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Genetic variation of *ABCB1* (rs1128503, rs1045642) and *CYP2E1* rs3813867 with the duration of tuberculosis therapy: a pilot study among tuberculosis patients in Indonesia

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Abstract

Objective: The risk of contracting tuberculosis (TB) and the efficacy of TB therapy are affected by several factors, including genetic variation among populations. In the Indonesian population, data on the genes involved in drug transport and metabolism of TB therapy are limited. The aim of this study was to identify the genetic profile of the *ABCB1* gene (rs1128503 and rs1045642) and *CYP2E1* gene (rs3813867) in Indonesians with TB. This study was a cross-sectional study of 50 TB outpatients in Jambi city, Indonesia. Sociodemographic characteristics were obtained from medical records. Whole blood was collected, and genomic DNA was isolated. Single nucleotide polymorphisms were determined using polymerase chain reaction-restriction fragment length polymorphism with *Haelll*, *Mbol*, and *Pstl* for rs1128503, rs1045642 (*ABCB1*), and rs3813867 (*CYP2E1*), respectively.

Result: The frequency of alleles of each gene was analyzed by Hardy–Weinberg equilibrium. The genetic profiles of *ABCB1* rs1128503 and rs1045642 were varied (CC, CT, TT), while *CYP2E1* rs3813867 was present in CC (wild type). The genetic variations of *ABCB1* and *CYP2E1* may have no significant correlation with the duration of TB therapy. Nevertheless, this study may provide as preliminary results for the genetic profiles of *ABCB1* (rs1128503, rs1045642) and *CYP2E1* (rs3813867) in the Indonesia population.

Keywords: ABCB1 C1236T, ABCB1 C3435T, CYP2E1-1293G>C, Single nucleotide polymorphism, Risk factor

Introduction

Tuberculosis (TB) is an infectious disease with a high mortality rate. TB is caused by *Mycobacterium tuberculosis* (*M. tuberculosis*) that mainly invades the lung. Indonesia has a pulmonary TB prevalence of 0.42% (1,017,290 cases), and in 2018, the incidence rate was 316 cases

per 100,000 population, making the country third highest in the world for TB [1, 2]. Moreover, TB resistant to rifampicin or multiple drug resistant tuberculosis (MDR-TB) is a serious threat. In 2018, there were 24,000 cases of MDR-TB in Indonesia [2].

First-line TB therapy is an adequate strategy for TB-sensitive cases; a combination of rifampicin, isoniazid, pyrazinamide, and ethambutol (2-month intensive phase) is continued with rifampicin and isoniazid for 4 months. These regimens are strongly recommended for countries with a high incidence of MDR-TB [3–5]. Strategies of

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Barliana et al. BMC Res Notes (2021) 14:295 Page 2 of 7

TB eradication focus on monitoring therapy and comprehensive patient care; however, the efficacy of therapy still presents challenges, especially in developing countries [6, 7]. Current TB therapy achieves > 95% cure and <5% relapse rates, but a small proportion of patients are not responsive to the therapy [8]. Factors affecting the efficacy of TB therapy include the host and bacteria. Genetic variations among individuals are known to affect the efficacy and toxicity of therapy [9]. Single nucleotide polymorphisms (SNPs) of genes involved in the metabolism or uptake of TB drugs show correlations with efficacy, such as the cytochrome P-450 (CYP) family and adenosine triphosphate (ATP)-binding cassette (ABC) family [10-12]. Alterations in these genes may influence the pharmacokinetics, sensitivity, or adverse reactions to drugs [13].

Adenosine triphosphate (ATP)-binding cassette B1 (ABCB1) is a membrane transporter, which is encoded by the ABCB1 gene, plays an important role in ATPdependent uptake and efflux of extracellular compounds and xenobiotics into and from cells [10, 14, 15]. Therefore, polymorphisms of ABCB1 determine the risk factor, efficacy, and toxicity of some therapies. ABCB1 gene C1236T (rs1128503) and/or C3435T (rs1045642) are involved in diseases such as TB, cancer, minor ischemic stroke, chronic liver disease, and mental health [14, 16-24]. The adverse effect of TB therapy is significantly correlated with hepatotoxicity [25-27]. Cytochrome P-450 2E1 (CYP2E1) encoded by the CYP2E1 gene, is mainly expressed in the liver, and catalyze xenobiotic metabolism. CYP2E1 is involved in isoniazid metabolism, and its activity and expression are affected by polymorphisms of CYP2E1 gene in the 5' upstream region (-1053C>T) [9, 28-32].

It is important to understand whether genetic variation is one of the risk factors for TB and severity progression. The present study was the first study to identify the genetic profile of the *ABCB1* gene (rs1128503 and rs1045642) and *CYP2E1* gene (rs3813867) of TB patients in Jambi city, Indonesia.

Main text

Methods

Subjects

This study was a cross-sectional study of TB outpatients in Abdul Manap Hospital, Jambi city, Indonesia. Fifty patients were examined to observe the SNPs of *ABCB1* (rs1128503 and rs1045642) and *CYP2E1* (rs3813867). The sample collection was conducted for 3 months. The population (male and female) that met the inclusion criteria, such as patients diagnosed with TB and currently undergoing TB therapy, was selected. The sociodemographic characteristics (age, gender, education, and occupation),

duration of therapy, therapy category, smoking status, and alcohol consumption were obtained from medical records.

Genotyping of ABCB1 (rs1128503 and rs1045642) and CYP2E1 (rs3813867)

The gene sequences were obtained from The National Center for Biotechnology Information (https://www.ncbi. nlm.nih.gov). Whole blood was collected and genomic DNA was isolated using Purelink Genomic DNA Mini Kit (Thermo Fischer Scientific, Waltham, MA, USA). SNP identification was determined using the polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) method. PCR was conducted by PCR SuperMix (Thermo Fisher Scientific, Waltham, MA, USA) using specific primers for each SNP. All specific primer (Invitrogen, Thermo Fisher Scientific, Waltham, MA, USA) sequences were obtained from previous studies [18, 33]. Each specific primer was confirmed with GENETYX version 10 software and Oligo Calc: Oligonucleotide Properties Calculator (http://www.basic.north western.edu/biotools/oligocalc.html) (Additional file 1: Table S1). PCR products were digested with MboI, HaeIII, or PstI (Promega, Madison, WI, USA), electrophoresed using 2.5% agarose gel containing SYBR® Safe DNA Gel Stain (Thermo Fischer Scientific, Waltham, MA, USA), and visualized under ultraviolet light at 312 nm. The MboI-digested PCR fragment (ABCB1 rs1045642) produced 88- and 162-bp for CC (wild-type); 88-, 162-, and 250-bp for CT; 250-bp for TT. HaeIII digested PCR fragments (ABCB1 rs1128503) produced 270-, 65-, and 35-bp for CC (wild-type); 270-, 100-, 65-, and 35-bp for CT; 270- and 100-bp for TT. The PstI digested PCR fragment (CYP2E1 rs3813867) produced: 412-bp for CC (wild-type), 118-, 294-, and 412-bp for CT, and 294-, and 412-bp for TT. GAPDH gene expression was used as an internal PCR control in the same samples.

Statistical analysis

Each locus was analyzed for allele frequencies using descriptive statistics with Hardy–Weinberg equilibrium (HWE, df=1). Statistically significant differences in sociodemographic characteristics were analyzed using Student's t-test with p<0.05 considered as statistically significant. The correlation between sociodemographic characteristic and duration of therapy was analyzed by Chi-square or Fisher exact test univariate analysis.

Results

Sociodemographics of TB patients

TB patients in Jambi city were mostly males of productive age (40 years old of age or younger) (Table 1; Additional file 2: Figure S1 and Additional file 3: Figure S2).

Barliana et al. BMC Res Notes (2021) 14:295 Page 3 of 7

Table 1 Correlation of sociodemographic characteristics with duration of therapy (n = 50)

Risk factors	n (%)	Duration of therapy ≤ 6 months n (%)	Duration of therapy > 6 months n (%)	<i>p</i> value
Gender				1.000*
Male	35 (70.0)	26 (68.4)	9 (75.0)	
Female	15 (30.0)	12 (31.6)	3 (25.0)	
Age group, years (N)				=-
<u>≤</u> 29	9 (18.0)	9 (23.7)	0 (0)	
1	8 (16.0)	6 (15.8)	2 (16.7)	
40–49	13 (26.0)	11 (28.9)	2 (16.7)	
50-59	12 (24.0)	7 (18.4)	5 (41.7)	
≥ 60	8 (16.0)	5 (13.2)	3 (25.0)	
Education (N)				_
Primary school	4 (8.0)	3 (7.9)	1 (8.3)	
Junior high school	7 (14.0)	5 (13.2)	2 (16.7)	
Senior high school	37 (74.0)	28 (73.7)	9 (75.0)	
Diploma/bachelor degree	2 (4.0)	2 (5.3)	0 (0)	
Occupation (N)				=-
Students/university students	4 (8.0)	4 (10.5)	0 (0)	
Employed	27 (54.0)	21 (55.3)	6 (50.0)	
Unemployed	19 (38.0)	13 (34.3)	6 (50.0)	
Smoking status (N)				0.125#
No	34 (68.0)	28 (73.7)	6 (50.0)	
Yes	16 (32.0)	10 (26.3)	6 (50.0)	
Alcohol consumption				0.621*
No	44 (88.0)	34 (89.5)	10 (83.3)	
Yes	6 (12.0)	4 (10.5)	2 (16.7)	
Genotype ABCB1 rs1045642				0.668*
CC	9 (18.0)	6 (15.8)	3 (25.0)	
CT+TT	41 (82.0)	32 (84.2)	9 (75.0)	
Genotype ABCB1 rs1128503				0.386*
CC	8 (16.0)	7 (30.4)	1 (11.1)	
CT+TT	25 (50.0)	16 (69.6)	8 (88.9)	
Missing	17 (34.0)	_	=	
Genotype <i>CYP2E1</i> rs3813867				-
CC	50 (100)	12 (24)	38 (76)	-
CT+TT	0 (0)	=	-	

Statistical comparisons were performed using *Chi-square test and *Fisher exact test

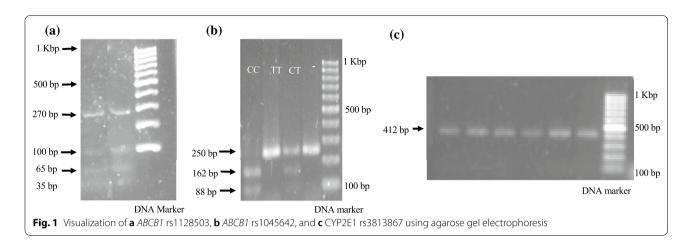
Most had graduated from senior high school and were either housewives or industrial laborers. Smoking and alcohol consumption status were only observed in only 16% and 6% of patients, respectively.

Genotyping of ABCB1 (rs1128503 and rs1045642) and CYP2E1 (rs3813867)

All samples were clearly identified by the RFLP method, but 17 samples were not identified for *ABCB1* rs1128503 (RFLP by *Hae*III) (Table 1, Fig. 1). Results showed that the genetic profiles of *ABCB1* rs1128503 were 7 (14%) CC, 14

(28%) CT, 12 (24%) TT, and 17 (34%) were not identified; ABCBI rs1045642 was 9 (18%) CC, 25 (50%) CT, and 16 (32%) TT. The T allele of both SNPs in ABCBI showed higher frequency than the C allele (Table 1). The CYP2EI rs3813867 was all observed in the CC (wild type) genotype (Additional file 4: Figure S3). Otherwise, the allele frequency of each genotype of all SNPs was not significant in the disequilibrium state, with p > 0.05 according to the Hardy–Weinberg equation (Additional file 5: Table S2; Additional file 6: Table S3; Additional file 7: Table S4).

Barliana et al. BMC Res Notes (2021) 14:295 Page 4 of 7



Duration of therapy and genetic variation

Most of patients were in first line therapy for TB (<6 months of therapy), but 1 patient was in the extensive phase, and 4 patients were confirmed to have MDR-TB (>6 months of therapy) (Table 1). All of the sociodemographic determinants were not significantly correlated with the duration of therapy (p>0.05). The genotype of ABCB1 (rs1045642 or rs1128503) showed variations rather than continuing for correlation analysis with duration of therapy. The variations in ABCB1 were divided into the C allele (CC, wild-type) and T allele (CT, TT) but showed no significant correlation with the duration of TB therapy (Table 1).

Discussion

Sociodemography characteristics are considered as risk factors for TB and MDR-TB incidence [34]. Our results showed that TB cases were higher in men, similar to most studies in Malaysia [35]. Active TB cases are shown to affect individuals of productive age, as shown in a Java island, Indonesia study [35–38]. However, we observed 17 patients who were less than 40 years of age and 33 who were more than 40 years old. Jambi city is located on Sumatera Island and might have different sociodemographic characteristics, including various ethnicities, from other parts of Indonesia.

Educational background is one TB determinants. A lower level of education correlated with a higher TB infection rate [35, 39]. Similarly, our data showed that 48 out of 50 patients had at least a senior high school educational. Lower educational background was assumed to have less exposure to health information; however this assumption has not been proven since TB-related health information can also be accessed by those with informal education [35, 40]. In the present study, most TB patients were employed. In Malaysia, employment status as a

determinant of TB infection was showed that patients who were unemployed completed TB therapy at higher rates [35]. Our result was similar to a study in Kenya [41]. Productive age and employed status of individuals are risk factors for TB infection because of high mobility and the increased likelihood of being exposed to TB [35, 42]. Behavioral factors, such as smoking status, alcohol consumption, and drug abuse, made individuals more susceptible to TB infection, thus affecting the incidence of active TB [43, 44]. We observed that only 16 patients smoked and 6 patients consumed alcohol. This might indicate that other factors affect the incidence of TB in Indonesia, especially in Jambi Province. Because the majority of Indonesians are Muslim, alcohol consumption is prohibited. Due to data limitations, we cannot statistically calculate the correlation of such sociodemographic characteristics with TB infection.

ABCB1 is involved in diseases and the efficacy of therapy, where SNPs in the ABCB1 gene affect its function. Polymorphisms of ABCB1 rs1128503 and rs1045642 are the most studied variants genetic of ABCB1 in diseases and indicate high frequencies in several populations [45, 46]. This was the first genotype variation study of ABCB1 in Indonesian TB patients, especially in the Jambi population, although we did not find a significant association with sociodemography characteristics or duration of therapy. We did find a higher number of T alleles in both polymorphisms of ABCB1. T alleles of ABCB1 rs1128503 are major alleles in Asia and minor alleles in Africa [14]. Both polymorphisms of ABCB1 rs1045642 and rs1128503 are synonymous SNPs, but they alter the stability of mRNA expression; therefore, they affect the drug pharmacokinetics, whether through reduced or increased drug bioavailability. An ABCB1 genotype study in Brazil showed that SNP rs1128503 had a significant correlation with the risk for MDR-TB. One of the limitations of our study was that we did not correlate the genetic variation with clinical outcomes or efficacy

Barliana et al. BMC Res Notes (2021) 14:295 Page 5 of 7

of therapy due to a lack of data. However, this study may become a preliminary study to identify the genetic profile of *ABCB1* rs1128503 (C1236T) and rs1045642 (C3435T) in Indonesia, especially in Jambi Province.

In the present study, all samples had the wild-type (CC) genotype; therefore we could not analyze the correlation with sociodemographic characteristics and duration of therapy. The variation in the CYP2E1 rs3813867 genotype in Malaysia (Asian and non-Asian) and in Turkey showed similar results to our study [47, 48]. The activity of CYP2E1 was isoniazid level-dependent and involved in acetyl hydrazine oxidation into diacetyl hydrazine and acetyl diazene ketene which was hepatotoxic [49]. CYP2E1 variation affects the efficacy of therapy, especially in adverse events causing anti-TB drug-induced hepatotoxicity (ATDH) [32]. Wild-type of CYP2E1 rs3813867 (c1/c1) was found to have increased activity compared to other variants of genotypes [9, 49]. These differences in the clinical outcomes of CYP2E1 rs3813867 have been studied in several populations. In Turkey, the heterozygosity of CYP2E1 rs3813867 was observed to increase the risk of ATDH, while in the North Indian population, the wild-type had a lower risk [25, 48]. As in China, most of Uyghur genetic variations were c1/c1 but not significantly associated with ATDH [50]. In our study, we found correlation between the genetic variation and ATDH. However, it is a potential issue for further investigation.

Conclusions

The ABCB1 and CYP2E1 genetic variations may have no significant correlation (p > 0.05) with the duration of TB therapy, although variations was occurred in ABCB1, due to small sample size. The result of the present work may provide as preliminary data on ABCB1 (rs1128503, rs1045642) and CYP2E1 (rs3813867) genetic profiles in Indonesian populations.

Limitations

- ABCB1 (rs1128503, rs1045642) and CYP2E1 (rs3813867) were considered responsible for drug efflux or metabolism, but due to the small number of patients, we could not find the significant involvement of these SNPs.
- We did not use a positive control for RFLP, although we used the GAPDH primer as an internal control for PCR.
- The visualization of PCR fragments was low quality due to low of sample concentrations; however, specific bands of correct size were observed.

Abbreviations

ABCB1: Adenosine triphosphate (ATP)-binding cassette B1; ATDH: Anti-TB drug-induced hepatotoxicity; CYP2E1: Cytochrome P-450 2E1; HWE: Hardy-Weinberg equilibrium; MDR-TB: Multiple drug resistant tuberculosis; *M. tuberculosis: Mycobacterium tuberculosis;* SNPs: Single nucleotide polymorphisms; PCR-RFLP: Polymerase chain reaction-restriction fragment length polymorphism; TB: Tuberculosis.

Supplementary Information

The online version contains supplementary material available at https://doi.org/10.1186/s13104-021-05711-8.

Additional file 1: Table S1. Primers for SNP genotyping [18, 33].

Additional file 2: Figure S1. Gender of TB patients.

Additional file 3: Figure S2. Age range of TB patients.

Additional file 4: Figure S3. Genotype distribution of a) *ABCB1* rs1128503, b) *ABCB1* rs1045642, and c) *CYP2E1* rs3813867.

Additional file 5: Table S2. Hardy–Weinberg Equilibrium for the Genotype of *ABCB1* rs1128503.

Additional file 6: Table S3. Hardy–Weinberg Equilibrium for the Genotype of *ABCB1* rs1045642.

Additional file 7: Table S4. Hardy–Weinberg Equilibrium for the Genotype of *CYP2E1* rs3813867.

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Authors' contributions

MIB and RA are designing the work, MIB is writing the manuscript, compiling and interpretation of data; ASWK, WNI, HH are collecting samples and compiling the data; SDA is analysis the data; AD, MM, TR, IMP, and AAS are substantively revised the draft. All authors read and approved the final manuscript.

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Availability of data and materials

All data generated or analysed during this study are included in this published article and available in the www.figshare.com repository [https://figshare.com/articles/dataset/Genotype_of_Samples_for_ABCB1_and_CYP2E1_pdf/14680884].

Declarations

Ethics approval and consent to participate

All patients were anonymized and informed about the research, then agreed and wrote informed consent to participate. All procedures performed in studies involving human participants comply with the ethical standards of the institutional and/or national research committee, Health Research Ethics Committee of Universitas Padjadjaran (No. 927/UN6.C.10/KEPK/PN/2017). The present research was also conducted in accordance with the 1964 Helsinki declaration and its later amendments or comparable ethical standards.

Consent for publication

Not applicable.

Competing interests

All authors declare that they have no competing interest in this work.

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Barliana et al. BMC Res Notes (2021) 14:295 Page 6 of 7

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References

- Ministry of Health Republic of Indonesia. Basic health research 2018. Jakarta: 2019. p. 1–580.
- 2. World Health Organization. Global tuberculosis report country profile 2019. Geneva: World Health Organization; 2019.
- World Health Organization. Guidelines for treatment of drug-susceptible tuberculosis and patient care. Geneva: World Health Organization; 2017.
- World Health Organization. Treatment of tuberculosis: guidelines. 4th ed. Geneva: World Health Organization; 2010.
- Ministri of Health Republic of Indonesia. Regulation of The Ministry of Health of The Republic of Indonesia No. 67 2016 of tuberculosis eradication. Jakarta: Ministry of Health Republic of Indonesia; 2016. p. 2016.
- Zumla AI, Gillespie SH, Hoelscher M, Philips PP, Cole ST, Abubakar I, et al. New antituberculosis drugs, regimens, and adjunct therapies: needs, advances, and future prospects. Lancet Infect Dis. 2014;14(4):327–40.
- Yew WW, Koh WJ. Emerging strategies for the treatment of pulmonary tuberculosis: promise and limitations? Korean J Intern Med. 2016;31(1):15–29.
- Menzies D, Benedetti A, Paydar A, Martin I, Royce S, Pai M, et al. Effect
 of duration and intermittency of rifampin on tuberculosis treatment outcomes: a systematic review and meta-analysis. PLoS Med.
 2009;6(9):e1000146.
- Ramachandran G, Swaminathan S. Role of pharmacogenomics in the treatment of tuberculosis: a review. Pharmgenomics Pers Med. 2012;5:89–98.
- Mooij MG, Nies AT, Knibbe CA, Schaeffeler E, Tibboel D, Schwab M, et al. Development of human membrane transporters: drug disposition and pharmacogenetics. Clin Pharmacokinet. 2016;55(5):507–24.
- Koo SH, Lo YL, Yee JY, Lee EJ. Genetic and/or non-genetic causes for interindividual and inter-cellular variability in transporter protein expression: implications for understanding drug efficacy and toxicity. Expert Opin Drug Metab Toxicol. 2015;11(12):1821–37.
- Ramos JSA, Felicio LP, Alves AA, Lopes MP, Soares TN, de Melo ESD. Unraveling CYP2E1 haplotypes in alcoholics from Central Brazil: a comparative study with 1000 genomes population. Environ Toxicol Pharmacol. 2018;62:30–9.
- Singer DR, Zair ZM. Clinical perspectives on targeting therapies for personalized medicine. Adv Protein Chem Struct Biol. 2016;102:79–114.
- Hodges LM, Markova SM, Chinn LW, Gow JM, Kroetz DL, Klein TE, et al. Very important pharmacogene summary: ABCB1 (MDR1, P-glycoprotein). Pharmacogenet Genomics. 2011;21(3):152–61.
- Sipeky C, Csongei V, Jaromi L, Safrany E, Maasz A, Takacs I, et al. Genetic variability and haplotype profile of MDR1 (ABCB1) in Roma and Hungarian population samples with a review of the literature. Drug Metab Pharmacokinet. 2011;26(2):206–15.
- Rodriguez-Castillo JA, Arce-Mendoza AY, Quintanilla-Siller A, Rendon A, Salinas-Carmona MC, Rosas-Taraco AG. Possible association of rare polymorphism in the ABCB1 gene with rifampin and ethambutol drugresistant tuberculosis. Tuberculosis (Edinb). 2015;95(5):532–7.
- Salvador-Martin S, Garcia-Gonzalez X, Garcia MI, Blanco C, Garcia-Alfonso P, Robles L, et al. Clinical utility of ABCB1 genotyping for preventing toxicity in treatment with irinotecan. Pharmacol Res. 2018;136:133–9.
- Pontual Y, Pacheco VSS, Monteiro SP, Quintana MSB, Costa MJM, Rolla VC, et al. ABCB1 gene polymorphism associated with clinical factors can predict drug-resistant tuberculosis. Clin Sci (Lond). 2017;131(15):1831–40.
- Pan Y, Chen W, Wang Y, Li H, Johnston SC, Simon T, et al. Association between ABCB1 polymorphisms and outcomes of clopidogrel treatment in patients with minor stroke or transient ischemic attack: secondary analysis of a randomized clinical trial. JAMA Neurol. 2019;76(5):552–60.

- Tan ZR, Zhou YX, Liu J, Huang WH, Chen Y, Wang YC, et al. The influence of ABCB1 polymorphism C3435T on the pharmacokinetics of silibinin. J Clin Pharm Ther. 2015;40(6):685–8.
- Sharif A, Kheirkhah D, Reza Sharif M, Karimian M, Karimian Z. ABCB1-C3435T polymorphism and breast cancer risk: a case-control study and a meta-analysis. J BUON. 2016;21(6):1433–41.
- Belmonte C, Ochoa D, Roman M, Saiz-Rodriguez M, Wojnicz A, Gomez-Sanchez CI, et al. Influence of CYP2D6, CYP3A4, CYP3A5 and ABCB1 polymorphisms on pharmacokinetics and safety of aripiprazole in healthy volunteers. Basic Clin Pharmacol Toxicol. 2018;122(6):596–605.
- Jiang B, Yan LJ, Wu Q. ABCB1 (C1236T) polymorphism affects P-glycoprotein-mediated transport of methotrexate, doxorubicin, actinomycin D, and etoposide. DNA Cell Biol. 2019;38(5):485–90.
- Hattori S, Suda A, Kishida I, Miyauchi M, Shiraishi Y, Fujibayashi M, et al. Effects of ABCB1 gene polymorphisms on autonomic nervous system activity during atypical antipsychotic treatment in schizophrenia. BMC Psychiatry. 2018;18(1):231.
- Rana SV, Sharma SK, Ola RP, Kamboj JK, Malik A, Morya RK, et al. N-acetyltransferase 2, cytochrome P4502E1 and glutathione S-transferase genotypes in antitubercular treatment-induced hepatotoxicity in North Indians. J Clin Pharm Ther. 2014;39(1):91–6.
- Yang S, Hwang SJ, Park JY, Chung EK, Lee JI. Association of genetic polymorphisms of CYP2E1, NAT2, GST and SLCO1B1 with the risk of anti-tuberculosis drug-induced liver injury: a systematic review and meta-analysis. BMJ Open. 2019;9(8):e027940.
- Perwitasari DA, Atthobari J, Wilffert B. Pharmacogenetics of isoniazidinduced hepatotoxicity. Drug Metab Rev. 2015;47(2):222–8.
- Hayashi S, Watanabe J, Kawajiri K. Genetic polymorphisms in the 5'-flanking region change transcriptional regulation of the human cytochrome P450IIE1 gene. J Biochem. 1991;110(4):559–65.
- 29. Tang K, Li Y, Zhang Z, Gu Y, Xiong Y, Feng G, et al. The Pstl/Rsal and Dral polymorphisms of CYP2E1 and head and neck cancer risk: a meta-analysis based on 21 case-control studies. BMC Cancer. 2010;10:575.
- 30. Roy PD, Majumder M, Roy B. Pharmacogenomics of anti-TB drugs-related hepatotoxicity. Pharmacogenomics. 2008;9(3):311–21.
- 31. Sheng YJ, Wu G, He HY, Chen W, Zou YS, Li Q, et al. The association between CYP2E1 polymorphisms and hepatotoxicity due to anti-tuber-culosis drugs: a meta-analysis. Infect Genet Evol. 2014;24:34–40.
- Singla N, Gupta D, Birbian N, Singh J. Association of NAT2, GST and CYP2E1 polymorphisms and anti-tuberculosis drug-induced hepatotoxicity. Tuberculosis (Edinb). 2014;94(3):293–8.
- Wu X, Shi H, Jiang H, Kemp B, Hong WK, Delclos GL, et al. Associations between cytochrome P4502E1 genotype, mutagen sensitivity, cigarette smoking and susceptibility to lung cancer. Carcinogenesis. 1997;18(5):967–73.
- 34. Pradipta IS, Forsman LD, Bruchfeld J, Hak E, Alffenaar JW. Risk factors of multidrug-resistant tuberculosis: a global systematic review and meta-analysis. J Infect. 2018;77(6):469–78.
- 35. Mohidem NA, Hashim Z, Osman M, Shaharudin R, Muharam FM, Makeswaran P. Demographic, socio-economic and behavior as risk factors of tuberculosis in Malaysia: a systematic review of the literature. Rev Environ Health. 2018;33(4):407–21.
- Soeroto AY, Lestari BW, Santoso P, Chaidir L, Andriyoko B, Alisjahbana B, et al. Evaluation of Xpert MTB-RIF guided diagnosis and treatment of rifampicin-resistant tuberculosis in Indonesia: a retrospective cohort study. PLoS ONE. 2019;14(2):e0213017.
- Pakasi TA, Karyadi E, Dolmans WM, van der Meer JW, van der Velden K. Malnutrition and socio-demographic factors associated with pulmonary tuberculosis in Timor and Rote Islands, Indonesia. Int J Tuberc Lung Dis. 2009;13(6):755–9.
- Verrall AJ, Alisjahbana B, Apriani L, Novianty N, Nurani AC, van Laarhoven A, et al. Early clearance of Mycobacterium tuberculosis: The INFECT case contact cohort study in Indonesia. J Infect Dis. 2020;221(8):1351–60.
- Ahmad RA, Mahendradhata Y, Utarini A, de Vlas SJ. Diagnostic delay amongst tuberculosis patients in Jogjakarta Province, Indonesia is related to the quality of services in DOTS facilities. Trop Med Int Health. 2011;16(4):412–23.
- 40. Adane K, Spigt M, Johanna L, Noortje D, Abera SF, Dinant GJ. Tuberculosis knowledge, attitudes, and practices among northern Ethiopian prisoners: implications for TB control efforts. PLoS ONE. 2017;12(3):e0174692.

Barliana et al. BMC Res Notes (2021) 14:295 Page 7 of 7

- 41. Mburu JW, Kingwara L, Ester M, Andrew N. Prognostic factors among TB and TB/DM comorbidity among patients on short course regimen within Nairobi and Kiambu counties in Kenya. J Clin Tuberc Other Mycobact Dis. 2018;12:9–13.
- 42. Snow K, Hesseling AC, Naidoo P, Graham SM, Denholm J, du Preez K. Tuberculosis in adolescents and young adults: epidemiology and treatment outcomes in the Western Cape. Int J Tuberc Lung Dis. 2017;21(6):651–7.
- 43. Soh AZ, Chee CBE, Wang YT, Yuan JM, Koh WP. Alcohol drinking and cigarette smoking in relation to risk of active tuberculosis: prospective cohort study. BMJ Open Respir Res. 2017;4(1):e000247.
- 44. Rao VG, Bhat J, Yadav R, Muniyandi M, Bhondeley MK, Sharada MA, et al. Tobacco smoking: a major risk factor for pulmonary tuberculosis—evidence from a cross-sectional study in central India. Trans R Soc Trop Med Hyg. 2014;108(8):474–81.
- Khabour OF, Alzoubi KH, Al-Azzam SI, Mhaidat NM. Frequency of MDR1 single nucleotide polymorphisms in a Jordanian population, including a novel variant. Genet Mol Res. 2013;12(1):801–8.

- Bossennec M, Di Roio A, Caux C, Menetrier-Caux C. MDR1 in immunity: friend or foe? Oncoimmunology. 2018;7(12):e1499388.
- 47. Goh LP, Chong ET, Chua KH, Chuah JA, Lee PC. Significant genotype difference in the CYP2E1 Pstl polymorphism of indigenous groups in Sabah, Malaysia with Asian and non-Asian populations. Asian Pac J Cancer Prev. 2014;15(17):7377–81.
- Ulusoy G, Arinc E, Adali O. Genotype and allele frequencies of polymorphic CYP2E1 in the Turkish population. Arch Toxicol. 2007;81(10):711–8.
- 49. Yue J, Peng RX, Yang J, Kong R, Liu J. CYP2E1 mediated isoniazid-induced hepatotoxicity in rats. Acta Pharmacol Sin. 2004;25(5):699–704.
- Xiang Y, Ma L, Wu W, Liu W, Li Y, Zhu X, et al. The incidence of liver injury in Uyghur patients treated for TB in Xinjiang Uyghur autonomous region, China, and its association with hepatic enzyme polymorphisms nat2, cyp2e1, gstm1 and gstt1. PLoS ONE. 2014;9(1):e85905.

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