

SHORT REPORT

Open Access

# Prostaglandin E<sub>2</sub> regulates the expression of connective tissue growth factor (CTGF/CCN2) in human osteoarthritic chondrocytes via the EP4 receptor

Kayo Masuko<sup>1\*</sup>, Minako Murata<sup>2</sup>, Kazuo Yudoh<sup>2</sup>, Hiroyuki Shimizu<sup>3</sup>, Moroe Beppu<sup>3</sup>, Hiroshi Nakamura<sup>4</sup>, Tomohiro Kato<sup>1</sup>

## Abstract

**Background:** The regulatory mechanisms of the expression of connective tissue growth factor/CCN family member 2 (CTGF/CCN2) in human articular chondrocytes have not been clarified. We investigated the effect of prostaglandin E<sub>2</sub> (PGE<sub>2</sub>) on CTGF/CCN2 expression in chondrocytes.

**Findings:** Articular cartilage samples were obtained from patients with osteoarthritis (OA) and chondrocytes were isolated and cultured in vitro. Chondrocytes were stimulated with PGE<sub>2</sub>, PGE receptor (EP)-specific agonists, or interleukin (IL)-1. CTGF expression was analyzed using quantitative polymerase chain reaction, Western blot, and enzyme-linked immunosorbent assay. The inhibitory effects of EP receptor antagonists (for EP2 and EP4) against PGE<sub>2</sub> stimulation were also investigated. Stimulation of chondrocytes with PGE<sub>2</sub> or IL-1 significantly suppressed CTGF expression. The suppressive effect of PGE<sub>2</sub> was reproduced by EP2/EP4 receptor agonists but not by EP1/EP3 receptor agonists, and was partially blocked by an EP4 receptor antagonist, suggesting that the EP4 receptor has a dominant role.

**Conclusions:** PGE<sub>2</sub> may be involved in the regulation of CTGF/CCN2 expression in human articular chondrocytes via the EP4 receptor. Elucidation of EP4-mediated signaling in chondrocytes may contribute to a better understanding of the effects of PGE<sub>2</sub> in arthritis.

## Background

Connective tissue growth factor/CCN family member 2 (CTGF/CCN2) is a member of the CCN family, which is a group of secreted multifunctional proteins that contain high levels of cysteine (reviewed in [1]). During skeletal development, CTGF is strongly expressed in the mesenchyme, including in hypertrophic chondrocytes, and plays an essential role in endochondral ossification by promoting angiogenesis, proliferation, and differentiation of chondrocytes [2]. In adult tissue, CTGF is expressed during wound healing and in fibrotic tissue [2,3]. Transforming growth factor (TGF)- $\beta$  stimulates CTGF expression, and TGF- $\beta$ -induced CTGF is involved

in scarring and fibrogenesis [3]. In chondrocytes, CTGF contributes to the production of the extracellular matrix by stimulating chondrocyte proliferation, the expression of type II collagen, and aggrecan among other factors, and the activation of integrin signaling [4,5]. CTGF expression is detected in normal human cartilage and in cartilage from patients with osteoarthritis (OA), suggesting that CTGF expression is involved in the development of fibrous tissue over the damaged OA cartilage [6,7].

Despite its potential involvement in OA, the regulatory mechanisms of CTGF expression in chondrocytes have not been fully clarified. Studies using chondrocytic cells and other cell types indicate that CTGF expression is modulated by several factors, including TGF- $\beta$  [8], dexamethasone [9], and macrophage colony stimulating factor [10]. A regulatory role of prostaglandin E<sub>2</sub> (PGE<sub>2</sub>)

\* Correspondence: k\_msk@mac.com

<sup>1</sup>Department of Biochemistry, St. Marianna University School of Medicine, 2-16-1 Sugao, Miyamae-ku, Kawasaki-shi, Kanagawa 216-8511, Japan

in CTGF expression has also been postulated [11,12]. PGE<sub>2</sub> is considered an important catabolic factor in OA cartilage [13]; therefore, the present study aimed to evaluate the potential contribution of PGE<sub>2</sub> in the regulation of CTGF expression in OA chondrocytes.

## Methods

### Samples

Human articular chondrocytes were obtained from 17 patients (M/F = 2:15, mean age 74.2 ± 5.38) with OA who underwent arthroplasty of a knee (15) or hip (2) joint at St. Marianna University School of Medicine Hospital. The diagnoses of OA was made according to the criteria of the American College of Rheumatology [14]. Written informed consent was obtained from each patient, and the study protocol was approved by the institution's ethics committee. The study was performed in compliance with the tenets of the Declaration of Helsinki proposed by the World Medical Association in 1964.

### Cell culture

After careful removal of the synovial tissue, the cartilage was minced, washed, and treated with collagenase. Isolated chondrocytes were then washed and cultured in vitro as a monolayer in Dulbecco's modified Eagle's medium supplemented with 10% fetal bovine serum and antibiotics. The fetal bovine serum used in the study was inactivated by incubation at 56°C for 30 min. The attached cells (P0) were grown on type I collagen-coated culture dishes, and subconfluent cells (P1 cells) were used in the experiments. The differentiated phenotypes of the cells used in the experiments were confirmed through macroscopic observation and on the basis of the expression of type II collagen and aggrecan mRNA by reverse transcription-polymerase chain reaction (PCR; data not shown).

Chondrocytes were serum-starved in medium with 0.5% fetal bovine serum for 24 h prior to the experiments and were either stimulated or not stimulated with PGE<sub>2</sub> (10 nM; Cayman Chemical Co., Ann Arbor, MI), butaprost (10 μM, Sigma-Aldrich, St. Louis, MO), PGE<sub>1</sub> alcohol (10 nM), sulprostone (1 μM, Sigma-Aldrich), or IL-1β (10 ng/ml, R&D Systems, Minneapolis, MN), for the indicated periods. In a separate experiment, cells were pretreated with either AH6809 (10 ng/ml) or GW627368X (5 μM; Cayman Chemical Co., Ann Arbor, MI) for 1 h before stimulation with PGE<sub>2</sub>. Cell viability was not affected by up to 500 nM PGE<sub>2</sub>, the vehicle, or any of the inhibitors during the culture period, as confirmed by trypan blue exclusion and an MTS [3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium] assay (data not shown). Stimulated chondrocytes and culture supernatants were collected and subjected to the following analyses.

### Real-time PCR

Messenger RNA was extracted from the cultured cells and converted to cDNA. Quantitative PCR was performed using specific primers and an ABP Prism 7000 according to the manufacturer's protocol. The sequences for the CTGF primer (GenBank No. NM\_001901) were as follows: forward, 5'-CCTGTGCAGCATGGACGTT-3'; reverse, 5'-GGACCAGGCAGTTGGCTCTAA-3', and the sequences for the GAPDH primer were as published elsewhere [15]. The oligonucleotides were synthesized by Takara Bio Co. Ltd. (Japan)

### Western blotting

Whole cell lysates were extracted from the cultured cells using standard lysis buffer (20 mM Tris-HCl, 250 mM NaCl, 1% NP-40, 1 mM dithiothreitol, 10 mM NaF, 2 mM Na<sub>3</sub>VO<sub>4</sub>, 10 mM Na<sub>4</sub>P<sub>2</sub>O<sub>7</sub>), and Protease Inhibitor Cocktail (Roche, Mannheim, Germany) and stored at -30°C until use. Protein concentration was determined using the Bradford method (Bio-Rad Protein Assay Reagent; BioRad Laboratories, Hercules, CA). The lysates were mixed with the loading dye and subjected to 15% sodium dodecyl sulphate (SDS)-polyacrylamide gel electrophoresis (PAGE). After transfer to a polyvinylidene difluoride membrane, a primary antibody was added: goat anti-CTGF polyclonal antibody (Catalog No. sc-14939, Santa Cruz Biotechnology Inc. Santa Cruz, CA, USA) or anti-glyceraldehyde-3-phosphate dehydrogenase (GAPDH; Abcam Ltd., Cambridge, UK). The membrane was then washed and reacted with the corresponding secondary antibody conjugated with horseradish peroxidase. Finally, the signals were visualized using the extended cavity laser system (GE Healthcare Biosciences KK, Tokyo, Japan). Densitometry of the signal bands was analysed using ImageJ software <http://rsb.info.nih.gov/ij/>. Statistical analyses were performed using Prism™ software (GraphPad Software Inc., San Diego, CA). The results are shown as mean ± SD. Student's t test was used to compare between two groups and a *p* value of less than 0.05 was considered significant.

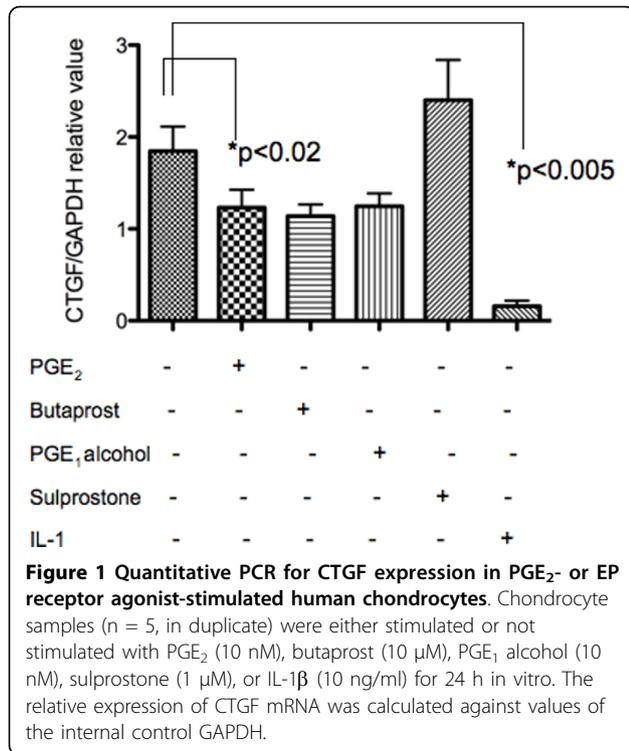
### Enzyme-linked immunosorbent assay (ELISA)

CTGF levels in the culture supernatant were measured in duplicate using the Human CTGF ELISA Kit (Pepro- tech, Rocky Hill, NJ) according to the manufacturer's instructions. According to the manufacturer, the detection range of CTGF using this kit is 63 to 4000 pg/ml. The results were analyzed using Student's t-test.

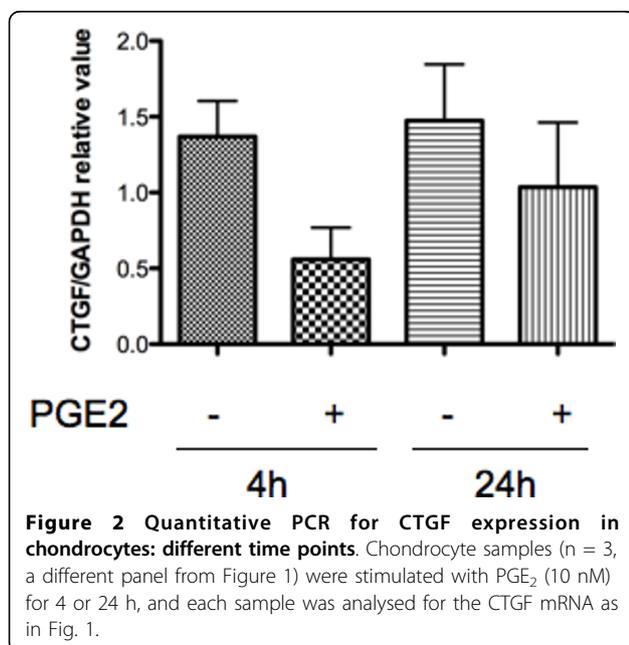
## Results

### PGE<sub>2</sub> downregulates CTGF expression in chondrocytes

Figure 1 shows the results of quantitative PCR. IL-1β and, to a lesser extent, PGE<sub>2</sub> significantly suppressed the expression of CTGF mRNA. We observed a similar effect of PGE<sub>2</sub> in chondrocyte samples from patients with traumatic fracture and rheumatoid arthritis (data



not shown). The suppressive effect of PGE<sub>2</sub> on CTGF mRNA expression tended to be more potent 4 h after stimulation than at 24 h after stimulation (Figure 2), suggesting that suppression occurred at the transcriptional level. A comparison of the effect of EP agonists and the effect of PGE<sub>2</sub> indicated that the effects of butaprost (EP2 agonist) and PGE<sub>1</sub> alcohol (EP2/EP4 agonist) were similar to those of PGE<sub>2</sub>. Sulprostone (EP1/EP3

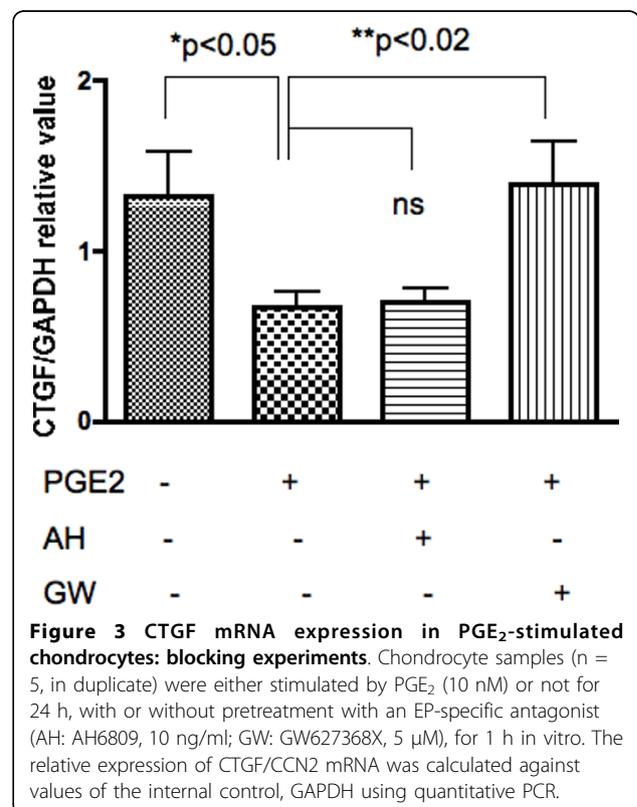


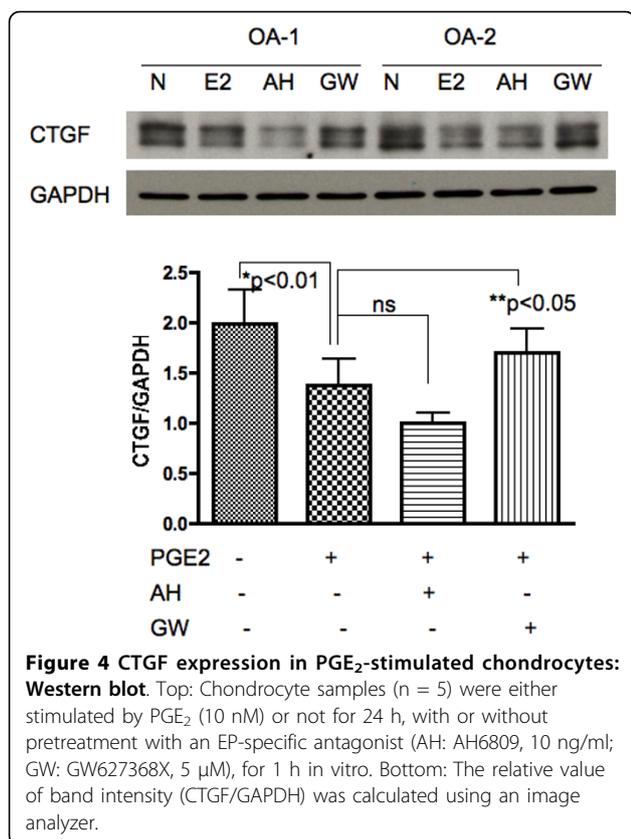
agonist), however, did not have a suppressive effect. These findings suggest that EP2 and/or EP4 receptors are involved in the suppression of CTGF expression by PGE<sub>2</sub> (Figures 1). Based on previous reports that butaprost might exert agonistic effects on EP4 at micromolar concentrations in some cells [16], we further analysed the effect of PGE<sub>2</sub> using EP2/EP4-specific antagonists. An EP4-specific agonist was not commercially available.

Chondrocytes were stimulated with PGE<sub>2</sub> in the presence or absence of AH6809 (EP2 antagonist) or GW627368X (EP4 antagonist). The results of quantitative PCR using these antagonists revealed that GW627368X, but not AH6809, blocked the suppressive effect of PGE<sub>2</sub> on CTGF expression (Figure 3), suggesting that the EP4 receptor has a dominant role in the effects of PGE<sub>2</sub> on CTGF. Western blot analyses produced similar results (Figure 4), confirming that the effects of PGE<sub>2</sub> were mediated through the EP4 receptor at the protein level. Of note, in the Western blot, CTGF protein was detected as several bands of approximately 30 and 35 kD (Figure 4, top). This finding suggests the presence of cleaved fragments, as reported in previous studies [5,8,17].

#### Suppression of CTGF secretion by PGE<sub>2</sub>

We measured secreted levels of CTGF from cultured OA chondrocytes by ELISA. Basal CTGF levels were detected in the culture supernatant of three of five OA



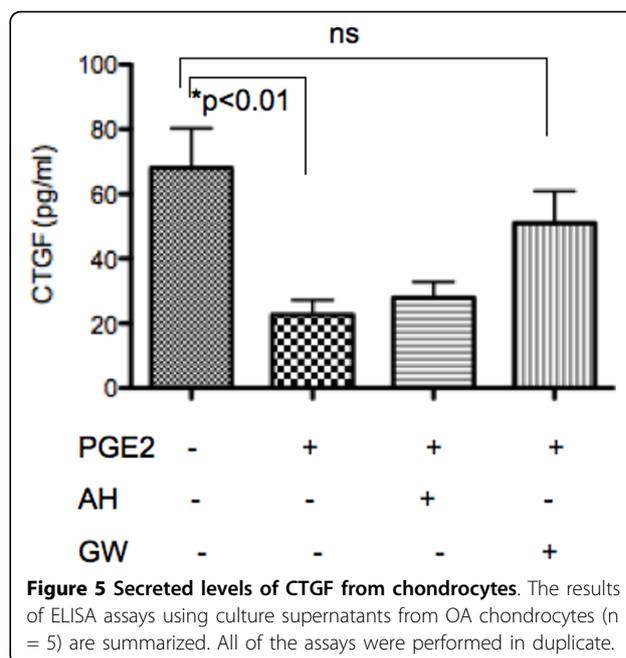


chondrocyte samples. In all tested samples, adding PGE<sub>2</sub> to the cells significantly decreased CTGF secretion (Figure 5). As a result, preincubation with AH6809 did not significantly prevent the decrease in CTGF secretion, whereas GW267368X abolished the suppressive effect of PGE<sub>2</sub> (Figure 5).

### Discussion and Conclusions

CTGF/CCN2 is currently attracting attention for its possible contribution to chondrocyte metabolism, including chondrogenesis, chondrocyte proliferation, and extracellular matrix production [4]. Although CTGF is expressed in human OA chondrocytes as well as in normal human chondrocytes [6,7], the regulatory mechanisms of its expression are not fully understood. Here, we evaluated whether the catabolic mediator PGE<sub>2</sub> is involved in the regulation of CTGF expression in articular chondrocytes. Both PGE<sub>2</sub> and IL-1β suppressed CTGF expression in human articular chondrocytes, and the inhibitory effect of PGE<sub>2</sub> was EP4 receptor-dependent.

PGE<sub>2</sub> exerts its effects through the family of EP receptors, i.e., EP1, EP2, EP3, and EP4. Each EP receptor has a distinct distribution pattern and shows different responses after PGE<sub>2</sub> stimulation (reviewed in [18]). EP2 and EP4 receptors are suggested to deliver important



signals to chondrocytes [19-21]. In this regard, Aoyama et al. previously reported the dominant expression of EP2 receptors in human articular cartilage and cultured chondrocytes, whereas significant expression of EP1 and EP4 receptors was not detected [20]. Otsuka and Aoyama et al. also reported that selective stimulation of the EP2 receptor promoted cartilage regeneration in an animal model with chondral defects [22], suggesting that EP2 has a dominant role in chondrocyte proliferation and cartilage regeneration. Attur et al. [13], however, recently demonstrated that all four EP receptors (EP1-EP4) are expressed in both OA and normal cartilage and in chondrocytes, and that the catabolic effects of PGE<sub>2</sub> are mediated by signaling through EP4 receptors in OA cartilage. Specifically, the authors demonstrated that PGE<sub>2</sub>-induced matrix metalloproteinase production and type II collagen degradation were both EP4-dependent [13]. In addition, Li et al. demonstrated the predominant expression of EP2 and EP4 receptors in human articular cartilage, and showed that PGE<sub>2</sub> works as a potent anti-anabolic factor via the suppression of proteoglycan production, which suggests a potential therapeutic application of EP2 and EP4 receptor antagonists [21]. Our results are consistent with their findings in regard to the implication of the involvement of EP4 receptors in the catabolic response to PGE<sub>2</sub> in chondrocytes. Specifically, CTGF mRNA and protein expression was suppressed by an EP4-selective antagonist. Thus, regulation of the EP4-mediated signal might be an important target to suppress the degradation process of inflamed articular cartilage. Of note, Li et al. also reported that the expression levels of each EP receptor

might vary between knee and ankle joints [21]. In the present study, we used 15 knee samples and 2 hip joints without any significant difference in the CTGF responses; however, further studies should be performed to clarify whether chondrocytes in various joints respond differently to PGE<sub>2</sub>.

CTGF has several potential functions in chondrocytes. During chondrogenesis, CTGF has an important role as a regulator of matrix remodeling and chondrocyte differentiation [3,23,24]. Woods et al. recently demonstrated that CTGF expression in chondrocytes is regulated by a small GTPase Rac1 and actin organization via the Smad binding site in the *CTGF* promoter gene, suggesting the involvement of TGF- $\beta$ /smad signaling in CTGF expression [25]. On the other hand, using rabbit auricular (not articular) cartilage, Fujisawa et al. reported that CTGF might increase proteoglycan synthesis and the expression of elastin and type II collagen, whereas it suppresses apoptotic cell death [26]. Another recent paper by Maeda et al. reported that CTGF interacts with bone morphologic protein 2 and the complex might regulate the differentiation of chondrocytic cells [27]. Further, Nishida et al. recently demonstrated that CTGF regulates the expression of vascular endothelial growth factor in chondrocytic cells under hypoxic conditions [28]. Based on these findings, the authors suggested a role for CTGF as a "signal conductor" in chondrocytes during endochondral ossification [27,28]. Nevertheless, how CTGF suppression by IL-1 and PGE<sub>2</sub> modifies the pathophysiology of OA remains unclear. Considering that CTGF might regulate extracellular matrix remodeling and is a pro-fibrotic factor [3], it is possible that CTGF is a downstream factor to inflammatory mediators such as IL-1 and PGE<sub>2</sub>, which modulate the balance between synthesis and degradation of the matrix in the cartilage in inflammatory arthritis.

As for the different bands of CTGF detected in Western blot (Figure 4), this finding may indicate the presence of degradation products and/or posttranscriptional modification by, for example, glycosylation of CTGF expressed in chondrocytes. The existence of similar fragments/byproducts has been reported in previous studies [5]: for example, Yang et al. reported that endogenous human CTGF is a secreted and glycosylated protein of approximately 32 to 38 kDa with 208 kDa of N-linked sugars and a 30-kD core [8]. Yang et al. also demonstrated the presence of differentially degraded forms of CTGF (18 kDa and 24 kDa) in biological fluids, including normal human sera. In addition, Zarrinkalam et al. demonstrated that CTGF protein is expressed in human peritoneal mesothelial cells at 36 to 38 kDa and also at proteolytically processed fragments of lower molecular weight (23 and 25 kDa) [17]. Western blot analysis in the present study revealed predominant bands at

approximately 30 to 32 kDa, suggesting that CTGF is deglycosylated in chondrocytes. This point should be further analysed to clarify the cleavage and function of each CTGF isoform in chondrocytes.

In conclusion, we demonstrated that PGE<sub>2</sub> regulates CTGF expression in chondrocytes at both the mRNA and protein levels via the EP4 receptor. Further investigation of the role of CTGF and the PGE<sub>2</sub> EP4 receptor in human chondrocytes might provide clues to establish novel therapeutic strategies against osteoarthritis.

#### Abbreviations

CTGF/CCN2: Connective tissue growth factor/CCN family member 2; TGF: transforming growth factor, PGE<sub>2</sub>: prostaglandin E<sub>2</sub>; OA: osteoarthritis; PCR: polymerase chain reaction; GAPDH: glyceraldehydes-3-phosphate dehydrogenase; ELISA: enzyme-linked immunosorbent assay; IL-1: interleukin-1

#### Acknowledgements

The authors thank Ms. T. Mogi, T. Kayanuma, M. Suzuki, and H. Ogasawara for their excellent technical assistance. The study was supported in part by Grants-in-Aid from the Japan Medical Women's Association, and from the Japanese Ministry of Education, Culture, Sports, Science, and Technology.

#### Author details

<sup>1</sup>Department of Biochemistry, St. Marianna University School of Medicine, 2-16-1 Sugao, Miyamae-ku, Kawasaki-shi, Kanagawa 216-8511, Japan.

<sup>2</sup>Department of Frontier Medicine, Institute of Medical Science, 2-16-1 Sugao, Miyamae-ku, Kawasaki-shi, Kanagawa 216-8512, Japan. <sup>3</sup>Department of Orthopedic Surgery, St. Marianna University School of Medicine, 2-16-1 Sugao, Miyamae-ku, Kawasaki-shi, Kanagawa 216-8511, Japan. <sup>4</sup>Department of Rheumatology, Nippon Medical School, 1-1-5 Sendagi, Bunkyo-ku 113-8603, Tokyo, Japan.

#### Authors' contributions

KM and MM designed and conducted the study. KY, HN, and TK interpreted the data. YS and MB collected the clinical samples and data. All authors approved the final version of the manuscript.

#### Competing interests

None of the authors in this study has any financial personal relationship with any organization that could influence (bias) this work.

Received: 1 October 2009

Accepted: 15 January 2010 Published: 15 January 2010

#### References

1. Perbal B: CCN proteins: multifunctional signalling regulators. *Lancet* 2004, **363**(9402):62-64.
2. Takigawa M, Nakanishi T, Kubota S, Nishida T: Role of CTGF/HCS24/ecogenin in skeletal growth control. *J Cell Physiol* 2003, **194**(3):256-266.
3. Shi-Wen X, Leask A, Abraham D: Regulation and function of connective tissue growth factor/CCN2 in tissue repair, scarring and fibrosis. *Cytokine Growth Factor Rev* 2008, **19**(2):133-144.
4. Nishida T, Kawaki H, Baxter RM, Deyoung RA, Takigawa M, Lyons KM: CCN2 (Connective Tissue Growth Factor) is essential for extracellular matrix production and integrin signaling in chondrocytes. *J Cell Commun Signal* 2007, **1**(1):45-58.
5. Leask A, Abraham DJ: All in the CCN family: essential matricellular signaling modulators emerge from the bunker. *J Cell Sci* 2006, **119**(Pt 23):4803-4810.
6. Kumar S, Connor JR, Dodds RA, Halsey W, Van Horn M, Mao J, Sathe G, Mui P, Agarwal P, Badger AM, et al: Identification and initial characterization of 5000 expressed sequenced tags (ESTs) each from adult human normal and osteoarthritic cartilage cDNA libraries. *Osteoarthritis Cartilage* 2001, **9**(7):641-653.
7. Omoto S, Nishida K, Yamaai Y, Shibahara M, Nishida T, Doi T, Asahara H, Nakanishi T, Inoue H, Takigawa M: Expression and localization of

- connective tissue growth factor (CTGF/Hcs24/CCN2) in osteoarthritic cartilage. *Osteoarthritis Cartilage* 2004, **12**(10):771-778.
8. Yang DH, Kim HS, Wilson EM, Rosenfeld RG, Oh Y: **Identification of glycosylated 38-kDa connective tissue growth factor (IGFBP-related protein 2) and proteolytic fragments in human biological fluids, and up-regulation of IGFBP-rP2 expression by TGF-beta in Hs578T human breast cancer cells.** *J Clin Endocrinol Metab* 1998, **83**(7):2593-2596.
  9. Kubota S, Moritani NH, Kawaki H, Mimura H, Minato M, Takigawa M: **Transcriptional induction of connective tissue growth factor/hypertrophic chondrocyte-specific 24 gene by dexamethasone in human chondrocytic cells.** *Bone* 2003, **33**(4):694-702.
  10. Nakao K, Kubota S, Doi H, Eguchi T, Oka M, Fujisawa T, Nishida T, Takigawa M: **Collaborative action of M-CSF and CTGF/CCN2 in articular chondrocytes: possible regenerative roles in articular cartilage metabolism.** *Bone* 2005, **36**(5):884-892.
  11. Ricupero DA, Rishikof DC, Kuang PP, Poliks CF, Goldstein RH: **Regulation of connective tissue growth factor expression by prostaglandin E(2).** *Am J Physiol* 1999, **277**(6 Pt 1):L1165-1171.
  12. Yu J, Prado GN, Schreiber B, Polgar P, Taylor L: **Role of prostaglandin E(2) EP receptors and cAMP in the expression of connective tissue growth factor.** *Arch Biochem Biophys* 2002, **404**(2):302-308.
  13. Attur M, Al-Mussawir HE, Patel J, Kitay A, Dave M, Palmer G, Pillinger MH, Abramson SB: **Prostaglandin E2 exerts catabolic effects in osteoarthritis cartilage: evidence for signaling via the EP4 receptor.** *J Immunol* 2008, **181**(7):5082-5088.
  14. Altman R, Asch E, Bloch D, Bole G, Borenstein D, Brandt K, Christy W, Cooke TD, Greenwald R, Hochberg M, et al: **Development of criteria for the classification and reporting of osteoarthritis. Classification of osteoarthritis of the knee. Diagnostic and Therapeutic Criteria Committee of the American Rheumatism Association.** *Arthritis Rheum* 1986, **29**(8):1039-1049.
  15. Murata M, Yudo K, Nakamura H, Chiba J, Okamoto K, Suematsu N, Nishioka K, Beppu M, Inoue K, Kato T, et al: **Hypoxia upregulates the expression of angiopoietin-like-4 in human articular chondrocytes: role of angiopoietin-like-4 in the expression of matrix metalloproteinases and cartilage degradation.** *J Orthop Res* 2009, **27**(1):50-57.
  16. Kopp UC, Cicha MZ, Nakamura K, Nusing RM, Smith LA, Hokfelt T: **Activation of EP4 receptors contributes to prostaglandin E2-mediated stimulation of renal sensory nerves.** *Am J Physiol Renal Physiol* 2004, **287**(6):F1269-1282.
  17. Zarrinkalam KH, Stanley JM, Gray J, Oliver N, Faull RJ: **Connective tissue growth factor and its regulation in the peritoneal cavity of peritoneal dialysis patients.** *Kidney Int* 2003, **64**(1):331-338.
  18. Sugimoto Y, Narumiya S: **Prostaglandin E receptors.** *J Biol Chem* 2007, **282**(16):11613-11617.
  19. Miyamoto M, Ito H, Mukai S, Kobayashi T, Yamamoto H, Kobayashi M, Maruyama T, Akiyama H, Nakamura T: **Simultaneous stimulation of EP2 and EP4 is essential to the effect of prostaglandin E2 in chondrocyte differentiation.** *Osteoarthritis Cart* 2003, **11**(9):644-652.
  20. Aoyama T, Liang B, Okamoto T, Matsusaki T, Nishijo K, Ishibe T, Yasura K, Nagayama S, Nakayama T, Nakamura T, et al: **PGE2 signal through EP2 promotes the growth of articular chondrocytes.** *J Bone Miner Res* 2005, **20**(3):377-389.
  21. Li X, Ellman M, Muddasani P, Wang JH, Cs-Szabo G, van Wijnen AJ, Im HJ: **Prostaglandin E2 and its cognate EP receptors control human adult articular cartilage homeostasis and are linked to the pathophysiology of osteoarthritis.** *Arthritis Rheum* 2009, **60**(2):513-523.
  22. Otsuka S, Aoyama T, Furu M, Ito K, Jin Y, Nasu A, Fukiage K, Kohno Y, Maruyama T, Kanaji T, et al: **PGE2 signal via EP2 receptors evoked by a selective agonist enhances regeneration of injured articular cartilage.** *Osteoarthritis Cart* 2009, **17**(4):529-538.
  23. Ivkovic S, Yoon BS, Popoff SN, Safadi FF, Libuda DE, Stephenson RC, Daluiski A, Lyons KM: **Connective tissue growth factor coordinates chondrogenesis and angiogenesis during skeletal development.** *Development* 2003, **130**(12):2779-2791.
  24. Pala D, Kapoor M, Woods A, Kennedy L, Liu S, Chen S, Bursell L, Lyons KM, Carter DE, Beier F, et al: **Focal adhesion kinase/Src suppresses early chondrogenesis: central role of CCN2.** *J Biol Chem* 2008, **283**(14):9239-9247.
  25. Woods A, Pala D, Kennedy L, McLean S, Rockel JS, Wang G, Leask A, Beier F: **Rac1 signaling regulates CTGF/CCN2 gene expression via TGFbeta/Smad signaling in chondrocytes.** *Osteoarthritis Cartilage* 2009, **17**(3):406-413.
  26. Fujisawa T, Hattori T, Ono M, Uehara J, Kubota S, Kuboki T, Takigawa M: **CCN family 2/connective tissue growth factor (CCN2/CTGF) stimulates proliferation and differentiation of auricular chondrocytes.** *Osteoarthritis Cartilage* 2008, **16**(7):787-795.
  27. Maeda A, Nishida T, Aoyama E, Kubota S, Lyons KM, Kuboki T, Takigawa M: **CCN family 2/connective tissue growth factor modulates BMP signalling as a signal conductor, which action regulates the proliferation and differentiation of chondrocytes.** *J Biochem* 2009, **145**(2):207-216.
  28. Nishida T, Kondo S, Maeda A, Kubota S, Lyons KM, Takigawa M: **CCN family 2/connective tissue growth factor (CCN2/CTGF) regulates the expression of Vegf through Hif-1alpha expression in a chondrocytic cell line, HCS-2/8, under hypoxic condition.** *Bone* 2009, **44**(1):24-31.

doi:10.1186/1756-0500-3-5

**Cite this article as:** Masuko et al.: Prostaglandin E<sub>2</sub> regulates the expression of connective tissue growth factor (CTGF/CCN2) in human osteoarthritic chondrocytes via the EP4 receptor. *BMC Research Notes* 2010 3:5.

**Submit your next manuscript to BioMed Central  
and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at  
www.biomedcentral.com/submit

