

SHORT REPORT

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Enhancing genome investigations in the mosquito *Culex quinquefasciatus* via BAC library construction and characterization

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Abstract

Background: *Culex quinquefasciatus* (Say) is a major species in the *Culex pipiens* complex and an important vector for several human pathogens including West Nile virus and parasitic filarial nematodes causing lymphatic filariasis. It is common throughout tropical and subtropical regions and is among the most geographically widespread mosquito species. Although the complete genome sequence is now available, additional genomic tools are needed to improve the sequence assembly.

Findings: We constructed a bacterial artificial chromosome (BAC) library using the plIndigoBAC536 vector and *HindIII* partially digested DNA isolated from *Cx. quinquefasciatus* pupae, Johannesburg strain (NDJ). Insert size was estimated by *NotI* digestion and pulsed-field gel electrophoresis of 82 randomly selected clones. To estimate genome coverage, each 384-well plate was pooled for screening with 29 simple sequence repeat (SSR) and five gene markers. The NDJ library consists of 55,296 clones arrayed in 144 384-well microplates. Fragment insert size ranged from 50 to 190 kb in length (mean = 106 kb). Based on a mean insert size of 106 kb and a genome size of 579 Mbp, the BAC library provides ~10.1-fold coverage of the *Cx. quinquefasciatus* genome. PCR screening of BAC DNA plate pools for SSR loci from the genetic linkage map and for four genes associated with reproductive diapause in *Culex pipiens* resulted in a mean of 9.0 positive plate pools per locus.

Conclusion: The NDJ library represents an excellent resource for genome assembly enhancement and characterization in *Culex pipiens* complex mosquitoes.

Keywords: Bacterial Artificial Chromosome, Culicidae, *Culex pipiens/quinquefasciatus*, Lymphatic filariasis, West Nile virus, *Wuchereria bancrofti*

Introduction

Culex quinquefasciatus (Say), the southern house mosquito, is a major vector for a number of important human pathogens including West Nile virus and *Wuchereria bancrofti*, the primary global etiologic agent for lymphatic filariasis (LF) [1-3]. It is estimated that more than 1.2 billion people are at risk for infection by parasites causing LF, with 120 million people presently infected [4]. Among these are over 40 million people who suffer from chronic morbidity associated with lymphedema and hydrocele [5]. Despite the availability of

effective antihelminthics to treat and prevent infections, the damage to the lymphatic system caused by these parasites is largely irreversible. Although efforts to eradicate LF globally using mass drug administration to human populations in endemic areas were initiated in 2000, the success of these efforts will likely also rely on the implementation of effective mosquito vector control strategies [6]. However, vector control efforts can be hindered by the rapid selection for emergence of insecticide resistance [7]. Consequently, the identification of new targets for insecticides as well as the development of novel vector control strategies is expected to play a large role in the successful control and/or eradication of mosquito-borne diseases [8].

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Cx. quinquefasciatus and *Cx. pipiens* (L.) are the two most common and geographically widespread species in the *Cx. pipiens* complex, a species complex with nearly worldwide distribution [9]. *Cx. quinquefasciatus* is common in tropical and subtropical regions while *Cx. pipiens*, the northern house mosquito, occupies more temperate regions. Both species are abundant in urban areas where they oviposit in stagnant, and often polluted water. They frequently enter homes and feed on humans during the night, hence the common name of house mosquito. The taxonomic status of this complex has been a subject of debate, and these taxa are sometimes placed within a single species, i.e., *Cx. pipiens quinquefasciatus* or *Cx. pipiens pipiens* [9]. Introgression between these species is common in the United States where hybrids can be found as far south as Louisiana and as far north as Illinois [10-12], yet in South Africa the populations remain largely distinct [13,14]. Females are morphologically indistinguishable, while differences in male genitalia have been used to identify species as well as interspecies hybrids [10-12,15]. Recently, however, PCR assays have been developed to aid in the differentiation of species in this complex [16-19].

Given their medical importance, *Cx. pipiens* complex mosquitoes have garnered considerable attention by the scientific community during the last 100 years [9]. Nevertheless, the current status of contemporary *Cx. pipiens* genetics remains considerably behind that of other important mosquito vectors such as *Anopheles gambiae* and *Aedes aegypti* [20]. The *Cx. quinquefasciatus* (Johannesburg strain) genome sequence was recently determined using the whole genome shotgun (wgs) approach, thus providing a valuable resource for advancing genome studies in this species complex [21]. However, the genome assembly remains highly fragmented and few (~40) of the 3171 supercontigs have been assigned to their respective chromosomes [21].

Bacterial artificial chromosome (BAC) genomic libraries are important resources for the assembly and characterization of complex genomes. They have been utilized for the assembly of numerous genomes including *Drosophila melanogaster* and *An. gambiae* [22,23]. BAC libraries have also been used for the development of genetic markers for non-model organisms [24,25]. Furthermore, BAC clones can be used for positional cloning to help identify and characterize genomic regions of interest [26,27], as well as for construction of BAC-based physical map assemblies [28-31]. These are useful for long-range contiguity and anchoring of wgs draft assemblies as well as targeted re-sequencing for high resolution using BAC pools [32]. The objective of this work was to construct a BAC library with comprehensive coverage of the *Cx. quinquefasciatus* genome, thereby providing a tool to aid in genome assembly,

marker development, and gene discovery in *Cx. pipiens* complex mosquitoes.

Methods

BAC library construction

High molecular weight DNA was extracted from pupae from the Johannesburg (JHB) strain. This strain was established using individuals from Johannesburg South Africa, and was the strain used in the *Cx. quinquefasciatus* genome project [21]. Pupae were gently homogenized in 1X PBS buffer containing 50 mM EDTA pH 8.0 and 0.1% BME and filtered through one layer of miracloth into 50 mL Falcon tubes. Cells were pelleted by centrifugation in a swinging-bucket rotor (Beckman) at 3,200 rpm for 15 minutes at 4°C. Pellets were washed 2 additional times with PBS and gently resuspended in 1 mL of PBS. The nuclei solution was warmed to 45°C in a waterbath, mixed gently with an equal volume of 1.5% low-melt agarose (Seaplaque) and aliquoted into plug molds (BioRad) using large-bore tips. Protein digestion and plug washing was performed exactly as the methods of Luo and Wing (2003) [33].

*Hind*III partial restriction enzyme digestion of DNA, as well as the preparation of high molecular weight DNA fragments was conducted following the procedure of Luo and Wing (2003) [33]. Preparation of the *Hind*III cloning-ready single copy pIndigoBAC536 vector from the high copy pCUGIBAC1 plasmid was performed according to Luo et al. (2001) [34]. The size selected high molecular fragments were ligated to the vector and transformed into *E. coli* strain DH10B competent cells (Invitrogen, Carlsbad, CA). White recombinant colonies were selected on LB plates containing chloramphenicol, X-Gal and IPTG, and picked robotically using the Genetix Q-bot (Genetix, UK). Recombinant clones were transferred into individual wells of microtiter plates, grown and then stored at -80°C. The BAC library was also gridded onto 10, 11.25 × 22.25 cm filters in high density, double spots (18,432 clones represented per filter) and 4 × 4 patterns.

To estimate the size of the BAC inserts, DNA from 82 randomly selected clones was prepared according to standard alkaline lysis protocol, digested with *Not*I, and separated by pulsed-field gel electrophoresis (PFGE) on a 1% agarose gel under the following conditions: 5-15 sec linear ramp time, 6 V/cm, 14°C in 0.5 × TBE buffer for 15 hours and stained with ethidium bromide. Insert sizes of the clones with endogenous *Not*I sites, evidenced by multiple restriction fragments, were estimated by summing the fragments. Southern blotting was used to confirm that all of the clones were truly *Cx. quinquefasciatus* and are not significantly contaminated by other types of DNA. One gel used for insert size determination was transferred to a positively charged nylon

Table 1 SSR primer sequences

SSR locus	GenBank accession	SSR Primer Sequence (F/R)	Product size (bp) ^a	# Positive plate pools
C127GAC1	GF102017	GCGTTTGGAGAGTGAAAAAG TGAGTTTTTCAGTGCCCTCCT	307	10
C32AC1B	GF110611	AAACGATCGCAATTCGAAAC GTGGCGAACAACATTCACAG	242	3
C32TC1B	GF110612	TCATCGTTCATTCGTTCCAA TGTCATTTTCTGCCTGCATC	179	2
C32TG1	GF102044	CGTGTTCATTGTTGGTG TTGGCTGTGCAACTGCTTC	400	29
C68ACAT1	GF102032	GGCCTTGCTGAGAAAATTG CCCAAAATCCAAGCTTCAAA	425	1
C68CA1	GF102033	ATAAAGCGACCAAGGCTCAA GCGAAACCATTCAAAAGCAT	294	7
C68GA1B	GF110613	CACCCACAGTTAACCCAAC CTCGAGAGATTTGGCCTTTG	245	8
C65AC1	GF102022	GGAGTTGTGCGGTTGAAAGT GCACTGCCTAACGGATCATT	305	19
C65CGC1	GF102023	TCTGGGTACAACCCCGTAAC AGAGAGTGCACAAAAGCAAT	221	20
C65TG1	GF102021	ACTGCGAAACGCTTACTGCT GTGTGTGGACTGTGGTGGAG	302	9
C474CT1B	GF110614	CCCAAACCTGCCACAAAAGT CTCACTCTCCGTGAACGACA	290	2
C48ATC1	GF102034	CATTTTTCGGGTGGCTTCTA CGAGATCGAAATGATGCTGA	337	7
C48CGA1B	GF110615	GCTTGGGAATCTGAATCTGC ACCTTGCAATCAACGAGCTT	251	4
C48GTT1B	GF110616	GTGGCCACCTGGTTGAGTT ACCACCGGTAGAACATCTCG	309	23
C175AT1	GF102036	GGACCAAGGGTACGATTTGA CAGACTGGTTAACGGCTTCC	185	14
C175TG1	GF102045	TCAGATCTCCGAGAGGAGGA CTGTCAGGGCCAGATTCAT	295	4
C134AC1	GF102037	GAAGGTCAGCCACTCAGGC ACAGCTGACTCTCGTCGAC	194	0
C129GT1	GF102038	AAGGTGCAAAACCAAACTGG TGGAGCACAGCCCCTACTCTT	377	1
C66CA1	GF102026	CGACTACTGCCCAATTTGT CACCCCTCCCCTACAGACGTA	213	2
C177CA1B	GF110618	AGGGCAATGTTACGACGAC CTTGCGCCTTAGTCATCCTC	293	2
C177TG1	GF102028	AGCACAAAAAGGCACGATTT TAAACGCAAGTAGCGGAGT	197	6
C99TGT1B	GF110619	GCAGTGGAGGATTCTGAGGA CAGAACGTTTGGCGAATTTT	358	5
C205CA1	GF102029	CAATGCGCCTTCTGGATTAT CTCGTGATGGCCATTTCTCT	227	3
C205TG1B	GF110620	ATTGCTCAAGTGTGCCTTT ATGACGACGAAAAACCGAAC	212	7
C139TG1B	GF110621	GGGATCGCTACGTGTTTTGT TCTCGGAATGCCAGTCTTTT	265	6
C446AC2	GF110622	CATACGACGTGGAACAAACG	162	17

Table 1 SSR primer sequences (Continued)

C446TG1	GF102043	ACGAGGTTGAGGTTGGTGAC GGAAAGGGGCACTTGTGTAA CGTTTGCTTCTTTCGAACC	397	22
CxpGT4	AY423738	GTCGTCGTAACCCTTGTT CGCGATAGTCGGTAATCGT	146	2
CxqTri4	AY958079	CTAGCCCGGTATTTACAAGAAC AACGCCAGTAGTCTCAGCAG	121	16

^aPredicted amplicon size based on nucleotide sequence in VectorBase [50].

Table 2 Primer sequences for genes used in library screening

Gene	VectorBase gene ID	GenBank accession	SSR Primer Sequence (F/R)	Product size (bp) ^a	# Positive plate pools
CHP*	CPIJ007110	GF110930	CGAGCAGTTCAAACACCAGA GCTTCTTCAGGTTGCTCCAC	207	12
FOXO	CPIJ016794	GF110931	CTGAGCCCAATTCAGTCCAT TCTGCTGTTAAAGTCAGCTCGTC	187	3
ILP-1	CPIJ018051	GF110932	AGTCCCTCGGAGGAGTTCAA TCGGCACAGTACTGCTTGAG	163	7
ILP-2	CPIJ018050	GF110933	TCCAGCAGATCTTCGATGC TGTAGATCGGGGAAGTCTGTC	140	10
ILP-5	CPIJ001698	GF110934	GGTTCATCACGCAGGAGT GTTGATCCGCTTGTCGAC	87	23

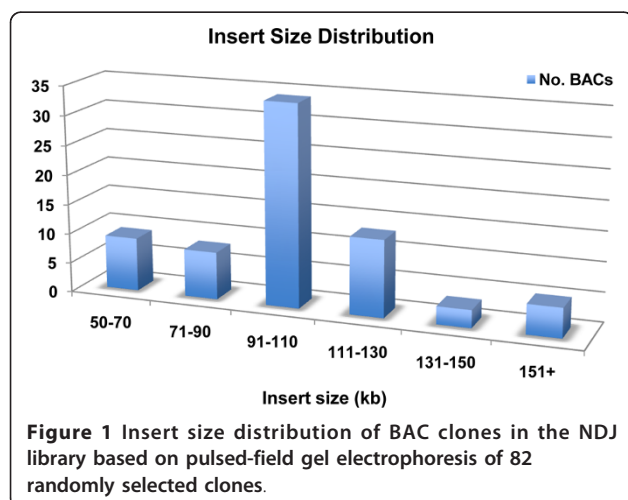
^aPredicted amplicon size based on nucleotide sequence in VectorBase [50]. *CHP: Conserved hypothetical protein.

membrane Hybond N⁺ (GE Healthcare) following the methods of Chomczynski (1992) [35]. BAC vector (pIndigoBAC536) and total *Culex* DNA were used as probes and radiolabeled with the DECAprime™ II kit (Ambion, Inc). The probes were mixed and denatured, and hybridization was carried out overnight at 60°C. The membrane was washed with 1× SSC, 0.1% SDS at 60°C twice for one hour each. The membrane was exposed to a phosphor screen (GE Healthcare) overnight and the image recorded by a Typhoon 9400 imager (GE Healthcare).

BAC library screening

Screening of the BAC library was generally performed as described by Jiménez *et al.* (2004) [36]. Briefly, we first prepared pools of DNA representing all clones within each of the individual 144 384-well microplates. Plates were initially replicated on LB agar plates containing 12.5 µg/ml chloramphenicol and incubated overnight at 37°C. The plates were then flooded with LB broth containing 12.5 µg/ml chloramphenicol, agitated for 4 h at 37°C and the slurries used to prepare 9.5 ml overnight cultures. These individual plate pool cultures were used for large-scale alkaline lysis DNA extractions [37] and subsequent PCR screening with SSR oligonucleotide primer sets.

A combination of PCR-based plate-pool DNA screening and radiolabeled oligonucleotide probe hybridization was used to screen the NDJ library. Initial PCR-based screening of BAC DNA representing individual plate pools was performed using 29 simple sequence repeats (SSRs) (Table 1) [38-40]. The Primer3 program [41] was used to design primers to amplify regions within a gene on supercontig 3.134 and four genes previously identified as having a role in reproductive diapause in *Cx. pipiens* s.s. (Table 2) [42,43]. PCR reactions were performed in a total volume of 25 µl containing 50 mM KCl, 10 mM Tris (pH 9.0), 0.1% Triton X, 1.5 mM MgCl₂, 200 µM dNTPs, 5 pmol of each primer (F and R), 25 ng of plate pool DNA and 1 unit of *Taq*



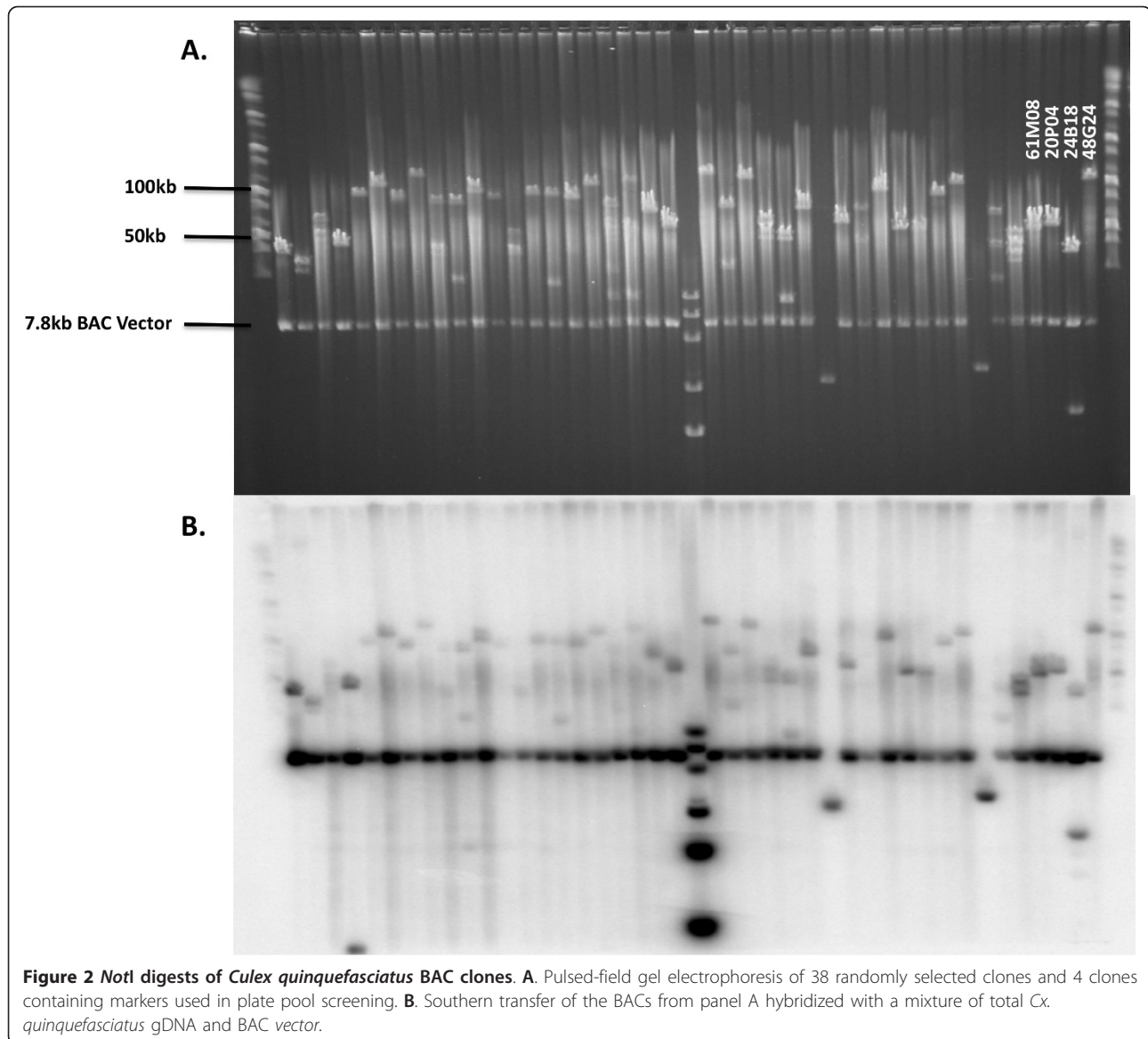


Figure 2 *NotI* digests of *Culex quinquefasciatus* BAC clones. **A.** Pulsed-field gel electrophoresis of 38 randomly selected clones and 4 clones containing markers used in plate pool screening. **B.** Southern transfer of the BACs from panel A hybridized with a mixture of total *Cx. quinquefasciatus* gDNA and BAC vector.

polymerase. PCR thermal cycling conditions were 5 min. at 94°C, followed by thirty cycles of 1 min. at 94°C, 1 min. at 60°C, 2 min. at 72°C, and then 10 min. at 72°C for a final extension. The SSR positive plate pools were identified by electrophoresis on 2% agarose gels using ethidium bromide and UV visualization.

Well position of marker loci within select positive microplates was determined by DNA-DNA hybridization. Four individual clones were identified by probing with P³²-labeled PCR amplicons (C127GAC1, C65AC1, C99TGT1, and FOXO) and thereafter sized with PFGE. Briefly, microplates representing positive pools were replicated to Colony/Plaque screen hybridization membranes (NEN™, Life Science Products) following Jiménez *et al.* [36]. Hybridizations and radiolabeling of the

target clones were conducted following our standard probing procedures [44]. The presence of the marker locus in each of the four clones was confirmed by PCR and UV visualization on 2% agarose gels, as described for plate pool screening.

Results and Discussion

We have constructed a BAC library for *Cx. quinquefasciatus*, an important human disease vector and a major species in the *Cx. pipiens* complex, using high molecular weight DNA extracted from Johannesburg strain pupae and partially digested with *HindIII*. The Notre Dame Johannesburg (NDJ) library consists of 144 384-well microplates containing 55,296 clones. *NotI* digestion and pulsed-field gel electrophoresis of 82 randomly selected

clones produced fragments ranging from 50 to 190 kb in length (mean = 106 kb) and no empty vectors (Figures 1 and 2). Southern blotting with BAC vector and *Cx. quinquefasciatus* gDNA indicated that the inserts are of *Culex* origin and all BACs appear to be fully digested (Figure 2B). Based on a mean insert size of 106 kb and a genome size of 579 Mbp, the BAC library provides ~10.1-fold coverage of the *Cx. quinquefasciatus* genome.

To further assess the quality of the library, we prepared and screened DNA extractions representing each of the 144 plate pools with 29 simple sequence repeat (SSR) markers representing all three linkage groups (Figure 3). The number of positive plate pools for each SSR ranged from 0 to 29, resulting in a mean of 8.7 positive plate pools per screen. Only one of the SSR markers (C134AC1) did not amplify in any of the plate pools. Nevertheless, a gene sequence (CPIJ007110) on the same supercontig (3.134), ~80 kb downstream, amplified in 12 plate pools. In addition to the SSRs, we screened the plate pools with primers designed to amplify sequences

within exons of four genes previously determined to have a role in reproductive diapause in *Cx. pipiens sensu stricto* (s.s.) [42,43]. The number of positive plate pools for the gene sequences ranged from three to 23, resulting in a mean of 10.8 positive plate pools per gene (Table 2). The size distribution of the four individual clones selected by probing with radiolabeled markers is similar to the size distribution of the library (Figure 2A). The overall mean number of positive plate pools for the 29 SSRs and five genes used to screen the library was 9.0, indicating that the NDJ BAC library represents ~9 BAC clones per marker across the *Cx. quinquefasciatus* genome, assuming that only one BAC clone per 384-well plate pool contains the target sequence.

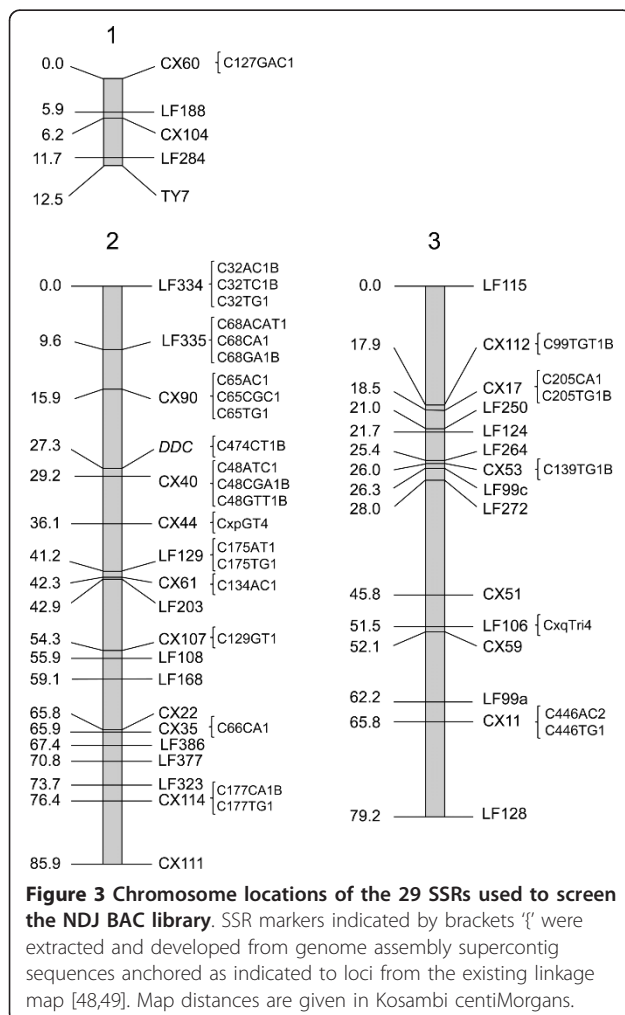
Detailed genetic and genomic studies among the *Cx. pipiens* complex could provide valuable insights into the molecular genetic mechanisms influencing important traits such as vector competence, insecticide resistance, and reproductive diapause. Despite morphological similarities and their ability to form hybrid populations, species within the complex differ in several life history traits. For example, *Cx. quinquefasciatus* requires a blood meal prior to laying eggs (anautogenous) and is unable to enter diapause and overwinter in cold climates. *Cx. pipiens* and *Cx. pipiens pallens* also are anautogenous but adult females are able to enter reproductive diapause and survive winter in temperate climates, and *Cx. pipiens molestus* is able to lay eggs without taking a blood meal (autogenous) but does not enter diapause [45-47]. Presently, detailed molecular analyses of these traits are limited by the fragmented genome assembly. Fingerprinting, end-sequencing and physical assembly of the NDJ BAC library would likely facilitate the construction of a more complete genome sequence assembly by serving as a template for genome finishing, including gap-filling, as well as providing resources to enable the assignment of the individual superscaffolds to their respective chromosome position via *in situ* hybridization. In summary, the NDJ BAC library provides a valuable resource for marker development, positional cloning, and genome sequence assembly enhancement for *Cx. quinquefasciatus* thus helping to advance genome studies in *Cx. pipiens* complex mosquitoes.

Library availability

The NDJ BAC library is available to researchers through the Clemson University Genomics Institute (see *Culex pipiens* library CPQLBa at <http://www.genome.clemson.edu/>).

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Authors' contributions

PVH drafted the manuscript and helped design probes for library screening. AM performed the initial genetic mapping and provided the pupae used in library construction. CAS constructed the library, estimated insert sizes and assisted in writing the manuscript. BD and DDL designed probes for library screening. BD performed plate pool screening. The project was conceived and supervised by DWS. All authors approved of the final version.

Competing interests

The authors declare that they have no competing interests.

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References

1. Turell MJ, O'Guinn ML, Dohm DJ, Jones JW: **Vector competence of North American mosquitoes (Diptera: Culicidae) for West Nile virus.** *J Med Ent* 2001, **38**:130-134.
2. Fonseca DM, Keyghobadi N, Malcolm CA, Mehmet C, Schaffner F, Mogi M, Fleischer RC, Wilkerson RC: **Emerging vectors in the *Culex pipiens* complex.** *Science* 2004, **303**:1535-1538.
3. Michalski ML, Erickson SM, Bartholomay LC, Christensen BM: **Midgut barrier imparts selective resistance to filarial worm infection in *Culex pipiens pipiens*.** *PLoS Negl Trop Dis* 2010, **4**:e875.
4. Michael E, Bundy DA, Grenfell BT: **Reassessing the global prevalence and distribution of lymphatic filariasis.** *Parasitology* 1996, **112**:409-428.
5. Addiss DG, Brady MA: **Morbidity management in the global programme to eliminate lymphatic filariasis: a review of the scientific literature.** *Filaria J* 2007, **6**:2-20.
6. Bockarie MJ, Pedersen EM, White GB, Michael E: **Role of vector control in the global program to eliminate lymphatic filariasis.** *Annu Rev Entomol* 2009, **54**:469-487.
7. Hemingway J, Ranson H: **Insecticide resistance in insect vectors of human disease.** *Annu Rev Entomol* 2000, **45**:371-391.
8. Hill CA, Kafatos FC, Stansfield SK, Collins FH: **Arthropod-borne diseases: vector control in the genomics era.** *Nature Rev Microbiol* 2005, **3**:262-268.
9. Vinogradova EB: *Culex pipiens pipiens* mosquitoes: taxonomy, distribution, ecology, physiology, genetics, applied importance and control Sofia: Pensoft; 2000.
10. Barr AR: **The distribution of *Culex p. pipiens* and *Culex p. quinquefasciatus* in North America.** *Am J Trop Med Hyg* 1957, **6**:153-165.
11. Cheng ML, Hacker CS, Pryor SC, Ferrell RE, Kitto GB: **The ecological genetics of the *Culex pipiens* complex in North America.** In *Recent Developments in The Genetics of Insect Disease Vectors*. Edited by: Steiner WWM, Tabachnick WJ, Rai S, Narang KS. Champaign: Stipes; 1982:581-627.
12. Kothera L, Zimmerman EM, Richards CM, Savage HM: **Microsatellite characterization of subspecies and their hybrids in *Culex pipiens* complex (Diptera: Culicidae) mosquitoes along a north-south transect in the central United States.** *J Med Ent* 2009, **46**:236-248.
13. Jupp PG: ***Culex (Culex) pipiens pipiens* Linnaeus and *Culex (Culex) pipiens quinquefasciatus* Say in South Africa: morphological and reproductive evidence in favour of their status as two species.** *Mosq Systematics* 1978, **10**:461-473.
14. Cornel AJ, McAbee RD, Rasgon J, Stanich MA, Scott TW, Coetzee M: **Differences in extent of genetic introgression between sympatric *Culex pipiens* and *Culex quinquefasciatus* (Diptera: Culicidae) in California and South Africa.** *J Med Entomol* 2003, **40**:36-51.
15. Sundararaman S: **Biometrical studies on intergradation in the genitalia of certain populations of *Culex pipiens* and *Culex quinquefasciatus* in the United States.** *Am J Hyg* 1949, **50**:307-314.
16. Crabtree MB, Savage HM, Miller BR: **Development of a species-diagnostic polymerase chain reaction assay for the identification of *Culex* vectors of St. Louis Encephalitis virus based on interspecies sequence variation in ribosomal DNA spacers.** *Am J Trop Med Hyg* 1995, **53**:105-109.
17. Bourguet D, Fonseca D, Yourch G, Dubois M-P, Chandre F, Severini C, Raymond M: **The acetylcholinesterase gene *Ace*: a diagnostic marker for the *quinquefasciatus* and *pipiens* forms of the *Culex pipiens* complex.** *J Am Mosq Control Assoc* 1998, **14**:390-396.
18. Aspen S, Savage H: **Polymerase chain reaction assay identifies North American members of the *Culex pipiens* complex based on nucleotide sequence differences in the acetylcholinesterase gene *Ace2*.** *J Am Mosq Cont Assoc* 2003, **19**:323-328.
19. Smith JL, Fonseca DM: **Rapid assays for identification of members of the *Culex (Culex) pipiens* complex, their hybrids, and other sibling species (Diptera: Culicidae).** *Am J Trop Med Hyg* 2004, **70**:339-345.
20. Severson DW, Brown SE, Knudson DL: **Genetic and physical mapping in mosquitoes: molecular approaches.** *Annu Rev Entomol* 2001, **46**:183-219.
21. Arensburger P, Megy K, Waterhouse RM, Abrudan J, Amedeo P, Antelo B, Bartholomay L, Bidwell S, Caler E, Camara F, Campbell CL, Campbell KS, Casola C, Castro MT, Chandramouliswaran I, Chapman SB, Christley S, Costas J, Eisenstadt E, Feschotte C, Fraser-Liggett C, Guigo R, Haas B, Hammond M, Hansson BS, Hemingway J, Hill SR, Howarth C, Ignell R, Kennedy RC, et al: **Sequencing of *Culex quinquefasciatus* establishes a platform for mosquito comparative genomics.** *Science* 2010, **330**:86-88.
22. Adams MD, Celniker SE, Holt RA, Evans CA, Gocayne JD, Amanatides PG, Scherer SE, Li PW, Hoskins RA, Galle RF, George RA, Lewis SE, Richards S, Ashburner M, Henderson SN, Sutton GG, Wortman JR, Yandell MD, Zhang Q, Chen LX, Brandon RC, Rogers YHC, Blaise RG, Champe M, Pfeiffer BD, Wan KH, Doyle C, Baxter EG, Helt G, Nelson CR, Beasley EM, Birman BP, Bolshakov S, Borkova D, Botchan MR, Bouch J, Brokstein P, Brottier P, Burtis KC, Busam DA, Butler H, Cadieu E, Center A, Chandra I, Cherry JM, Cawley S, Dahlke C, Davenport LB, Davies A, de Pablos B, Delcher A, Deng ZM, Mays AD, Dew I, Dietz SM, Dodson K, Doup LE, Downes M, Dugan-Rocha S, Dunkov BC, Dunn P, Durbin KJ, Evangelista CC, Ferraz C, Ferreira S, Fleischmann W, Fosler C, Gabriellian AE, Garg NS, Gelbart WM, Glasser K, Glodek A, Gong FC, Gorrell JH, Gu ZP, Guan P, Harris M, Harris NL, Harvey D, Heiman TJ, Hernandez JR, Houck J, Hostin D, Houston DA, Howland TJ, Wei MH, Ibegwam C, Jalali M, Kalush F, Karpen GH, Ke ZX, Kennison JA, Ketchum KA, Kimmel BE, Kodira CD, Kraft C, Kravitz S, Kulp D, Lai ZW, Lasko P, Lei YD, Levitsky AA, Li JY, Li ZY, Liang Y, Lin XY, Liu XJ, Mattei B, McIntosh TC, McLeod MP, McPherson D, Merkulov G, Milshina NV, Mobjary C, Morris J, Moshrefi A, Mount SM, Moy M, Murphy B, Murphy L, Muzny DM, Nelson DL, et al: **The genome sequence of *Drosophila melanogaster*.** *Science* 2000, **287**:2185-2195.
23. Holt RA, Subramanian GM, Halpern A, Sutton GG, Charlab R, Nusskern DR, Wincker P, Clark AG, Ribeiro JMC, Wides R, Salzberg SL, Loftus B, Yandell M, Majoros WH, Rusch DB, Lai ZW, Kraft CL, Abril JF, Anthouard V, Arensburger P, Atkinson PW, Baden H, de Berardinis V, Baldwin D, Benes V, Biedler J, Blass C, Bolanos R, Boscus D, Barnstead M, et al: **The genome sequence of the malaria mosquito *Anopheles gambiae*.** *Science* 2002, **298**:129-149.
24. Temnykh S, DeClerck G, Lukashova A, Lipovich L, Cartinhour S, McCouch S: **Computational and experimental analysis of microsatellites in rice (*Oryza sativa* L.): frequency, length variation, transposon associations, and genetic marker potential.** *Genome Res* 2001, **11**:1441-1452.
25. Ellison CK, Shaw KL: **Mining non-model genomic libraries for microsatellites: BAC versus EST libraries and the generation of allelic richness.** *BMC Genomics* 2010, **11**:e428.
26. Perelygin AA, Scherbik SV, Zhulin IB, Stockman BM, Li Y, Brinton MA: **Positional cloning of the murine flavivirus resistance gene.** *Proc Natl Acad Sci USA* 2002, **99**:9322-9327.
27. Luo M, Yu Y, Kim H-R, Kudrna D, Itoh Y, Agate RJ, Melamed E, Goicoechea JL, Talag J, Mueller C, Wang W, Currie J, Sisneros NB, RA W, Arnold AP: **Utilization of a zebra finch BAC library to determine the structure of an avian androgen receptor genomic region.** *Genomics* 2006, **87**:181-190.
28. Quiniou SM-A, Waldbieser GC, Duke MV: **A first generation BAC-based physical map of the channel catfish genome.** *BMC Genomics* 2007, **8**:e40.
29. Palti Y, Luo MC, Hu Y, Genet C, You FM, Vallejo RL, Thorgaard GH, Wheeler PA, Rexroad CE III: **A first generation BAC-based physical map of the rainbow trout genome.** *BMC Genomics* 2009, **10**:e462.
30. Fang G-C, Blackmon BP, Henry DC, Staton ME, Saski CA, Hodges SA, Tomkins JP, Luo H: **Genomic tools development for *Aquilegia*: construction of a BAC-based physical map.** *BMC Genomics* 2010, **11**:e621.
31. Xia JH, Feng F, Lin G, Wang CM, Yue GH: **A first generation BAC-based physical map of the Asian seabass (*Lates calcarifer*).** *PLoS One* 2010, **5**: e11974.

32. Rounsley S, Marri PR, Yu Y, He R, Sisneros N, Goicoechea JL, *et al*: **De novo next generation sequencing of plant genomes.** *Rice* 2009, **2**:35-43.
33. Luo M, Wing RA: **An improved method for plant BAC library construction.** In *Plant Functional Genomics*. Edited by: Grotewold E. Totowa: Humana; 2003:3-19.
34. Luo M, Wang YH, Frisch D, Joobeur T, Wing RA, Dean RA: **Melon bacterial artificial chromosome (BAC) library construction using improved methods and identification of clones linked to the locus conferring resistance to melon *Fusarium* wilt (Fom-2).** *Genome* 2001, **44**:154-162.
35. Chomczynski P: **One-hour downward alkaline capillary transfer for blotting of DNA and RNA.** *Anal Biochem* 1992, **201**:134-139.
36. Jiménez LV, Kang B, deBruyn B, Lovin DD, Severson DW: **Characterization of an *Aedes aegypti* bacterial artificial chromosome (BAC) library and chromosomal assignment of BAC clones for physical mapping quantitative trait loci that influence *Plasmodium* susceptibility.** *Insect Mol Biol* 2004, **13**:37-44.
37. Birnboim HC, Doly J: **A rapid alkaline extraction procedure for screening recombinant plasmid DNA.** *Nucleic Acids Res* 1979, **7**:1513-1523.
38. Keyghobadi N, Matrone MA, Ebel GD, Kramer LD, Fonseca DM: **Microsatellite loci from the northern house mosquito (*Culex pipiens*), a principal vector of West Nile virus in North America.** *Mol Ecol Notes* 2004, **4**:20-22.
39. Smith JL, Keyghobadi N, Matrone MA, Escher RL, Fonseca DM: **Cross-species comparison of microsatellite loci in the *Culex pipiens* complex and beyond.** *Mol Ecol Notes* 2005, **5**:697-700.
40. Hickner PV, deBruyn B, Lovin DD, Mori A, Behura SK, Pinger R, Severson DW: **Genome-based microsatellite development in the *Culex pipiens* complex and comparative microsatellite frequency with *Aedes aegypti* and *Anopheles gambiae*.** *PLoS One* 2010, **5**:e13062.
41. Rozen S, Skaletsky HJ: **Primer3 on the WWW for general users and for biologist programmers.** In *Bioinformatics Methods and Protocols: Methods in Molecular Biology*. Edited by: Krawetz S, Misener S. Totowa: Humana; 2000:365-386.
42. Sim C, Denlinger DL: **Insulin signaling and FOXO regulate the overwintering diapause of the mosquito *Culex pipiens*.** *Proc Natl Acad Sci USA* 2008, **105**:6777-6781.
43. Sim C, Denlinger DL: **A shut-down in expression of an insulin-like peptide, ILP-1, halts ovarian maturation during the overwintering diapause of the mosquito *Culex pipiens*.** *Insect Mol Biol* 2009, **18**:325-332.
44. Severson DW: **RFLP analysis of insect genomes.** In *The Molecular Biology of Insect Disease Vectors: a Methods Manual*. Edited by: Crampton JM, Beard CB, Louis C. London: Chapman and Hall; 1997:309-320.
45. Tekle A: **The physiology of hibernation and its role in the geographical distribution of populations of the *Culex pipiens* complex.** *Am J Trop Med Hyg* 1960, **9**:321-330.
46. Eldridge BF: **Environmental control of ovarian development in mosquitoes of the *Culex pipiens* complex.** *Science* 1966, **151**:826-828.
47. Eldridge BF: **The effect of temperature and photoperiod on blood-feeding and ovarian development in mosquitoes of the *Culex pipiens* complex.** *Am J Trop Med Hyg* 1968, **17**:133-140.
48. Mori A, Severson DW, Christensen BM: **Comparative linkage maps for the mosquitoes (*Culex pipiens* and *Aedes aegypti*) based on common RFLP loci.** *J Hered* 1999, **90**:160-164.
49. Mori A, Romero-Severson J, Severson DW: **Genetic basis for reproductive diapause is correlated with life history traits within the *Culex pipiens* complex.** *Insect Mol Biol* 2007, **16**:515-524.
50. Lawson D, Arensburg P, Atkinson P, Besansky NJ, Bruggner RV, Butler R, Campbell KS, Christophides GK, Christley S, Dialynas E, Emmert D, Hammond M, Hill CA, Kennedy RC, Lobo NF, MacCallum MR, Madey G, Megy K, Redmond S, Russo S, Severson DW, Stinson EO, Topalis P, Zdobnov EM, Birney E, Gelbart WM, Kafatos FC, Louis C, Collins FH: **VectorBase: a data resource for invertebrate vector genomics.** *Nucleic Acids Res* 2009, **37**:583-587.

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