 2009;117(3):95–109.
- 17. Panee J. Monocyte Chemoattractant Protein 1 (MCP-1) in obesity and diabetes. Cytokine. 2012;60(1):1–12.
- Camargo A, Delgado-Lista J, Garcia-Rios A, Cruz-Teno C, Yubero-Serrano EM, Perez-Martinez P, et al. Expression of proinflammatory, proatherogenic genes is reduced by the Mediterranean diet in elderly people. Br J Nutr. 2012;108(3):500–8.
- Medina-Remón A, Casas R, Tressserra-Rimbau A, Ros E, Martínez-González MA, Fitó M, et al. Polyphenol intake from a Mediterranean diet decreases inflammatory biomarkers related to atherosclerosis: a substudy of the PREDIMED trial. Br J Clin Pharmacol. 2017;83(1):114–28.
- Ricordi C, Garcia-Contreras M, Farnetti S. Diet and inflammation: possible effects on immunity, chronic diseases, and life span. J Am Coll Nutr. 2015;34(sup1):10–3.
- 21. Tonstad S, Butler T, Yan R, Fraser GE. Type of vegetarian diet, body weight, and prevalence of type 2 diabetes. Diabetes Care. 2009;32(5):791–6.
- Azadbakht L, Esmaillzadeh A. Dietary diversity score is related to obesity and abdominal adiposity among Iranian female youth. Public health nutrition. 2011;14(1):62–9.
- Ruiz-Canela M, Zazpe I, Shivappa N, Hebert JR, Sanchez-Tainta A, Corella D, et al. Dietary inflammatory index and anthropometric measures of obesity in a population sample at high cardiovascular risk from the PREDIMED (PREvencion con Dleta MEDiterranea) trial. Br J Nutr. 2015;113(6):984–95.
- 24. Salehi-Abargouei A, Esmaillzadeh A, Azadbakht L, Keshteli AH, Feizi A, Feinle-Bisset C, et al. Nutrient patterns and their relation to general and abdominal obesity in Iranian adults: findings from the SEPAHAN study. Eur J Nutr. 2016;55(2):505–18.
- Mirmiran P, Esfahani FH, Mehrabi Y, Hedayati M, Azizi F. Reliability and relative validity of an FFQ for nutrients in the Tehran lipid and glucose study. Public health nutrition. 2010;13(5):654–62.
- Ghaffarpour M, Houshiar-Rad A, Kianfar H. The manual for household measures, cooking yields factors and edible portion of foods. Tehran Nashre Olume Keshavarzy. 1999;7:213.
- Shivappa N, Steck SE, Hurley TG, Hussey JR, Hébert JR. Designing and developing a literature-derived, population-based dietary inflammatory index. Public health nutrition. 2014;17(8):1689–96.
- Craig CL, Marshall AL, Sjorstrom M, Bauman AE, Booth ML, Ainsworth BE, et al. International physical activity questionnaire: 12-country reliability and validity. Med Sci Sports Exerc. 2003;35(8):1381–95.
- Correa-Rodríguez M, Rueda-Medina B, González-Jiménez E, Correa-Bautista JE, Ramírez-Vélez R, Schmidt-RioValle J. Dietary inflammatory index, bone health and body composition in a population of young adults: a cross-sectional study. Int J Food Sci Nutr. 2018. https://doi. org/10.1080/09637486.2018.1446915.
- Ruiz-Canela M, Zazpe I, Shivappa N, Hebert JR, Sánchez-Tainta A, Corella D, et al. Dietary inflammatory index and anthropometric measures of obesity in a population sample at high cardiovascular risk from the PREDIMED (PREvencion con Dleta MEDiterranea) trial. Br J Nutr. 2015;113(6):984–95.

- Muhammad HFL, van Baak MA, Mariman EC, Sulistyoningrum DC, Huriyati E, Lee YY, et al. Dietary inflammatory index score and its association with body weight, blood pressure, lipid profile, and leptin in Indonesian adults. Nutrients. 2019;11(1):148.
- Brinkley TE, Hsu F-C, Beavers KM, Church TS, Goodpaster BH, Stafford RS, et al. Total and abdominal adiposity are associated with inflammation in older adults using a factor analysis approach. J Gerontol Series A. 2012;67(10):1099–106.
- Cesari M, Kritchevsky SB, Baumgartner RN, Atkinson HH, Penninx BW, Lenchik L, et al. Sarcopenia, obesity, and inflammation—results from the trial of angiotensin converting enzyme inhibition and novel cardiovascular risk factors study. Am J Clin Nutr. 2005;82(2):428–34.
- Londhe P, Guttridge DC. Inflammation induced loss of skeletal muscle. Bone. 2015;80:131–42.
- Kelaiditi E. Diet, inflammation and skeletal muscle mass in women. Norwich: University of East Anglia; 2013.
- Jin C, Flavell RA. Innate sensors of pathogen and stress: linking inflammation to obesity. J Allergy Clin Immunol. 2013;132(2):287–94.
- Salas-Salvado J, Bullo M, Garcia-Lorda P, Figueredo R, Del Castillo D, Bonada A, et al. Subcutaneous adipose tissue cytokine production is not responsible for the restoration of systemic inflammation markers during weight loss. Int J Obes. 2006;30(12):1714–20.
- Wu GD, Chen J, Hoffmann C, Bittinger K, Chen Y-Y, Keilbaugh SA, et al. Linking long-term dietary patterns with gut microbial enterotypes. Science. 2011;334(6052):105–8.
- Martínez JA, Etxeberría U, Galar A, Milagro FI. Role of dietary polyphenols and inflammatory processes on disease progression mediated by the gut microbiota. Rejuvenation Rese. 2013;16(5):435–7.
- 40. Chassaing B, Gewirtz AT. Gut microbiota, low-grade inflammation, and metabolic syndrome. Toxicol Pathol. 2014;42(1):49–53.
- Engström G, Hedblad B, Stavenow L, Lind P, Janzon L, Lindgärde F. Inflammation-sensitive plasma proteins are associated with future weight gain. Diabetes. 2003;52(8):2097–101.

- 42. Kievit P, Howard JK, Badman MK, Balthasar N, Coppari R, Mori H, et al. Enhanced leptin sensitivity and improved glucose homeostasis in mice lacking suppressor of cytokine signaling-3 in POMC-expressing cells. Cell Metab. 2006;4(2):123–32.
- McCormick SM, Gowda N, Fang JX, Heller NM. Suppressor of cytokine signaling (SOCS) 1 regulates interleukin-4 (IL-4)-activated insulin receptor substrate (IRS)-2 tyrosine phosphorylation in monocytes and macrophages via the proteasome. J Biol Chem. 2016;291(39):20574–87.
- 44. Ueki K, Kondo T, Kahn CR. Suppressor of cytokine signaling 1 (SOCS-1) and SOCS-3 cause insulin resistance through inhibition of tyrosine phosphorylation of insulin receptor substrate proteins by discrete mechanisms. Mol Cell Biol. 2005;25(19):8762.
- Mars M, de Graaf C, van Rossum C, de Groot C, Seidell J, Kok F, editors. Leptin and appetite responses induced by a four-day energy restriction; preliminary results. Food Choice Wageningen; 2002. p. 247–247.
- Shivappa N, Hébert JR, Rietzschel ER, De Buyzere ML, Langlois M, Debruyne E, et al. Associations between dietary inflammatory index and inflammatory markers in the Asklepios Study. Br J Nutr. 2015;113(4):665–71.
- Shivappa N, Hebert JR, Marcos A, Diaz LE, Gomez S, Nova E, et al. Association between dietary inflammatory index and inflammatory markers in the HELENA study. Mol Nutr Food Res. 2017;61(6):1600707.
- Casas R, Urpi-Sardà M, Sacanella E, Arranz S, Corella D, Castañer O, et al. Anti-inflammatory effects of the Mediterranean diet in the early and late stages of atheroma plaque development. Mediators inflamma. 2017;2017:3674390.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

