

RESEARCH ARTICLE

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Emerging threat of multidrug resistant bugs – *Acinetobacter calcoaceticus baumannii* complex and Methicillin resistant *Staphylococcus aureus*

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Abstract

Background: Infections caused by bacteria such as multidrug resistant (MDR) *Acinetobacter* spp. and methicillin-resistant *Staphylococcus aureus* (MRSA) constitute a worldwide pandemic. Without gathering information about these strains, we cannot reduce the morbidity and mortality due to infections caused by these notorious bugs.

Methods: This study was conducted to identify the status of MDR *Acinetobacter* spp. and MRSA in a tertiary care centre of Nepal. Sputum, endotracheal aspirate and bronchial washing specimens were collected and processed from patients suspected of lower respiratory tract infection following standard microbiological methods recommended by the American Society for Microbiology (ASM). Double disk synergy test method was employed for the detection of extended-spectrum beta-lactamase (ESBL) in *Acinetobacter* isolates. Methicillin resistance in *S. aureus* was confirmed by using ceftioxin and oxacillin disks.

Results: Different genomespecies of *Acinetobacter* were isolated; these consisted of *Acinetobacter calcoaceticus baumannii* complex and *A. Iwoffii*. Around 95% of *Acinetobacter* isolates were MDR, while 12.9% were ESBL-producer. Of the total 33 isolates of *S. aureus*, 26 (78.8%) were MDR and 14 (42.4%) were methicillin resistant.

Conclusions: A large number of MDR *Acinetobacter* spp. and MRSA has been noted in this study. The condition is worsened by the emergence of ESBL producing *Acinetobacter* spp. Hence, judicious use of antimicrobials is mandatory in clinical settings. Moreover, there should be vigilant surveillance of resistant clones in laboratories.

Keywords: *Acinetobacter* spp., MDR, MRSA, ESBL

Background

Worldwide emergence of multidrug resistance among gram-negative and gram-positive bacteria has resulted in a confounding scene in treatment modalities. Bacteria like *Acinetobacter calcoaceticus baumannii* (Acb) complex and methicillin resistant *Staphylococcus aureus* (MRSA) have created a dilemma regarding the appropriate antibiotic therapy to use against them. Infections caused by these bugs are often difficult to treat. Further, these bacteria survive for a long time in hospital environment, with enhanced opportunities for transmission between patients

[1]. It is noteworthy that hospital-acquired pneumonia (HAP) developing ≥ 5 days after hospitalization (late-onset) is often caused by aerobic gram-negative bacilli (e.g., *Acinetobacter* spp.) or MRSA [2].

MRSA is one of the most important nosocomial pathogens worldwide, but recently it is increasingly identified as the etiological agent of infections acquired in community. Molecular epidemiological studies indicate that community associated (CA)-MRSA and healthcare associated (HA)-MRSA may have distinctive phenotypic and genetic features [3]. Traditionally, CA-MRSA are attributed with characteristics, such as smaller staphylococcal chromosomal cassette (SCC)*mec* cassettes – types IV and V – and a more restricted resistance pattern to antibiotics other than β -lactams than HA-MRSA. However,

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recently, a bidirectional crossing of borders between HA- and CA-associated infections is occurring [4]. These days, MRSA infections acquired outside of the hospital setting have been increasingly reported. The resistance of MRSA against various antimicrobials is globally increasing at an alarming rate. As a result, treatment of MRSA infections has become more challenging. This is a disturbing revelation and a major concern among health care professionals.

Studies of multidrug resistant (MDR) bacterial isolates like *Acb* complex and MRSA are crucial not only for the proper management of infections caused by them, but also for the prevention of the dissemination of such strains in the community and in hospitals. We undertook this study to find out the current trend of drug resistant *Acinetobacter* spp. and MRSA isolates in clinical samples from patients suspected of lower respiratory tract infections (LRTIs). Additionally, we aimed to determine the prevalence of ESBL in *Acinetobacter* and to compare the role of oxacillin and cefoxitin discs in detecting methicillin resistance in *S. aureus*.

Methods

A hospital-based prospective study was carried out in the Bacteriology Laboratory of Tribhuvan University Teaching Hospital (TUTH) in Kathmandu, Nepal over a period of six months. During the study period, 1120 specimens were processed.

Inclusion criteria

Specimens obtained from the lower respiratory tract [sputum, endotracheal (ET) secretion and bronchial washing] were included for culture and sensitivity test. Only those specimens which met the criteria recommended by the American Society for Microbiology (ASM) [5] were selected for further processing.

Culture of the specimen and identification of isolated organisms

The specimens were cultured on Chocolate agar (CHA), 5% Sheep Blood agar (BA) and MacConkey agar (MA) (Oxoid, UK) plates. The CHA plates were incubated in a CO₂ incubator (10% CO₂) at 37°C for 24 hours. The BA and MA plates were incubated at 37°C for 24 hours in an aerobic atmosphere. Identification of significant isolates was done following standard microbiological techniques [5]. A purity plate was employed to ensure that the inoculum used for the biochemical tests was pure.

Duplicate isolates from the same patient were excluded from the study.

Antibiotic susceptibility testing

The antibiotic sensitivity test of the pathogens isolated from the clinical specimen was done in Mueller Hinton

agar (MHA) (Oxoid, UK) by Kirby-Bauer method, as recommended by Clinical and Laboratory Standards Institute (CLSI) [6].

For disk susceptibility testing, penicillin, amoxicillin, oxacillin, ciprofloxacin, cotrimoxazole, cephalixin, gentamicin, erythromycin, clindamycin, teicoplanin, vancomycin, amoxicillin-clavulanate, cloxacillin, linezolid and cefoxitin were used for *S. aureus*. Likewise, ciprofloxacin, cotrimoxazole, gentamicin, amikacin, cefotaxime, ceftriaxone, ceftazidime, cefepime, amoxicillin-clavulanate, piperacillin, piperacillin-tazobactam, tigecycline, cefoperazone-sulbactam, meropenem and imipenem were used for *Acinetobacter* spp. The initial screen test for the production of ESBL was performed by using ceftriaxone (CRO) (30 µg), ceftazidime (CAZ) (30 µg) and cefotaxime (CTX) (30 µg) disks (Oxoid, UK). If the zone of inhibition (ZOI) was ≤ 25 mm for CRO, ≤ 22 mm for CAZ and/or ≤ 27 mm for CTX, the isolate was considered a potential ESBL-producer as recommended by CLSI [6]. Isolates those that were suspected as ESBL-producer by screen test were tested further by double disk synergy test (DDST). In DDST method, amoxicillin-clavulanic acid (AMC) disk (20/10 µg) was placed at the center and disks containing the 30 µg of CAZ, CTX and CRO were placed separately beside 15 mm distance (edge to edge), away from the central disk, in a horizontal manner [7]. Any enhancement of the ZOI between the disks (either of the cephalosporin disks and clavulanic acid containing disk) indicated the presence of ESBL.

In this study, if the isolates were resistant to at least three classes of first line antimicrobial agents, they were regarded as MDR [8]. Inducible macrolide-lincosamide-streptogramin B (iMLS_B) resistance was detected in *S. aureus* by Disk approximation test placing a 2 microgram clindamycin disk 15 mm away from the edge of a 15 microgram erythromycin disk on a MHA plate. Following incubation, organisms that showed flattening of the clindamycin zone adjacent to the erythromycin disk (referred to as a "D" zone) were considered to exhibit inducible clindamycin resistance [6].

To test for MRSA, a 1 µg oxacillin disk and a 30 µg cefoxitin disk were used, and the plates were incubated at 35°C. Plates containing oxacillin disk were read following a 24 hour incubation period. The diameter of the ZOI of growth was recorded and interpreted as susceptible or resistant by the criteria of CLSI. *S. aureus* isolates were deemed methicillin resistant when the ZOI was ≤ 10 mm with the oxacillin disk test or ≤ 21 mm with the cefoxitin disk method [6].

Control strains of *E. coli* ATCC 25922 and *S. aureus* ATCC 25923 were used for the standardization of the antibiotic sensitivity test. For ESBL test standardization, *E. coli* ATCC 25922 and *K. pneumoniae* ATCC 700603 were used as negative and positive controls respectively.

S. aureus strains ATCC 25923 and ATCC 43300 were used as negative and positive controls respectively for MRSA test standardization.

Ethical consideration

Permission was obtained from the Institutional Review Board at the Institute of Medicine, Tribhuvan University, Kathmandu. Written informed consent for participation in the study was obtained from participants or where participants were children, the consent was obtained from their parent or guardian.

Data processing and analysis

Data were analyzed using Microsoft Excel 2007 and represented as frequency distribution and percentage.

Results

Number of specimens and result pattern

Among the total processed specimens (n=1120), 497 showed significant growth (44.38%). Of all the isolates, *Acinetobacter* spp., the organism of our interest, was the fourth most common bacterial isolate (11.23%). *Acinetobacter* spp. comprised of both *Acb* complex and *A. Iwoffii*. Likewise, *S. aureus* was the seventh most common significant bacteria (6.19%) to cause LRTIs. A majority of *Acinetobacter* spp. (88.3%) and *S. aureus* (75.76%) were isolated from inpatients.

Antimicrobial susceptibility test result of *Acinetobacter* spp.

Nearly 58%, 38.7% and 35% of *Acinetobacter* isolates showed resistance to piperacillin-tazobactam, tigecycline

and imipenem respectively (Table 1). Approximately 95% of *Acinetobacter* isolates were MDR, while 12.9% were ESBL-producer (Data not shown in table).

Antimicrobial susceptibility test result of *S. aureus*

Most of the commonly used oral antibiotics were not effective against *S. aureus*. Only vancomycin, teicoplanin and linezolid were found to be the most effective antibiotics (Table 2).

MRSA and Inducible clindamycin resistance

Of the total 33 isolates of *S. aureus*, 14 (42.4%) were methicillin-resistant. Incidence of inducible clindamycin resistance was also seen (3.0%).

Number of MDR bacterial isolates

Nearly 95% of *Acinetobacter* spp. and 78.8% of *S. aureus* isolates were MDR.

Distribution of *Acinetobacter* and MRSA

Acinetobacter spp. and MRSA were more common in intensive care units (ICUs) (Table 3).

Comparison of Cefoxitin and Oxacillin methods for MRSA detection

Among the 33 *S. aureus* isolates subjected to the cefoxitin (30 µg) and oxacillin (1 µg) disk methods, the former detected 14 (42.4%) to be MRSA and the latter detected 12 (36.4%). The cefoxitin disk method detected more MRSA ($P>0.05$).

Table 1 Antimicrobial susceptibility test result of *Acinetobacter* spp. (n=62)

Antibiotics	Resistant (%)
Ciprofloxacin	64.5
Cotrimoxazole	75.8
Gentamicin	62.9
Amikacin	54.8
Ceftriaxone	87.1
Cefotaxime	88.7
Ceftazidime	82.3
Cefepime	85.5
Amoxicillin-Clavulanate	88.7
Piperacillin	77.4
Piperacillin-Tazobactam	58.0
Cefoperazone-Sulbactam	54.8
Meropenem	50.0
Imipenem	35.5
Tigecycline	38.7

Table 2 Antimicrobial susceptibility test result of *S. aureus* (n=33)

Antibiotics	Resistant (%)
Penicillin	93.9
Amoxicillin	90.9
Ciprofloxacin	69.7
Cotrimoxazole	72.7
Cephalexin	84.6
Gentamicin	54.5
Amoxycillin-clavulanate	69.7
Erythromycin	63.6
Clindamycin	45.5
Oxacillin	36.4
Cefoxitin	42.4
Cloxacillin	42.4
Vancomycin	0.0
Teicoplanin	0.0
Linezolid	0.0

Table 3 Wardwise distribution of ESBL- producing *Acinetobacter* spp. and MRSA

Wards	<i>Acinetobacter</i> spp.	MRSA
	No.	No.
Surgical	9	2
Medical	14	4
Cardiac care unit (CCU)	5	1
ICU	23	6

Discussion

This study was conducted among the patients attending TUTH, Kathmandu to find out the bacteriology of LRTIs with special reference to *Acinetobacter* spp. and MRSA.

A higher prevalence of *Acinetobacter* spp. was encountered in this study. *Acinetobacter* spp. consisted of *Acb* complex (10.85%) and *A. lwoffii* (0.38%). A prevalence study carried out a decade ago at TUTH showed the growth of *Acinetobacter* spp. to be 1.19% of the total number of cases [9]. Another study in the same setting in 2004 showed the increment in growth of these bacteria by twofold [10]. Likewise, the present finding on the prevalence of *Acinetobacter* spp. is higher than reported 3.9% of LRTI cases in Western Nepal [11]. These findings show an increasing trend in the prevalence of *Acinetobacter* spp.

In this study, out of total 62 *Acinetobacter* isolates, 51 (85.0%) were from inpatients and 48 (77.42%) were from elderly patients. The infection caused by *Acinetobacter* most frequently involved the respiratory tract of intubated patients. Studies have shown that *Acinetobacter* pneumonia is more common in critically ill patients in Asian (range 4-44%) and European hospitals (0-35%); however, it has a low incidence in United States hospitals (6-11%) [12].

Fluoroquinolones have replaced other quinolones over the last two decades. They are now widely used in clinical medicine as broad-spectrum antimicrobial agents. As a result, fluoroquinolone resistance is increasing rapidly [13]. In our study, ciprofloxacin resistance was found in 64.52% of *Acinetobacter* spp. isolates. Higher resistance (72.9% and >90%) to this antibiotic has been reported in India by Joshi et al. [13] and Mezzatesta et al. [14]. *Acinetobacter* spp. are known to harbour resistance plasmids (R-plasmids), and therefore often act as a reservoir of multidrug-resistance [15]. These pathogens can mutate to acquire quinolone resistance by either target alteration that modifies DNA gyrase (DNA topoisomerase II), or diminished drug accumulation by modifying porin system and efflux pump [16].

Out of 62 *Acinetobacter* isolates, 59 (95.16%) were MDR and 88.14% of the MDR *Acinetobacter* isolates were from inpatients. This startling rate of MDR *Acinetobacter* underscores the need for a cogent step in treatment options.

Higgins et al. [16] suggested that sulbactam had better activity over clavulanate and tazobactam and it might represent an alternative treatment option for infections due to the MDR *Acb* complex. Ling et al. [17] also reported a higher sensitivity of *Acb* complex against cefoperazone-sulbactam than piperacillin-tazobactam in Shanghai (91% vs. 29%) and Hong-Kong (95% vs. 81%). Complying with the previous studies, this study also showed cefoperazone-sulbactam had a comparatively lower resistance rate (54.84%) than piperacillin-tazobactam (58.07%) and amoxicillin-clavulanate (88.71%); however, the difference is not significant for the former. Tigecycline, a glycylicycline, was found to be a promising antibiotic. However, caution should be used in considering tigecycline treatment for *A. baumannii* infection in sites where drug levels may be suboptimal, such as the bloodstream. Development of resistance to tigecycline has been reported recently [18]. This demands prudent use of this novel antibiotic in treatment modalities.

The resistance of *Acinetobacter* spp. towards the carbapenems is much higher in this study (50% for meropenem and 35.48% for imipenem) as compared to different studies in Indian hospitals, viz., All India Institute of Medical Sciences (34.7% for meropenem and 27.2% for imipenem) [19], and St. John's Hospital (14.0% for meropenem) [20].

The decreased susceptibility of gram-negative isolates towards the third and fourth generation cephalosporins (26.79% to 28.57%) could be attributable to ESBL or AmpC beta-lactamase production. We found that 12.9% of *Acb* complex were ESBL-producer. However, we did not perform test for AmpC beta lactamase. Due to the increase in ESBL producing strains, there has been increased use of beta-lactam/ beta-lactamase inhibitor combinations, monobactams and carbapenems. However, the last few years have witnessed an increasing resistance to these drugs as well. In India, 22.2% and 17.3% of ESBL producers showed resistance to meropenem and imipenem respectively [19]. Though this reported finding is slightly higher than the present study finding, the possibility of the emergence of carbapenem-resistant isolates cannot be denied.

Medical wards and ICUs comprised the major domicile of ESBL producers. Third generation cephalosporin, such as ceftriaxone, cefotaxime and ceftazidime, are highly used in ICUs, even in our setting. Therefore, the resistance observed here may have appeared under the selective influence of the extensive usage of these antibiotics. In any nosocomial setting, carbapenems are used as the last resort for treatment of MDR *Acinetobacter* infections. However, over the last 15 years, acquired resistance to this life saving antimicrobial has been increasingly reported [21]. The pan-resistant gram-negative ESBL producing isolates were sensitive to polymyxin B and colistin sulphate. Because no fundamentally new anti-infective drugs are

currently available, it compelled us to re-evaluate the 'old drugs' and, fortunately, they proved to be as effective in this study as in other studies [22].

Apart from drug resistance in gram-negative bacteria, a worldwide increase in antibiotic resistance has been observed among gram-positive isolates. Today the concern of MRSA has reached the pinnacle. It is noteworthy that MRSA can cause both community- and hospital-acquired infections. However, there are only a few reports depicting the situation of MRSA among LRTI isolates in Nepal. In this study, out of total 33 *S. aureus* isolates, 14 (42.42%) were identified as MRSA. Nearly 93% of MRSA were isolated from inpatients ($P < 0.04$). Considering the specimenwise distribution, 78.59% of the MRSA were isolated from sputum specimens, while the rest were from ET secretion. In eastern Nepal, Kumari et al. found that 60% of *S. aureus* isolates from respiratory tract specimens were methicillin resistant. Though the finding of the present study showed less prevalence of MRSA as compared to Kumari et al. [23], it is much higher than that of 13.01% reported by Pokhrel et al. [24] from TUTH in 1993 and 15.4% by Subedi et al. [25] from Western Nepal in 2005.

Prior antibiotic use is the most common risk factor for colonization and infection with MRSA. Other risk factors for pneumonia due to MRSA include use of corticosteroids and chronic obstructive pulmonary disease. Mortality rates are higher in patients with pneumonia caused by MRSA than in those with pneumonia caused by methicillin susceptible *S. aureus* [26]. This heightened mortality likely reflects more serious comorbidities rather than differences in the virulence of the organisms [27]. In Japan, MRSA isolates from sputum specimens dramatically increased from 0% in 1985 to 51.4% in 1988. Moreover, 7 out of 13 cases of bronchopulmonary infections due to MRSA died [28]. The SENTRY antimicrobial surveillance (1997–1999) program found that the prevalence of MRSA in hospitals was very high in Singapore (62.9%), Taiwan (61.1%) and Portugal (54.4%) [29].

This study demonstrated that the overall sensitivity pattern of *S. aureus* isolates, with the exception to clindamycin, linezolid, teicoplanin, and vancomycin, other commonly prescribed antibiotics, had their spectrum of action much below 70%. Nearly 54.54% of the isolates were sensitive to clindamycin and 100% to linezolid, teicoplanin and vancomycin. Agents like erythromycin, ciprofloxacin and cotrimoxazole were effective in only 36.36%, 30.30%, and 27.27% cases, respectively. Resistance to penicillin was found in 93.94% of the *S. aureus* isolates. This finding complies with that of Dar et al. [30], who showed 92.0% of the *S. aureus* were resistant to penicillin.

In this study, a very low sensitivity of MRSA strains towards ciprofloxacin is probably due to the indiscriminate empirical use of these drugs because of their low cost

and ease of availability in our country. High resistance to ciprofloxacin has also been observed by Subedi et al. [25]. The susceptibility towards erythromycin was also lower in this study when compared with that of Kumari et al. (28.69%) [23]. Clindamycin is one of the drugs of choice in MRSA infections. Macrolide-resistant isolates of *S. aureus* may have constitutive or inducible resistance to clindamycin (methylation of the 23S rRNA encoded by the *erm* gene also referred to as MLS_B i.e., Macrolide, Lincosamide and type B Streptogramin resistance) or may be resistant only to macrolides (efflux mechanism encoded by the *msrA* gene) [6]. In this study, one erythromycin inducible clindamycin resistant case in an MRSA strain was also observed by D test. Vancomycin is used to treat MDR infections. In this study, no isolate was found to be vancomycin resistant. Vancomycin seemed to be the only option as the last resort antibiotic for MDR staphylococcal infection.

Two different methods were employed for the detection of MRSA. The cefoxitin disk method was better than the oxacillin disk method ($P > 0.05$), as the former detected all 14 MRSA cases while the latter missed two cases. According to CLSI [6], the cefoxitin disk test is comparable to the oxacillin disk test for the prediction of *mecA*-mediated resistance to oxacillin. The cefoxitin disk test is easier to read and thus is the preferred method. Besides, cefoxitin is an inducer of the *mecA* gene.

Conclusions

Notorious bugs such as MDR *Acinetobacter* spp. and MRSA have emerged. Although combination drugs and carbapenems are relatively more effective against *Acinetobacter* spp., there is a confounding scene in antibiotic armamentarium due to reports on resistance of these bugs against the last resort antibiotics. Thus, there is a great threat of dissemination of such resistant clones in the community as well as in hospital settings. It is mandatory to control this situation before it leads to a disaster. Hospital infection control program should be effectively implemented for the management of infected patients and to breach the transmission of such organisms to other patients, health care workers, and to the community.

Abbreviations

MDR: Multidrug resistant; MRSA: Methicillin resistant *Staphylococcus aureus*; ASM: American Society for Microbiology; ESBL: Extended-spectrum beta-lactamase; HAP: Hospital-acquired pneumonia; LRTI: Lower respiratory tract infection; TUTH: Tribhuvan University Teaching Hospital; ET: Endotracheal; MHA: Mueller Hinton agar; CLSI: Clinical and Laboratory Standard Institute; CA-MRSA: Community associated Methicillin resistant *Staphylococcus aureus*; HA-MRSA: Healthcare associated Methicillin resistant *Staphylococcus aureus*; $iMLS_B$: Inducible macrolide lincosamide and streptogramin B; DDST: Double disk synergy test; ZOI: Zone of inhibition; ATCC: American type culture collection; Fig.: Figure; No.: Number; ICU: Intensive care unit; CCU: Cardiac care unit.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

SKM carried out the study, analyzed data and prepared the manuscript. BPR and BMP designed the study and supervised the work and the manuscript. All authors have read and approved the manuscript.

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