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The pulsed ultrasound strategy effectively decreases the *S. aureus* population of chronic rhinosinusitis patients



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Abstract

Objective: Staphylococcus aureus with the ability of biofilm formation and the drug resistance acquisition is one of the most frequently isolated pathogens from chronic rhinosinusitis patients. Ultrasound as an alternative therapy is effectively able to kill the bacteria by cavitation in or on the bacterial cells and peroxide generation and hence improving antibiotic treatment efficacy.

Results: Staphylococcus aureus was detected in 4 and 6 out of 14 patients by phenotypic and qPCR assays, respectively. Four patients were completely resolved after pulsed ultrasound treatment. However, presence of the *S. aureus* was confirmed in three healthy controls by bacterial cultivation. Pulsed ultrasound have been quantitatively decreased the *S. aureus* population in chronic rhinosinusitis patients (p < 0.05). Further studies need to be investigated the effectiveness of pulsed ultrasound as an alternative course of CRS patient's treatment.

Keywords: Pulsed ultrasound, Chronic rhinosinusitis, *Staphylococcus aureus*, Treatment, Real-time PCR

Introduction

Bacterial communities within a polymeric matrix which called biofilms are difficult for cultivation and highly persistent during antibiotic treatment [1]. Approximately, 65% of human bacterial infections are accompanied with biofilms including sinusitis [2]. Chronic rhinosinusitis (CRS) is a prevalent disease and have been documented with bacterial biofilms complicated to control. *Staphylococcus aureus* with the ability of biofilm formation and the drug resistance acquisition is one of the most frequently isolated pathogens from CRS patients [3]. Because of the complex and not well-known nature of the rhinosinusitis and drug resistance development of involved pathogens, treatment of the

disease is sophisticated [4]. Medical management of the CRS is difficult and needs a prolonged antibiotic therapy. In addition to the complications of long-term medical treatment, persistence promotion and emerging the drug resistant bacterial populations made investigators to explore alternative therapies [4, 5].

Therapeutic ultrasound (US) has been advocated as a treatment option for CRS patients in recent years [1, 4]. The high-frequency sound waves are used in ultrasound treatment and can be implemented as continuous or pulsed type. Ultrasound is effectively able to kill the bacteria by cavitation in or on the bacterial cells and peroxide generation and hence improving antibiotic treatment efficacy [1]. It has been demonstrated that ultrasound could completely treat CRS patients [6]. Therefore, the aim of this study was to survey the impact of pulsed ultrasound on the *S. aureus* population of chronic rhinosinusitis patients.

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Main text

Materials and methods

Study setting

This study was a double blind, randomized clinical trial performed from April to September 2018. The included individuals were CRS infected adult patients confirmed by clinical diagnostic criteria (more than or equal to two major symptoms or one major symptom (nasal obstruction, facial pain/pressure, postnasal drip and hyposmia) and two minor symptoms (headache, halitosis, fatigue, dental pain and ear pain)) and also the CT scan results based on the Lund-Mackay staging system [7]. Exclusion criteria were having any head and neck malignancy. Totally, fourteen CRS patients were referred by an Ear, Nose, and Throat (ENT) surgeon. Additionally, ten voluntarily healthy people were enrolled in this study. The informed consent form was filled before entering the study.

Specimen collection

Specimen collection from all participants were carried out in two times within a 21 days interval (before and after of US exposure to CRS patients, first and second time without any therapy for healthy control group). Briefly, Dacron swabs was used to collect specimens from three sites, including nasopharynx, the right and left meatus and kept in sterile tubes, containing 1 ml saline solution. Following sampling, all specimens were transferred to clinical laboratory for further analysis. Also, CT scan was done for all precipitants in two times, as described above and SNOT-20 questionnaire (Additional file 1) was filled out.

Therapeutic ultrasound

Patients were subjected to 10 times pulsed US therapy with 4 cm/s of the probe velocity movement. Ultrasound frequency was 1 MHz. The intensity for maxillary and frontal sinuses were reported 1 W/cm² and 0.5 W/cm², respectively. Duty cycle for ultrasound was 10%. Specimen collection, CT scans and SNOT20 questionnaire was repeated following the last exposure of pulsed US therapy. If recovery was not seen by CT scan findings or patient didn't report recovery, routinely medical therapy started for patients by ENT specialist.

Phenotypic identification by conventional methods

All the specimens were vortexed carefully and worked out as soon as possible after collection. Ten microliters of each specimen were subjected to serial dilution from 10^1 to 10^{-6} concentration in PBS. By spreading plate technique, 10 microliters of each diluted suspensions were cultured separately in 2 sheep Blood agar, 2 chocolate agar and

MacConkey agar plates and were incubated aerobically and anaerobically at 37 °C for 24 h. After the incubation period, the colony-forming units (CFUs)/ml of each specimen was analyzed using Miles and Misra Method [8]. Also, the Gram staining, standard routine biochemical and microbiological tests were used to identify all the bacterial colonies [9].

DNA extraction

Total Bacterial DNA was extracted from clinical specimens using Macherey–Nagel (MN) DNA purification kit according to the manufacturer's instruction (Macherey–Nagel, Germany). Finally, extracted DNA was stored in $-20\,^{\circ}\mathrm{C}$ for further analysis. *S. aureus* ATCC 29213 strain was used as standard strain.

Quantitative real time PCR

The primers and probe were used to detect fibronectin-binding protein A (FnbA) gene of *S. aureus* are as follows: forward primer 5-AGTGAGCGACCATACAAC AG-3, reverse primer 5-CATAATTCCCGTGACCAT TT-3 and probe 5-FAM-AAGCACAAGGACCAATCG AGG-BHQ-1-3 [10]. First, by using the Nanodrop 1000 (Thermo Scientific, USA), the amounts of DNA templates were checked. For calculating the DNA molecules of each template per gram, following formula was used; molecules of DNA = mass (in gram) Avogadro's number/average molecular weight of a base × template length [11]. A standard curve was drawn using tenfold serially diluted DNA to gain the efficacy of the PCR reactions. The qPCR assay was performed by LinGene K Real Time

Table 1 The patient's scores from CT and SNOT-20 questionnaire before and after the treatment

Patient ID	CT scores		Questionnaire scores	
	Before	After	Before	After
1	8	1	56	10
2	17	0	38	8
3	15	5	48	25
4	7	0	24	1
5	27	4	36	9
6	6	1	17	2
7	18	4	17	1
8	24	6	40	25
9	33	11	85	25
10	11	2	45	15
11	6	0	64	1
12	17	2	48	6
13	24	0	54	0
14	17	2	62	10

The CT score and questionnaire scores were decreased after treatment showing patient's remission

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PCR apparatus (Bioer, Hangzhou, China). The control strain was *S. aureus* ATCC 29213 and all the samples were run in duplicate.

Statistical analysis

All data were analyzed by SPSS software version 21. The presence of *S. aureus* among patients and control groups were analyzed by Chi Square test. *S. aureus* detected in samples before and after the therapy by conventional culture technique were analyzed by nonparametric McNemar's test. For qPCR results between related groups before and after the treatment, Wilcoxon test was used. A P value of < 0.05 was considered as statistically significant.

Results

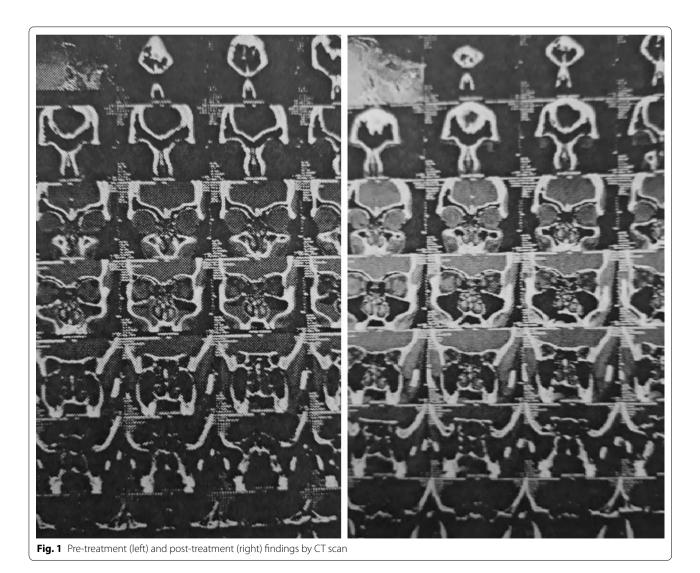
Bacterial isolates

According to conventional bacterial identification, *S. aureus* was detected in both CRS patients and healthy

group before treatment. In this case, *S. aureus* was detected in 3 and 4 cases in healthy and CRS patients, respectively. Importantly, although *S. aureus* prevalence among CRS patients and healthy group was not significant (p>0.05), but the presence of other pathogenic bacteria such as *Streptococcus pneumoniae*, *Streptococcus pyogenes*, *Haemophilus influenza*, *Klebsiella pneumoniae* and *Escherichia coli* was detected only in CRS patients.

Ultrasound treatment output

US therapy effectively resolved *S. aureus* in 3 cases and decreased bacterial CFU/ml in one case by phenotypic analysis. This decreased bacterial load was in concomitant with the site changing of *S. aureus* from the right meatus to the nasopharynx. Although this difference is considered to be not statistically significant (p>0.05), but is clinically significant. In addition, *S. pneumoniae*,



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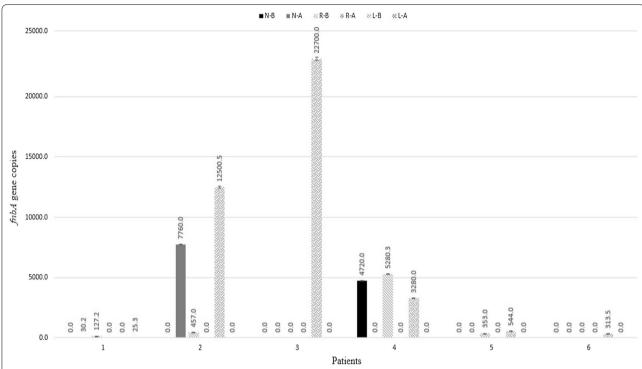


Fig. 2 qPCR results of *fnbA* gene in positive patients. (*N-B* nasopharynx-before treatment, *N-A* nasopharynx-after treatment, *R-B* right meatus-before treatment, *L-A* left meatus-after treatment).

S. pyogenes, H. influenza, K. pneumoniae and E. coli were removed from different sites of patients except one case that, decreasing of the Streptococcus pneumoniae load was seen phenotypically. The CT score of all the patients was decreased considerably after the treatment. As well as, the patient's scores from the SNOT-20 questionnaire were less after treatment compared to the initial stage. All these findings are in favor of patient's remission due to pulsed ultrasound treatment (Table 1, Fig. 1).

Quantitative PCR findings

Six patients were quantitatively demonstrated to have S. aureus. From these, 4 patients were completely recovered after treatment and the bacterial load was significantly reduced (p=0.02). However, two patients (No. 1, 2) that were negative at the beginning showed S. aureus after treatment. This may be because of bacterial transferring during sampling or subsequent colonization of bacteria. The qPCR results are detailed in Fig. 2.

Discussion

Treatment of CRS patients is encountering to the problem and is challengeable. Mucosal surfaces of the sinuses of these patients is source of multi species of bacterial population within the extracellular matrix which called biofilm [10]. This matrix is elaborated by bacterial population and it causes tolerance and resistance against harsh conditions such as the presence of antimicrobial agents [12]. Already, *S. aureus* is considered as an important infectious agent which can produce the thick layer of biofilm [13, 14].

Despite classical antibiotic therapy in CRS patients commonly is led to failure in therapy, but antibiotics are used as an empiric choice for CRS patients [15, 16] which resulting development of resistant bacteria and thus treatment failure [16]. It has also been shown, exposure of S. aureus living in the sinuses to antibiotics led emerging and distributing of methicillin resistant S. aureus (MRSA) in CRS patients [16]. Complication of CRS patients by the noticed challenges convinced researchers to study and elaborate alternative strategy for treatment of CRS patients [17]. In this regard, US therapy is a suggested strategy for treatment of CRS patients by breaking down the biofilm structure of the bacterial population. It has been reported that, US can decrease bacterial load through biofilm destruction [18]. In pulsed US strategy, it reduces thermal activities by allowing time for heat to dissipate from the coupling medium during treatment [19]. Although, biofilm formation of bacterial strains and its disruption by therapy was not evaluated in our study, but according to previous studies and published paper (see above), it has been demonstrated that,

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S. aureus is considered as an important CRS agent which can produce the thick layer of biofilm. In another hand, according to literature, US therapy was using as biofilm disruption option [13–18].

It has also been shown, Pulsed US therapy combined with gentamicin decrease bacterial viability in E. coli model [1, 20]. A study conducted on pulsed US therapy against P. aeruginosa, E. coli in implants [21]. These researchers reported that, pulsed US therapy only was effective against E. coli. It shows pulsed US therapy is more effective when combined with the antibiotics, which can decrease bacterial viability dramatically [22]. Ansari et al. [23] treated 57 CRS cases using low-intensity pulsed US successfully. According to their reports, most major and minor symptoms indicated significant changes after pulsed US therapy. Young et al. [24] treated 22 CRS cases with six treatments of low-intensity pulsed US which who were being considered for endoscopic sinus surgery. In accordance to these studies, our results derived from bacterial culture and quantitative molecular tests showed a significant reduction in S. aureus load in majority of patients after pulsed US treatment.

In conclusion, the clinical improvements of patients showed beneficial recovery and significantly reduced bacterial load were seen in target patients. However, the effectiveness of pulsed ultrasound is thought to be worthy of investigation in further studies.

Limitations

The low number of volunteers to participate in the study and difficulty to access and following up of all members are limitation of this study.

Supplementary information

Supplementary information accompanies this paper at https://doi.org/10.1186/s13104-019-4579-3.

Additional file 1. SINO-NASAL OUTCOME TEST (SNOT-20).

Abbreviations

CRS: chronic rhinosinusitis; US: ultrasound; CT scan: computed tomography scan; ENT surgeon: nose, and throat surgeon; SNOT: Sino-Nasal Outcome test; CFUs: colony-forming units; FnbA: fibronectin-binding protein A; qPCR: quantitative real time PCR.

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Authors' contributions

JS and MMF: conceived, designed and supervised the study; MF revised the manuscript; SHS, JKY: specimen collection; NF: US treatment; HK: Microbiological analysis; NF, BVG, MD: collected and analyzed the data; NF: drafted the manuscript. All authors read and approved the final manuscript.

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Availability of data and materials

The datasets used and/or analyzed during the current study are presented within the manuscript, supplementary file and also additional information such as patients' history are available from the corresponding author on reasonable request.

Ethics approval and consent to participate

The study protocol and ethical issue were approved by the Ethics Committee of Iran University of Medical sciences (IRCT20140810018754N8).

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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References

- Bartley J, Young D. Ultrasound as a treatment for chronic rhinosinusitis. Med Hypotheses. 2009;73(1):15–7.
- Costerton JW, Stewart PS. Biofilms and device-related infections. Persistent bacterial infections. Washington, D.C.: American Society of Microbiology; 2000. p. 423–39.
- Dworniczek E, Frączek M, Seniuk A, Kassner J, Sobieszczańska B, Adamski J, et al. Bacterial biofilms in patients with chronic rhinosinusitis. Folia Microbiol. 2009;54(6):559–62.
- Nakhostin Ansari N, Naghdi S, Farhadi M. Physiotherapy for chronic rhinosinusitis: the use of continuous ultrasound. Int J Ther Rehabil. 2007;14(7):306–10.
- Ansari NN, Fathali M, Naghdi S, Hasson S. Effect of pulsed ultrasound on chronic rhinosinusitis: a case report. Physiother Theory Pract. 2010;26(8):558–63.
- Ansari NN, Naghdi S, Farhadi M. Therapeutic ultrasound as a treatment for chronic sinusitis. Physiother Res Int. 2004;9(3):144–6.
- Lund VJ, Mackay IS. Staging in rhinosinusitus. Rhinology. 1993;31(4):183–4.
- 8. Miles AA, Misra S, Irwin J. The estimation of the bactericidal power of the blood. Epidemiol Infect. 1938;38(6):732–49.
- Mahon CR, Lehman DC, Manuselis G. Textbook of diagnostic microbiology. Amsterdam: Elsevier Health Sciences; 2014.
- Ghodousi A, Nomanpour B, Davoudi S, Maleknejad P, Omrani M, Kashef N, et al. Application of fnbA gene as new target for the species-specific and quantitative detection of Staphylococcus aureus directly from lower respiratory tract specimens by real time PCR. Indian J Pathol Microbiol. 2012;55(4):490.
- Adams PS. Data analysis and reporting. Real-time PCR. Abingdon: Taylor & Francis: 2007. p. 65–88.
- Kazemian H, Bourbour S, Beheshti M, Bahador A. Oral colonization by nosocomial pathogens during hospitalization in intensive care unit and prevention strategies. Recent Pat Anti-Infect Drug Discov. 2017;12(1):8–20.

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- Mahdiyoun SM, Kazemian H, Ahanjan M, Houri H, Goudarzi M. Frequency of aminoglycoside-resistance genes in methicillin-resistant staphylococcus aureus (MRSA) isolates from hospitalized patients. Jundishapur J Microbiol. 2016;9(8):e35052.
- Ghafourian S, Mohebi R, Rezaei M, Raftari M, Sekawi Z, Kazemian H, et al. Comparative analysis of biofilm development among MRSA and MSSA strains. Roum Arch Microbiol Immunol. 2012;71(4):175–82.
- Smith SS, Evans CT, Tan BK, Chandra RK, Smith SB, Kern RC. National burden of antibiotic use for adult rhinosinusitis. J Allergy Clin Immunol. 2013;132(5):1230–2.
- Bhattacharyya N, Kepnes LJ. Assessment of trends in antimicrobial resistance in chronic rhinosinusitis. Ann Otol Rhinol Laryngol. 2008;117(6):448–52.
- Bordin A, Sidjabat HE, Cottrell K, Cervin A. Chronic rhinosinusitis: a microbiome in dysbiosis and the search for alternative treatment options. Microbiol Austr. 2016;37(3):149–52.
- 18. Bartley J, Ansari NN, Naghdi S. Therapeutic ultrasound as a treatment modality for chronic rhinosinusitis. Curr Infect Dis Rep. 2014;16(3):398.
- Polat BE, Hart D, Langer R, Blankschtein D. Ultrasound-mediated transdermal drug delivery: mechanisms, scope, and emerging trends. J Control Release. 2011;152(3):330–48.
- 20. Rediske AM, Roeder BL, Nelson JL, Robison RL, Schaalje GB, Robison RA, et al. Pulsed ultrasound enhances the killing of *Escherichia coli* biofilms

- by aminoglycoside antibiotics in vivo. Antimicrob Agents Chemother. 2000;44(3):771–2.
- 21. Carmen JC, Roeder BL, Nelson JL, Ogilvie RLR, Robison RA, Schaalje GB, et al. Treatment of biofilm infections on implants with low-frequency ultrasound and antibiotics. Am J Infect Control. 2005;33(2):78–82.
- Ensing G, Roeder B, Nelson JLE, Van Horn J, Van der Mei H, Busscher H, et al. Effect of pulsed ultrasound in combination with gentamicin on bacterial viability in biofilms on bone cements in vivo. J Appl Microbiol. 2005;99(3):443–8.
- 23. Ansari NN, Naghdi S, Farhadi M, Jalaie S. A preliminary study into the effect of low-intensity pulsed ultrasound on chronic maxillary and frontal sinusitis. Physiother Theory Pract. 2007;23(4):211–8.
- Young D, Morton R, Bartley J. Therapeutic ultrasound as treatment for chronic rhinosinusitis: preliminary observations. J Laryngol Otol. 2010;124(5):495–9.

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