

RESEARCH NOTE

Open Access



First report of *Staphylococcus pseudintermedius* ST71-SCCmec III and ST45- Ψ SCCmec₅₇₃₉₅ from canine pyoderma in Argentina

Mariela E. Srednik^{1*}, Claudia A. Perea¹, Gabriela I. Giacoboni², Jessica A. Hicks¹ and Linda K. Schlater¹

Abstract

Staphylococcus pseudintermedius is an opportunistic pathogen commonly associated with skin infections in dogs. Twenty-three methicillin-resistant *S. pseudintermedius* (MRSP) isolated in Argentina from dogs with pyoderma were analyzed using whole genome sequencing (WGS) and classified into sequence types (ST) by multilocus sequence typing (MLST) and staphylococcal chromosome cassette *mec* (SCC*mec*) types.

Based on the WGS analysis, MLST, and SCC*mec* type results, we report for the first time in Argentina two MRSP strains, one each, belonging to ST71-SCC*mec* III and ST45- Ψ SCC*mec*₅₇₃₉₅ from dogs with pyoderma. We also identified seven isolates with ST339, which had been previously reported in only two isolates in Argentina. Additionally, we identified ten MRSP isolates harboring variants of the SCC*mec* V found in *S. aureus*, seven SCC*mec* V (5C2&5) with two *ccrC1* recombinases, and three SCC*mec* V (5C2) with one *ccrC1* recombinase.

Our findings provide important insights into the evolution and geographic spread of these hypervirulent dominant clones that threaten the health of our companion animals and represent a significant risk for zoonotic infections.

Keywords *Staphylococcus pseudintermedius*, MRSP, SCC*mec*, MLST, ST71, ST45, Canine pyoderma

Introduction

Staphylococcus pseudintermedius is an important opportunistic pathogen in canine companions and is commonly associated with skin infections [1]. This bacterium is sporadically associated with human infections because it can be transmitted easily via close contact with animals, and

it has the potential to cause severe disease [2]. Methicillin-resistant staphylococci of the intermedius group (SIG) emerged in canines in 1999 [3], and *S. pseudintermedius* was first described in 2005 [4]. Methicillin-resistant *S. pseudintermedius* (MRSP) has been spreading worldwide through the expansion and dissemination of dominant clonal lineages with specific genetic characteristics, including the sequence type (ST) 71 in Europe, ST68 in North America and ST45/ST112 in Asia [5, 6]. The first infection of MRSP in humans was reported in 2006 in Belgium [7] and the first MRSP isolated from a human patient in Argentina was reported in 2020 [8]. Furthermore, dominant clones are multi-drug resistant (MDR), suggesting that the spread of horizontally transferrable

*Correspondence:

Mariela E. Srednik
mariela.srednik@gmail.com

¹National Veterinary Services Laboratories, Animal and Plant Health Inspection Service, U.S. Department of Agriculture, Ames, IA, U.S.A.

²Laboratorio de Bacteriología y Antimicrobianos, Departamento de Microbiología, Facultad de Ciencias Veterinarias, Universidad Nacional de La Plata, La Plata, Argentina



© The Author(s) 2023. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

resistance genes is a contributing factor for the dissemination of certain sequence types [9].

Antimicrobial resistance patterns differ in the three most prevalent MRSP clonal lineages [5]. Clonal complexes (CCs) are groups of sequence types (STs) sharing at least six identical alleles of the seven *S. pseudintermedius* MLST genes (*ack*, *cpn60*, *fdh*, *pta*, *purA*, *sar*, and *tuf*), with the primary founder being the ST with the largest number of single locus variants (SLVs) and all other strains diverge from the predicted clonal ancestor [10]. MRSP belonging to clonal complexes CC71 and CC68 often contain several genes that confer resistance to multiple antimicrobials in addition to the *mecA* gene located within the *SCCmec* cassette [11, 12]. For CC45, isolates often harbor resistance genes and mutations that make them resistant to almost all antimicrobials used in veterinary medicine [13].

In 2010, the global population structure of MRSP gradually started to change and it became more heterogeneous than previously described, with evidence of dissemination through clonal expansion of MRSP dominant lineages over large distances [14]. In Europe, there was an apparent decrease of ST71 [6, 15, 16] with the emergence of two novel MRSP lineages (ST258 and ST496) of European and Australian origin [6, 17]. Likewise, ST71 clones began to spread worldwide over more distant locations and this clone has now been reported in Asia and in North and South America, with high prevalence in many countries in these regions. This change in the global population structure of *S. pseudintermedius* may be the consequence of importation from other countries due to the mobilization of animals and people across geographical locations [9, 18, 19]. In other parts of the world, the MRSP population appears to be more diverse. In Argentina, the MRSP population consists of genetically distinct STs not closely related to the more prevalent ST71 and ST68 lineages [20].

Staphylococcal chromosome cassette *mec* (*SCCmec*) typing is one of the molecular techniques currently used to understand the epidemiology and the clonal relationships of methicillin-resistant *S. aureus* (MRSA) strains [21]. Consequently, *SCCmec* typing for *S. pseudintermedius* has been progressively adapted from the work done for *S. aureus*. Existing reports of *S. pseudintermedius* *SCCmec* type III (previously described as II-III by Descloux et al. [22]) associated it with the European epidemic clone ST71, and Ψ *SCCmec*₅₇₃₉₅ was significantly associated with ST45 [5, 11, 13]. To date, no knowledge exists regarding *S. pseudintermedius* belonging to the ST71 and ST45 clones in Argentina. Here we report for the first time in Argentina ST71-*SCCmec* III and ST45- Ψ *SCCmec*₅₇₃₉₅.

Main text

Methods

Isolate selection Thirty *S. pseudintermedius* isolates from dogs with pyoderma collected during 2016 from the Buenos Aires Metropolitan Area (Ciudad Autónoma de Buenos Aires, Gran Buenos Aires and La Plata, Argentina) were selected randomly from the strain collection of the Laboratory of Bacteriology and Antimicrobials, Department of Microbiology, Faculty of Veterinary Sciences, National University of La Plata, Argentina (Laboratorio de Bacteriología y Antimicrobianos, Departamento de Microbiología, Facultad de Ciencias Veterinarias, Universidad Nacional de La Plata, Argentina). Identification was confirmed by MALDI-TOF and whole genome sequencing (WGS) at the National Veterinary Services Laboratories (NVSL) in Ames, Iowa, U.S.A. Twenty-three *S. pseudintermedius* isolates were identified as methicillin-resistant (MRSP) due to the presence of the *mecA* gene, which encodes methicillin resistance, through WGS analysis (described below).

Whole genome sequencing and genomic analysis

Sequencing was performed with the Illumina MiSeq platform using 2×250 paired-end chemistry and the NexteraXT library preparation kit. Multilocus sequence typing (MLST) was determined using ABRicate (<https://github.com/tseeman/abricate/>) with the *S. pseudintermedius* PubMLST database, and new alleles and sequence types (STs) were submitted to PubMLST (<http://pubmlst.org/spseudintermedius>) for curation and number designation by Vincent Perreten (vincent.perreten@vetsuisse.unibe.ch). *SCCmec* types were determined using *SCCmecFinder* 1.2 [23] (<https://cge.food.dtu.dk/services/SCCmecFinder-1.2/>), a database with *SCCmec* types I through XII, including *SCCmec* IV and V subtypes (as of the preparation of this manuscript), based on those identified in *S. aureus*. For the predicted *SCCmec* types III and V, additional manual alignment/mapping was performed using the available reference sequences for these *SCCmec* types for *S. aureus* and *S. pseudintermedius* (AB03671.1, AM904732.1 for *SCCmec* type III; HE984157.2 for Ψ *SCCmec*₅₇₃₉₅; and FJ544922.1, ERR175868, AB512767.1, AB505629.1, AB462393.1, AB121219.1 for *SCCmec* type V), using Geneious Prime v11.0.9 (Biomatters Ltd., NZ).

Results

For the 23 MRSP isolates analyzed, a total of 14 sequence types (STs) were identified, five previously described: ST339 (n=7), ST1412 (n=3), ST71 (n=2), ST45 (n=1) and ST313 (n=1); and nine newly identified STs (Table 1).

SCCmecFinder successfully classified twelve isolates as *SCCmec* type IIIa (n=2), *SCCmec* type V (5C2) (n=3) and *SCCmec* type V (5C2&5) (n=7). The remainder of the isolates could not be typed.

Table 1 Multilocus sequence types (MLST) and SCCmec types of methicillin resistant *Staphylococcus pseudintermedius* isolates obtained from dogs with pyoderma in Argentina

MLST	SCCmec type
ST339 (n=7)	SCCmec V (5C2) (in only 2 isolates)
ST1412 (n=3)	SCCmec V (5C2&5)
ST71 (n=2)	SCCmec III (previously described as II-III)
ST45 (n=1)	ΨSCCmec ₅₇₃₉₅
ST313 (n=1)	none
<i>Newly identified</i>	
ST2233 (n=1), ST2234 (n=1), ST2235 (n=1), ST2242 (n=1)	SCCmec V (5C2&5)
ST2261 (n=1)	SCCmec V (5C2)
ST2243 (n=1), ST2244 (n=1), ST2236 (n=1), ST2237 (n=1)	none

The two isolates classified as SCCmec type III belonged to ST71. These were mapped against the *S. pseudintermedius* KM1381 (AM904732.1) genome reference that harbors a hybrid SCCmec type II-III, described to be a combination of SCCmec II from *S. epidermidis* and SCCmec III from *S. aureus*, but lacking the cadmium resistance operon [22]. Both isolates showed high homology (99.9%) to this reference (Fig. 1A).

For one isolate identified as ST45, a SCCmec type could not be determined using SCCmecFinder, but alignment/mapping to HE984157.2 resulted in high homology (98.8%) classifying it as ΨSCCmec₅₇₃₉₅ (Fig. 1B).

Of the SCCmec type V, three were predicted as SCCmec type V (5C2), with only one *ccrC1* recombinase, and seven were predicted as SCCmec type V (5C2&5), with two *ccrC1* recombinases. When these 10 isolates were compared against SCCmec V subtype references (Va, Vb and Vc), isolates with SCCmec type V (5C2) (BI-1991, BI-2002, BI-2008) showed 79.4–90.6% homology to the *S. aureus* type Va (5C2) reference strain. The rest showed 84.8–99.8% homology to *S. pseudintermedius* 06-3228 (FJ544922.1) and *S. pseudintermedius* 23,929 (ERR175868), which are both references for *S. pseudintermedius* SCCmec V (5C2&5) [12, 24]. We classified five of these isolates as SCCmec Vb due to their homology with *S. aureus* AB462393.1 (Vb). Furthermore, two of these SCCmec Vb (BI-1980, BI-1990) showed evidence of harboring a truncated *mecR1* gene. Finally, we classified two isolates (BI-1991, BI-2003) as SCCmec Vc (5C2&5) because they harbored the *czrC* gene that is present in the SCCmec Vc but is absent in Vb. (Figure 1C, D and E).

Discussion

This study is the first report of *S. pseudintermedius* ST71-SCCmec III and ST45-ΨSCCmec₅₇₃₉₅ in Argentina, obtained from a cohort of isolates recovered from dogs with pyoderma in the Buenos Aires Metropolitan

Area in 2016. A previous study in Argentina described a population of MRSP from dogs with clinical disease that consisted of six genetically distinct STs: ST339, ST649, ST919, ST920, ST921, and ST922 [20]. Here, among 23 MRSP, ST339 (n=7) was also identified, as well as an additional thirteen sequence types, including ST1412 (n=3), ST71 (n=2), ST45 (n=1), ST313 (n=1) and nine newly identified STs (ST2233-2237, ST2242-2244 and ST2261). These data contribute to the characterization of the population structure of MRSP in Argentina, which now includes two globally prevalent clones (ST71 and ST45). ST71 was initially described as the predominant clone in Europe but is now spread worldwide, whereas ST45 was described as the most prevalent clone in Asia [5]. Ggetti et al. [20] identified two isolates with sequence type ST339 in Argentina. The first MRSP recovered from a human patient in Argentina was ST1412 [8]. Interestingly ST1412 is a double locus variant of ST45, the sequence type that originated in Asia.

The ST71 clone has mainly been associated to SCCmec type III [11]. This SCCmec, first identified in 2005, was initially classified as a hybrid SCCmec II-III [22]. The distribution of this clone was primary found in Europe, but is now disseminated worldwide [23, 25]. The first report of an ST71 MRSP in South America was from a dog in Brazil in 2013 [26] and this study is the first report of this clone in Argentina. As in previous reports, the two isolates identified in this study as ST71 harbored SCCmec type III.

Pseudo (Ψ) SCCmec elements have been identified in *S. haemolyticus* with no evidence of *ccr* genes, but with a *mec* complex [27, 28]. A novel ΨSCCmec₅₇₃₉₅ was described in MRSP CC45 from companion animals in Thailand and Israel [13]. In Australia, MRSP belonging to ST45 was also associated to this novel ΨSCCmec₅₇₃₉₅ element [18]. Even though no particular SCCmec type is usually associated to MRSP-ST45 [25, 29], some reports identified ΨSCCmec₅₇₃₉₅ with this clone [13, 18]. The results from this study show evidence to also classify the MRSP-ST45 isolate from Argentina as an ST45-ΨSCCmec₅₇₃₉₅, making this the first report of this element in the country.

Lastly, almost half (10/23) of the isolates were predicted as SCCmec V. SCCmec V has been associated to different STs [5], and variation has been observed in SCCmec type V for *S. pseudintermedius* in comparison to *S. aureus*. Currently, this element is classified into three subtypes for *S. aureus*, according to Uehara [30]: Va (5C2), Vb (5C2&5) and Vc (5C2&5). To provide clarity, it's important to mention how the classification for subtype Vb has evolved. Initially, it was classified as V_T (AB462393.1) [31]. Later, Black et al. [12] described a homologous SCCmec type V element in *S. pseudintermedius* (FJ44922.1), which only differed in a deleted section of a gene in *S.*

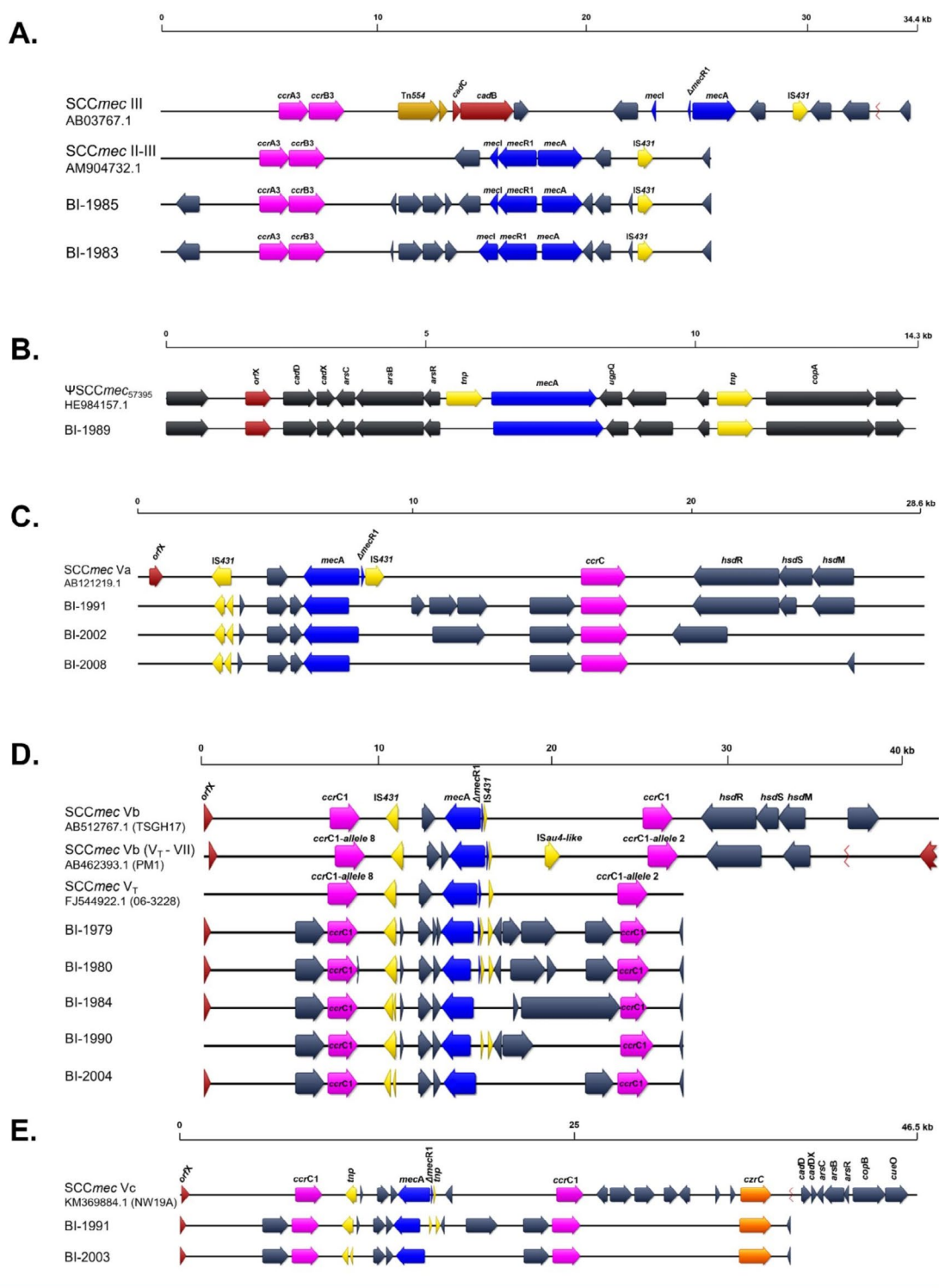


Fig. 1 **A**, Alignment of *S. pseudintermedius* BI-1983 and BI-1985 SCCmec elements to *S. pseudintermedius* KM1381 (AM904732, first described as hybrid II-III) and *S. aureus* 85/2082 (AB037671.1, SCCmec III). **B**, Alignment of BI-1989 to *S. pseudintermedius* 57,395 (HE984157.2, Ψ SCCmec₅₇₃₉₅). **C**, Alignment of SCCmec V (5C2) predicted elements for BI-1991, BI-2002 and BI-2008 to *S. aureus* SCCmec Va (5C2) [AB121219.1]. **D**, Alignment of SCCmec Vb (5C2&5) predicted elements for BI-1979, BI-1980, BI-1984, BI-1990, and BI-2004 to *S. aureus* Vb (5C2&5) [AB462393.1; AB512767.1] and *S. pseudintermedius* SCCmec V_T (FJ544922.1). **E**, Alignment of SCCmec Vc (5C2&5) predicted elements BI-1981 and BI-2003 to *S. aureus* Vc (5C2&5) [KM369884].

pseudintermedius with respect to *S. aureus*. Then, Takano et al. [32] proposed reclassification of Vb to as SCCmec type VII. Finally, Perreten et al. [11], described an SCCmec V in *S. pseudintermedius* that was highly homologous to the previously named V_T or VII from *S. aureus*, which was designated as SCCmec V (5C2&5). In this study, three MRSP isolates, belonging to ST339, showed one *ccrC1* recombinase only and were most homologous to SCCmec V (5C2). In contrast, the remaining seven MRSP isolates showed two *ccrC1* recombinases and were most homologous to SCCmec V (5C2&5). Additionally, there was evidence to suggest that the *mecR1* gene was truncated in two of these isolates (BI-1979 and BI-1980). Worthing et al. [18] reported similar results for the SCCmec V_T identified in their study. Prior to our study, SCCmec V (5C2&5) was the only SCCmec type reported in MRSP in Argentina [20].

Conclusion

Using whole-genome sequencing we identified two MRSP isolates, one belonging to sequence type 71 and carrying staphylococcal cassette chromosome *mec* type III (ST71-SCCmec III), and the other belonging to sequence type 45 and carrying the Ψ SCCmec₅₇₃₉₅ (ST45- Ψ SCCmec₅₇₃₉₅), neither of which had been previously reported in Argentina. Even though these sequence types were first identified and distributed in Europe and Asia, respectively, our results support the current worldwide spread observed for these *S. pseudintermedius* clones. These findings highlight the importance of WGS for understanding the circulating populations of MRSP and the spread of multidrug-resistant *S. pseudintermedius* in companion animals, which can consequently have a significant impact on public health.

Limitation

- Complete fragment coverage of the SCCmec elements was limited due to the inevitable gaps present in assemblies from short read technology, therefore fully closed genomes were not available.
- There are inconsistencies in the literature regarding nomenclature and classification of SCCmec elements, which makes interpretation and comparative analysis more complex.
- There is an evident need for a formal SCCmec nomenclature that includes SCCmec elements from *Staphylococcus pseudintermedius* and other *Staphylococcus* species.

Abbreviations

CC	Clonal complex
MDR	Multi-drug resistant
MLST	Multilocus sequence typing
MRSP	Methicillin-resistant <i>Staphylococcus pseudintermedius</i>
SCCmec	Staphylococcal cassette chromosome <i>mec</i>
SIG	Staphylococci of the <i>intermedius</i> group

ST	Sequence type
WGS	Whole genome sequencing

Acknowledgements

Not applicable.

Author Contributions

M.S. and C.P. analyzed and interpreted the data. M.S. was involved in the study design, bacterial identification confirmation, whole genome sequencing, and writing of the draft manuscript. C.P. performed the figures and corrected the manuscript. G.G. carried out the bacterial isolation and biochemical identification. J.H. carried out the bioinformatic analysis. L.S. provided project oversight, obtained resources, and revised the manuscript. All authors read and approved the final manuscript.

Funding

This project was supported in part by an appointment to the Research Participation Program at the Animal and Plant Health Inspection Service, United States Department of Agriculture, administered by the Oak Ridge Institute for Science and Education through an interagency agreement between the U. S. Department of Energy and USDA APHIS.

Data Availability

All sequence data was deposited in NCBI under BioProject PRJNA848756.

Declarations

Ethics approval and consent to participate

Ethical approval and guidelines are not applicable to the study as no animals were involved in the research. Consent was obtained from all owners for the use of the samples for research purposes.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Received: 13 July 2022 / Accepted: 3 February 2023

Published online: 23 February 2023

References

1. Bannoehr J, Guardabassi L. *Staphylococcus pseudintermedius* in the dog: taxonomy, diagnostics, ecology, epidemiology and pathogenicity. *Vet Dermatol* 2012;23(4):253–66, e51–2. doi: <https://doi.org/10.1111/j.1365-3164.2012.01046.x>. PMID: 22515504.
2. Somayaji R, Rubin JE, Priyantha MA, Church D. Exploring *Staphylococcus pseudintermedius*: an emerging zoonotic pathogen? *Future Microbiol*. 2016;11:1371–4. <https://doi.org/10.2217/fmb-2016-0137>.
3. Gortel K, Campbell KL, Kakoma I, Whittam T, Schaeffer DJ, Weisiger RM. Methicillin resistance among staphylococci isolated from dogs. *Am J Vet Res*. 1999;60:1526–30.
4. Devriese LA, Vancanneyt M, Baele M, Vanechoutte M, De Graef E, Snaeuwaert C, Cleenwerck I, Dawyndt P, Swings J, Decostere A, Haesebrouck F. *Staphylococcus pseudintermedius* sp. nov., a coagulase-positive species from animals. *Int. J. Syst. Evol. Microbiol* 2005;55(Pt 4):1569–1573. doi: <https://doi.org/10.1099/ijs.0.63413-0>. PMID: 16014483.
5. Pires dos Santos T, Damborg P, Moodley A, Guardabassi L. Systematic review on global epidemiology of methicillin-resistant *Staphylococcus pseudintermedius*: inference of population structure from multilocus sequence typing data. *Front Microbiol*. 2016;7:1599. <https://doi.org/10.3389/fmicb.2016.01599>.
6. Bergot M, Martins-Simoes P, Kilian H, Châtre P, Worthing KA, Norris JM, Madec JY, Laurent F, Haenni M. Evolution of the Population structure of *Staphylococcus pseudintermedius* in France. *Front Microbiol*. 2018;9:3055.
7. Van Hoovels L, Vankeerberghen A, Boel A, Van Vaerenbergh K, De Beenhouwer H. First case of *Staphylococcus pseudintermedius* infection in a human. *J Clin Microbiol*. 2006 Dec;44(12):4609–12. <https://doi.org/10.1128/JCM.01308-06>. Epub 2006 Oct 18. PMID: 17050817; PMCID: PMC1698428.

8. Gagetti P, Errecalde L, Wattam AR, De Belder D, Ojeda Saavedra M, Corso A, Rosato AE. Characterization of the first *mecA*-positive Multidrug-Resistant *Staphylococcus pseudintermedius* isolated from an argentinian patient. *Microb Drug Resist*. 2020;26(7):717–21. <https://doi.org/10.1089/mdr.2019.0308>. PMID: 32031908; PMCID: PMC7368382.
9. Bruce S, Smith JT, Mydosh JL, Ball J, Needle DB, Gibson R, Andam CP. Accessory Genome Dynamics of Local and Global *Staphylococcus pseudintermedius* populations. *Front Microbiol*. 2022;13:798175. <https://doi.org/10.3389/fmicb.2022.798175>. PMID: 35222331; PMCID: PMC8867027.
10. Feil EJ, Li BC, Aanensen DM, Hanage WP, Spratt BG. eBURST: inferring patterns of evolutionary descent among clusters of related bacterial genotypes from multilocus sequence typing data. *J Bacteriol*. 2004 Mar;186(5):1518–30. PMID: 14973027; PMCID: PMC344416.
11. Perreten V, Kadlec K, Schwarz S, Groenlund Andersson U, Finn M, Greko C, et al. Clonal spread of methicillin-resistant *Staphylococcus pseudintermedius* in Europe and North America: an international multicentre study. *J Antimicrob Chemother*. 2010;65:1145–54. <https://doi.org/10.1093/jac/dkq078>.
12. Black CC, Solyman SM, Eberlein LC, Bemis DA, Woron AM, Kania SA. Identification of a predominant multilocus sequence type, pulsed-field gel electrophoresis cluster, and novel staphylococcal chromosomal cassette in clinical isolates of *mecA*-containing, methicillin-resistant *Staphylococcus pseudintermedius*. *Vet. Microbiol* 2009;139(3–4):333–8. doi: <https://doi.org/10.1016/j.vetmic.2009.06.029>. PMID: 19604657.
13. Perreten V, Chanchaithong P, Prapasarakul N, Rossano A, Blum SE, Elad D, Schwendener S. Novel pseudo-staphylococcal cassette chromosome *mec* element (ψ SCC*mec*₅₇₃₉₅) in methicillin-resistant *Staphylococcus pseudintermedius* CC45. *Antimicrob Agents Chemother*. 2013;57(11):5509–15. <https://doi.org/10.1128/AAC.00738-13>. PMID: 23979735; PMCID: PMC3811289.
14. Smith JT, Amador S, McGonagle CJ, Needle D, Gibson R, Andam C. Population genomics of *Staphylococcus pseudintermedius* in companion animals in the United States. *Commun Biology*. 2020. <https://doi.org/10.1038/s42003-020-1009-y>.
15. Grönthal T, Eklund M, Thomson K, Piiparinen H, Sironen T, Rantala M. Antimicrobial resistance in *Staphylococcus pseudintermedius* and the molecular epidemiology of methicillin-resistant *S. pseudintermedius* in small animals in Finland. *J Antimicrob Chemother*. 2017;72(4):1021–30.
16. Silva V, Oliveira A, Manageiro V, et al. Clonal diversity and Antimicrobial Resistance of Methicillin-Resistant *Staphylococcus pseudintermedius* isolated from Canine Pyoderma. *Microorganisms*. 2021;9(3):482. <https://doi.org/10.3390/microorganisms9030482>.
17. Duim B, Verstappen KM, Broens EM, Laarhoven LM, Van Duijkeren E, Hordijk J, et al. Changes in the population of methicillin-resistant *Staphylococcus pseudintermedius* and dissemination of antimicrobial-resistant phenotypes in the Netherlands. *J Clin Microbiol*. 2016;54:283–8. <https://doi.org/10.1128/JCM.01288-15>.
18. Worthing KA, Schwendener S, Perreten V, Saputra S, Coombs GW, Pang S, Davies MR, Abraham S, Trott DJ, Norris JM. Characterization of Staphylococcal Cassette chromosome *mec* elements from Methicillin-Resistant *Staphylococcus pseudintermedius* Infections in australian animals. *mSphere*. 2018;3(6):e00491–18. <https://doi.org/10.1128/mSphere.00491-18>. PMID: 30404937; PMCID: PMC6222048.
19. Nisa S, Bercker C, Midwinter AC, Bruce I, Graham CF, Venter P, Bell A, French NP, Benschop J, Bailey KM, Wilkinson DA. Combining MALDI-TOF and genomics in the study of methicillin resistant and multidrug resistant *Staphylococcus pseudintermedius* in New Zealand. *Sci Rep*. 2019;9(1):1271. <https://doi.org/10.1038/s41598-018-37503-9>. PMID: 30718644; PMCID: PMC6361924.
20. Gagetti P, Wattam AR, Giacoboni G, De Paulis A, Bertona E, Corso A, Rosato AE. Identification and molecular epidemiology of methicillin resistant *Staphylococcus pseudintermedius* strains isolated from canine clinical samples in Argentina. *BMC Vet Res*. 2019;15:264.
21. Ito T, Katayama Y, Hiramoto K. Cloning and nucleotide sequence determination of the entire *mec* DNA of pre-methicillin-resistant *Staphylococcus aureus* N315. *Antimicrob Agents Chemother*. 1999;43:1449–58.
22. Descloux S, Rossano A, Perreten V. Characterization of new staphylococcal cassette chromosome *mec* (SCC*mec*) and topoisomerase genes in fluoroquinolone- and methicillin-resistant *Staphylococcus pseudintermedius*. *J Clin Microbiol*. 2008;46:1818–23. <https://doi.org/10.1128/JCM.02255-07>.
23. Kaya H, Hasman H, Larsen J, Stegger M, Johannesen TB, Allesøe RL, Lemvig CK, Aarestrup FM, Lund O, Larsen AR. SCC*mec*Finder, a Web-Based Tool for Typing of Staphylococcal Cassette Chromosome *mec* in *Staphylococcus aureus* Using Whole-Genome Sequence Data. *mSphere*. 2018;3(1). <https://doi.org/10.1128/mSphere.00612-17>.
24. McCarthy AJ, Harrison EM, Stanczak-Mrozek K, Leggett B, Waller A, Holmes MA, Lloyd DH, Lindsay JA, Loeffler A. Genomic insights into the rapid emergence and evolution of MDR in *Staphylococcus pseudintermedius*. *J Antimicrob Chemother*. 2015;70(4):997–1007. doi: <https://doi.org/10.1093/jac/dku496>. PMID: 25527273.
25. Couto N, Monchique C, Belas A, Marques C, Gama LT, Pomba C. Trends and molecular mechanisms of antimicrobial resistance in clinical staphylococci isolated from companion animals over a 16 year period. *J Antimicrob Chemother*. 2016;71(6):1479–87. doi: <https://doi.org/10.1093/jac/dkw029>. PMID: 26944924.
26. Quitoco IM, Ramundo MS, Silva-Carvalho MC, Souza RR, Beltrame CO, de Oliveira TF, Araújo R, Del Peloso PF, Coelho LR, Figueiredo AM. First report in South America of companion animal colonization by the USA1100 clone of community-acquired methicillin-resistant *Staphylococcus aureus* (ST30) and by the european clone of methicillin-resistant *Staphylococcus pseudintermedius* (ST71). *BMC Res Notes*. 2013;6:336. <https://doi.org/10.1186/1756-0500-6-336>. PMID: 23981343; PMCID: PMC3765899.
27. Bouchami O, Ben Hassen A, de Lencastre H, Miragaia M. High prevalence of *mec* complex C and *ccrC* is independent of SCC*mec* type V in *Staphylococcus haemolyticus*. *Eur J Clin Microbiol Infect Dis*. 2012;31:605–14.
28. Zong Z. Characterization of a complex context containing *mecA* but lacking genes encoding cassette chromosome recombinases in *Staphylococcus haemolyticus*. *BMC Microbiol*. 2013;13:64. <https://doi.org/10.1186/1471-2180-13-64>.
29. Wegener A, Damborg P, Guardabassi L, Moodley A, Mughini-Gras L, Duim B, Wagenaar JA, Broens EM. Specific staphylococcal cassette chromosome *mec* (SCC*mec*) types and clonal complexes are associated with low-level amoxicillin/clavulanic acid and cefalotin resistance in methicillin-resistant *Staphylococcus pseudintermedius*. *J Antimicrob Chemother*. 2020;75(3):508–11. <https://doi.org/10.1093/jac/dkz509>.
30. Uehara Y. Current status of Staphylococcal Cassette chromosome *mec* (SCC*mec*). *Antibiot (Basel)*. 2022;11(1):86. <https://doi.org/10.3390/antibiotics11010086>. PMID: 35052963; PMCID: PMC8772726.
31. Boyle-Vavra S, Ereshesky B, Wang CC, Daum RS. Successful multiresistant community-associated methicillin-resistant *Staphylococcus aureus* lineage from Taipei, Taiwan, that carries either the novel Staphylococcal chromosome cassette *mec* (SCC*mec*) type VT or SCC*mec* type IV [published correction appears in *J Clin Microbiol* 2005;43(12):6223]. *J. Clin. Microbiol*. 2005;43(9):4719–4730. doi:<https://doi.org/10.1128/JCM.43.9.4719-4730.2005>.
32. Takano T, Higuchi W, Zaraket H et al. Novel characteristics of community-acquired methicillin-resistant *Staphylococcus aureus* strains belonging to multilocus sequence type 59 in Taiwan [published correction appears in *Antimicrob Agents Chemother* 2012;56(12):6441. Zaraket, Hassan [added]. *Antimicrob Agents Chemother* 2008;52(3):837–845. doi:<https://doi.org/10.1128/AAC.01001-07>.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.